



US012612634B2

(12) **United States Patent**
Kim et al.

(10) **Patent No.:** **US 12,612,634 B2**
(45) **Date of Patent:** ***Apr. 28, 2026**

(54) **COMPOSITIONS FOR INDUCING MODIFICATIONS OF TARGET ENDOGENOUS NUCLEIC ACID SEQUENCES IN NUCLEUSES OF EUKARYOTIC CELLS**

(71) Applicant: **TOOLGEN INCORPORATED**, Seoul (KR)

(72) Inventors: **Jin-Soo Kim**, Seoul (KR); **Seung Woo Cho**, Seoul (KR); **Sojung Kim**, Seoul (KR)

(73) Assignee: **TOOLGEN INCORPORATED**, Seoul (KR)

(*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 0 days.

This patent is subject to a terminal disclaimer.

(21) Appl. No.: **19/027,181**

(22) Filed: **Jan. 17, 2025**

(65) **Prior Publication Data**

US 2025/0171788 A1 May 29, 2025

Related U.S. Application Data

(63) Continuation of application No. 18/932,745, filed on Oct. 31, 2024, which is a continuation of application No. 17/004,338, filed on Aug. 27, 2020, which is a continuation of application No. 14/685,568, filed on Apr. 13, 2015, now Pat. No. 10,851,380, which is a continuation of application No. PCT/KR2013/009488, filed on Oct. 23, 2013.

(60) Provisional application No. 61/837,481, filed on Jun. 20, 2013, provisional application No. 61/803,599, filed on Mar. 20, 2013, provisional application No. 61/717,324, filed on Oct. 23, 2012.

(51) **Int. Cl.**

- C12N 9/22** (2006.01)
- C12N 9/16** (2006.01)
- C12N 15/10** (2006.01)
- C12N 15/11** (2006.01)
- C12N 15/52** (2006.01)
- C12N 15/63** (2006.01)
- C12N 15/82** (2006.01)
- C12N 15/85** (2006.01)
- C12N 15/90** (2006.01)

(52) **U.S. Cl.**

CPC **C12N 15/52** (2013.01); **C12N 9/16** (2013.01); **C12N 9/22** (2013.01); **C12N 15/102** (2013.01); **C12N 15/111** (2013.01); **C12N 15/63** (2013.01); **C12N 15/8216** (2013.01); **C12N 15/85** (2013.01); **C12N 15/907** (2013.01); **C12Y 301/21** (2013.01); **C12N 2310/10** (2013.01); **C12N 2310/20** (2017.05); **C12N 2310/531** (2013.01)

(58) **Field of Classification Search**

None
See application file for complete search history.

(56) **References Cited**

U.S. PATENT DOCUMENTS

- 5,766,900 A 6/1998 Shillito et al.
- 5,767,367 A 6/1998 Dudits et al.
- 5,925,517 A 7/1999 Tyagi et al.
- 6,040,295 A 3/2000 Rolland et al.
- 8,697,359 B1 4/2014 Zhang
- 8,771,495 B2 7/2014 Panecasio et al.
- 8,883,233 B2 11/2014 Gillessen
- 8,889,559 B2 11/2014 Trapp et al.
- 8,993,233 B2 3/2015 Zhang et al.
- 8,999,641 B2 4/2015 Zhang et al.
- 9,023,649 B2 5/2015 Mali et al.
- 9,260,723 B2 2/2016 Mali et al.
- 9,493,779 B2 11/2016 Ainley et al.
- 9,637,739 B2 5/2017 Siksnyis et al.

(Continued)

FOREIGN PATENT DOCUMENTS

- AU 733057 B2 5/2001
- AU 2011203213 A1 8/2011

(Continued)

OTHER PUBLICATIONS

Wefers et al., Direct production of mouse disease models by embryo microinjection of TALENs and oligodeoxynucleotides. PNAS (2013), 110: 3782-3787 (Year: 2013).*
Dadi et al., Decreased growth factor expression through RNA interference inhibits development of mouse preimplantation embryos. Comparative Medicine (2009), 59: 331-338 (Year: 2009).*
O'Meara et al., Gene silencing in bovine zygotes: siRNA transfection versus microinjection. Reproduction, Fertility and Development (2011), 23: 534-543 (Year: 2011).*
Bartlett and Davis, Effect of siRNA Nuclease Stability on the In Vitro and In Vivo Kinetics of siRNA-Mediated Gene Silencing. Biotechnology and Bioengineering (2007), 97: 909-92 (Year: 2007).*
Kim et al., Highly efficient RNA-guided genome editing in human cells via delivery of purified Cas9 ribonucleoproteins. Genome Research (2014), 24: 1012-1019 (Year: 2014).*

(Continued)

Primary Examiner — Catherine Konopka

(74) Attorney, Agent, or Firm — Gemini Law LLP

(57)

ABSTRACT

The present disclosure relates to targeted genome editing in eukaryotic cells or organisms. More particularly, the present disclosure provides for compositions and methods that may induce modifications in target endogenous nucleic acid sequences in nucleuses of eukaryotic cells. For example, methods disclosed herein may comprise preparing a Cas9/RNA complex, wherein the Cas9/RNA complex comprises a Cas9 protein and a guide RNA. Methods may further comprise introducing the Cas9/RNA complex into a non-human embryo, wherein the Cas9/RNA complex induces a modification at a target endogenous nucleic acid of the non-human embryo to provide for a genome modified embryo. Methods may further comprise transferring the genome modified embryo into a foster mother and allowing the foster mother to produce a F0 animal having the modification at the target endogenous nucleic acid.

16 Claims, 61 Drawing Sheets

Specification includes a Sequence Listing.

(56)

References Cited

U.S. PATENT DOCUMENTS

| | | | |
|--------------|-----|---------|-------------------------------------|
| 10,851,380 | B2 | 12/2020 | Kim et al. |
| 2005/0220796 | A1 | 10/2005 | Dynan et al. |
| 2009/0111119 | A1 | 4/2009 | Doyon et al. |
| 2010/0001189 | A1 | 1/2010 | Federici |
| 2010/0076057 | A1 | 3/2010 | Sontheimer et al. |
| 2011/0223638 | A1 | 9/2011 | Wiedenheft et al. |
| 2012/0058102 | A1 | 3/2012 | Wilson et al. |
| 2014/0068797 | A1* | 3/2014 | Doudna C12N 15/907 435/375 |
| 2014/0090113 | A1 | 3/2014 | Cogan et al. |
| 2014/0093913 | A1 | 4/2014 | Cost et al. |
| 2014/0179770 | A1 | 6/2014 | Zhang et al. |
| 2014/0186843 | A1 | 7/2014 | Zhang et al. |
| 2014/0186919 | A1 | 7/2014 | Zhang et al. |
| 2014/0189896 | A1 | 7/2014 | Zhang et al. |
| 2014/0242664 | A1 | 8/2014 | Zhang et al. |
| 2014/0273230 | A1 | 9/2014 | Chen et al. |
| 2014/0310830 | A1 | 10/2014 | Zhang et al. |
| 2014/0342456 | A1 | 11/2014 | Mali et al. |
| 2014/0349405 | A1 | 11/2014 | Sontheimer et al. |
| 2014/0357530 | A1 | 12/2014 | Zhang et al. |
| 2015/0020223 | A1 | 1/2015 | Zhang et al. |
| 2015/0031134 | A1 | 1/2015 | Zhang et al. |
| 2015/0050699 | A1 | 2/2015 | Siksnys et al. |
| 2015/0067921 | A1 | 3/2015 | Cogan et al. |
| 2015/0067922 | A1 | 3/2015 | Yang et al. |
| 2015/0079681 | A1 | 3/2015 | Zhang |
| 2015/0166615 | A1 | 6/2015 | Xia et al. |
| 2015/0203872 | A1 | 7/2015 | Zhang |
| 2015/0225734 | A1 | 8/2015 | Voytas et al. |
| 2015/0232882 | A1 | 8/2015 | Zhang et al. |
| 2015/0247150 | A1 | 9/2015 | Zhang et al. |
| 2015/0291965 | A1 | 10/2015 | Zhang et al. |
| 2015/0344912 | A1 | 12/2015 | Kim et al. |
| 2015/0351340 | A1 | 12/2015 | Bundock et al. |
| 2015/0356239 | A1 | 12/2015 | Zhang et al. |
| 2016/0017366 | A1 | 1/2016 | Chen et al. |
| 2016/0032274 | A1 | 2/2016 | Church et al. |
| 2016/0046961 | A1 | 2/2016 | Jinek et al. |
| 2016/0153004 | A1 | 6/2016 | Zhang et al. |
| 2016/0153005 | A1 | 6/2016 | Zhang et al. |
| 2016/0153006 | A1 | 6/2016 | Zhang et al. |
| 2016/0168594 | A1 | 6/2016 | Zhang et al. |
| 2016/0175462 | A1 | 6/2016 | Zhang et al. |
| 2016/0186213 | A1 | 6/2016 | Zhang et al. |
| 2021/0047648 | A1 | 2/2021 | Kim et al. |

FOREIGN PATENT DOCUMENTS

| | | | |
|----|--------------|----|---------|
| JP | 2005006578 | A | 1/2005 |
| NZ | 228948 | A | 6/1991 |
| WO | WO1998010084 | A1 | 3/1998 |
| WO | WO2000055378 | A1 | 9/2000 |
| WO | WO2002067966 | A1 | 9/2002 |
| WO | WO2002067996 | A2 | 9/2002 |
| WO | WO2005054494 | A2 | 6/2005 |
| WO | WO2007024029 | A1 | 3/2007 |
| WO | WO2007025195 | A1 | 3/2007 |
| WO | WO2008108989 | A2 | 9/2008 |
| WO | WO2009042164 | A1 | 4/2009 |
| WO | WO2010001189 | A1 | 1/2010 |
| WO | WO2010052341 | A1 | 5/2010 |
| WO | WO2010076939 | A1 | 7/2010 |
| WO | WO2011007193 | A1 | 1/2011 |
| WO | WO2011056186 | A1 | 5/2011 |
| WO | WO2011130346 | A1 | 10/2011 |
| WO | WO2011146121 | A1 | 11/2011 |
| WO | WO2012012738 | A1 | 1/2012 |
| WO | WO2012138939 | A1 | 10/2012 |
| WO | WO2013098244 | A1 | 7/2013 |
| WO | WO2013188522 | A2 | 12/2013 |
| WO | WO2014018423 | A2 | 1/2014 |
| WO | WO2014022702 | A2 | 2/2014 |
| WO | WO2014065596 | A1 | 5/2014 |

| | | | | | |
|----|---------------|------|---------|-------|------------|
| WO | WO-2014089290 | A1 * | 6/2014 | | A61K 38/00 |
| WO | WO2014093635 | A1 | 6/2014 | | |
| WO | WO2014093694 | A1 | 6/2014 | | |
| WO | WO2014093709 | A1 | 6/2014 | | |
| WO | WO2014093718 | A1 | 6/2014 | | |
| WO | WO2014099750 | A2 | 6/2014 | | |
| WO | WO2014144155 | A1 | 9/2014 | | |
| WO | WO2014197568 | A2 | 12/2014 | | |
| WO | WO2014204725 | A1 | 12/2014 | | |
| WO | WO2015026883 | A1 | 2/2015 | | |

OTHER PUBLICATIONS

Hudson and Ortlund, The structure, function and evolution of proteins that bind DNA and RNA. *Nature Reviews Molecular Cell Biology* (2014), 15: 749-760 (Year: 2014).*

Sternberg et al., Mechanism of substrate selection by a highly specific CRISPR endoribonuclease. *RNA* (2012), 18: 661-672 (Year: 2012).*

Jinek et al., A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity. *Science* (2012), 337: 816-821 (Year: 2012).*

Halfime of Spontaneous RNA Hydrolysis at 25° C., B10NUM3R5, <https://bionumbers.hms.harvard.edu/bionumber.aspx?id=105354&ver=3> (1 page) Referencing 2001 study.

Farboud B., et al. Enhanced Genome Editing with Cas9 Ribonucleoprotein in Diverse Cells and Organisms. *J. Visualized Experiments*. 2018, 135:e57370 (13 pages).

Hennig S. K., et al. Evaluation of Mutation Rates, Mosaicism and Off Target Mutations when Injecting Cas9 mRNA or Protein for Genome Editing of Bovine Embryos. *Scientific Reports*. (2020) 10:22309 (9 pages).

Lin, S., et al., Enhanced homology-directed human genome engineering by controlled timing of CRISPR/Cas9 delivery. *eLife*. 2014, 3:e04766 (with supplemental materials). (18 pages).

Mehravar M., et al. Mosaicism in CRISPR/Cas9-mediated Genome Editing. *Developmental Biology*. 445 (2019) 156-162 (7 pages).

Sung Y. H., et al. Highly Efficient Gene Knockout in Mice and Zebrafish with RNA-guided Endonucleases. *Genome Research*. (2014) 24(1):125-31 (8 pages).

Li, D., et al. Heritable gene targeting in the mouse and rat using a CRISPR-Cas system. *Nat Biotechnol*. 2013, 31 (8):681-683 and Supplementary Materials.

Li, J-F., et al., Multiplex and homologous recombination-mediated genome editing in *Arabidopsis* and *Nicotiana benthamiana* using guide RNA and Cas9. *Nat Biotechnol*. 2013, 31(8):688-691 and Supplementary Materials.

Li, T., et al., Modularly Assembled Designer TAL Effector Nucleases for Targeted Gene Knockout and Gene Replacement in Eukaryotes. *Nucleic Acids Research*, 2011, 39(14):6315-6325.

Li, T., et al., TAL nucleases (TALNs): hybrid protein composed of TAL effectors and FokI DNA-cleavage domain. *Nucleic Acids Research*. 2011, 39(1):359-372. (14 pages).

Li, W., et al. Simultaneous generation and germline transmission of multiple gene mutations in rat using CRISPR-Cas systems. *Nat Biotechnol*. 2013, 31(8):684-686 and Supplementary Materials.

Lieber, M. R., The Mechanism of Double-Strand DNA Break Repair by the Nonhomologous DNA End Joining Pathway. *Annual Review of Biochemistry*. 2010, 79:181-211. (34 pages).

Lino, C. A., et al., Delivering CRISPR: a review of the challenges and approaches. *Drug Delivery*. 2018, 25(1):1234-1257.

Liu, J., et al., Efficient and Specific Modifications of the *Drosophila* Genome by Means of an Easy TALEN Strategy. *Journal of Genetics and Genomics*. 2012, 39(5):209-215. (7 pages).

Liu, P-Q., et al., Generation of a triple-gene knockout mammalian cell line using engineered zinc-finger nucleases. *Biotechnology and Bioengineering*. 2010, 106(1):97-105.

Liu, Y., et al., Polyethylene glycol (PEG)-mediated transformation of the fused egfp-hph gene into *Pleurotus ostreatus*. *African Journal of Biotechnology*. 2012, 11(19):4345-4353. (9 pages).

Liu, Y-C., et al., Efficient Polyethylene Glycol (PEG) Mediated Transformation of the Moss *Physcomitrella patens*. *J. Vis. Exp*. 2011, 50:2560. (4 pages).

(56)

References Cited

OTHER PUBLICATIONS

- Lo, T-W., et al., Precise and Heritable Genome Editing in Evolutionarily Diverse Nematodes Using TALENs and CRISPR/Cas9 to Engineer Insertions and Deletions. *Genetics*. 2013, 195(2):331-348. (26 pages).
- Lu, C. and Li, P., Preparation of Short RNA by In Vitro Transcription. In: Conn, G. (eds) *Recombinant and In Vitro RNA Synthesis. Methods in Molecular Biology*, 2013, vol. 941. Humana Totowa, NJ. (12 pages).
- Ma, S. et al., Highly efficient and specific genome editing in silkworm using custom TALENs. *PLoS ONE*. 2012, 7(9): e45035. (7 pages).
- Madrigal, P., and Pawel, K., Current bioinformatic approaches to identify DNase I hypersensitive sites and genomic footprints from DNase-seq data. *Frontiers in Genetics*. 2012, 3(230):1-3. (3 pages).
- Maeder, M. L. et al., CRISPR RNA-guided activation of endogenous human genes. *Nat. Methods*. 2013, 10:977-979.
- Magnani, L., et al., Pioneer factors: directing transcriptional regulators within the chromatin environment. *Trends in Genetics*. 2011, 27(11):465-474. (10 pages).
- Mahfouz, M. M., et al., De novo-engineered transcription activator-like effector (TALE) hybrid nuclease with novel DNA binding specificity creates double-strand breaks. *PNAS*. 2011, 108(6):2623-2628.
- Mak, A. N-S., et al., The Crystal Structure of TAL Effector PthXo1 Bound to Its DNA Target. *Science*. 2012, 335 (6069):716-719. (5 pages).
- Makarova, K. S., et al., Evolution and classification of the CRISPR-Cas systems. *Nat Rev Microbiol*. 2011, 9:467-477. (11 pages).
- Mali, P., et al., Cas9 as a Versatile Tool for Engineering Biology. *Nat Methods*. 2013, 10(10):957-963.
- Mali, P., et al., RNA-Guided Human Genome Engineering via Cas9. *Science*. 2013, 339(6121):823-826 and Supplementary Materials. (40 pages).
- Mali, P., et al., CAS9 transcriptional activators for target specificity screening and paired nickases for cooperative genome engineering. *Nat Biotechnol*. 2013, 31:833-838.
- Marchfelder, A., et al., Small RNAs for defence and regulation in archaea. *Extremophiles*. 2012, 16:685-696. (12 pages).
- Maresca, M., et al., Obligate ligation-gated recombination (ObliGaRe): Custom-designed nuclease-mediated targeted integration through nonhomologous end joining. *Genome Res*. 2013, 23:539-546.
- Marfori, M., et al., Molecular basis for specificity of nuclear import and prediction of nuclear localization. *Biochimica et Biophysica Acta (BBA)—Molecular Cell Research*. 2011, 1813(9):1562-1577. (16 pages).
- Marraffini, L. A. and Sonthheimer, E. J., Self versus non-self discrimination during CRISPR RNA-directed immunity. *Nature*. 2010, 463(7280):568-571. (13 pages).
- Mashimo, T., et al., Generation of Knockout Rats with X-Linked Severe Combined Immunodeficiency (X-SCID) Using Zinc-Finger Nucleases. *PLoS ONE* 2010, 5(1):e8870. (7 pages).
- Mazzara, G. P., et al., Maturation events leading to transfer RNA and ribosomal RNA. *Cell Biology*. 2012, 3:439-545.
- Miao, J., et al., Targeted mutagenesis in rice using CRISPR-Cas system. *Cell Res*. 2013, 23(10):1233-1236.
- Mika, J. T., and Poolman, B., Macromolecule diffusion and confinement in prokaryotic cells. *Current Opinion in Biotechnology*. 2011, 22(1):117-126. (10 pages).
- Miller, J. C., et al., A Tale nuclease architecture for efficient genome editing. *Nat Biotechnol*. 2011, 29(2):143-148 and online methods.
- Mougiakos, I., et al., Characterizing a thermostable Cas9 for bacterial genome editing and silencing. *Nat Commun*. 2017, 8:1647. (11 pages).
- Mussolino, C., et al., A novel TALE nuclease scaffold enables high genome editing activity in combination with low toxicity. *Nucleic Acids Research*. 2011, 39(21):9283-9293.
- Mussolino, C., and Cathomen, T., RNA guides genome engineering. *Nat. Biotechnol*. 2013, 31:208-209.
- Mussolino, C., and Cathomen, T., Tale nucleases: tailored genome engineering made easy. *Current Opinion in Biotechnology*. 2012, 23(5):644-650. (7 pages).
- Musunuru, K., Genome editing of human pluripotent stem cells to generate human cellular disease models. *Dis Model Mech*. 2013, 6(4):896-904. (17 pages).
- Nakayama, T., et al., Simple and efficient CRISPR/Cas9-mediated targeted mutagenesis in *Xenopus tropicalis*. *Genesis*. 2013, 51(12):835-843. (15 pages).
- Nature Biotechnology Journal webpage, vol. 30 issue 9, Sep. 10, 2012, available at <https://www.nature.com/nbt/volumes/30/issues/9> (last accessed on Mar. 22, 2021), (15 pages).
- NCBI record for NC-002737.2, 2019, “*Streptococcus pyogenes* M1 Gas, complete sequence”, retrieved from the internet at < https://www.ncbi.nlm.nih.gov/nucleotide/NC_002737.2 > (1 page).
- Nekrasov, V., et al., Targeted mutagenesis in the model plant *Nicotiana benthamiana* using Cas9 RNA-guided endonuclease. *Nat Biotechnol*. 2013;31(8):691-693 and Supplemental Materials.
- Ng, Y. S., et al., Chemical transfection of dye-conjugated microRNA precursors for microRNA functional analysis of M2 macrophages. *Journal of Cellular Biochemistry*. 2012, 113(5):1714-1723. (10 pages).
- Niinaka, Y., et al., Silencing of autocrine motility factor induces mesenchymal-to-epithelial transition and suppression of osteosarcoma pulmonary metastasis. *Cancer Res*. 2010, 70(22):9483-9493. (11 pages).
- Noland, C. L., et al., siRNA Repositioning for Guide Strand Selection by Human Dicer Complexes. *Molecular Cell*. 2011, 43(1):110-121. (12 pages).
- Ogawa, A., Rational design of artificial riboswitches based on ligand-dependent modulation of internal ribosome entry in wheat germ extract and their applications as label-free biosensors. *RNA*. 2011, 17:478-488.
- Pandika, M., 2014, “Jennifer Doudna, CRISPR Code Killer”, www.ozy.com/rising-stars-and-provocateurs/jennifer-doudna-crispr-code-killer/4690, Jan. 7, 2014, pp. 1-6.
- Pardo, R., et al., pARIS-htt: an optimised expression platform to study huntingtin reveals functional domains required for vesicular trafficking. *Mol Brain*. 2010, 3:17. (17 pages).
- Pattanayak, V., et al., Revealing off-target cleavage specificities of zinc finger nucleases by in vitro selection. *Nat Methods*, 2011, 8(9):765-770.
- Pattanayak, V., et al., High-throughput profiling of off-target DNA cleavage reveals RNA-programmed Cas9 nuclease specificity. *Nat. Biotechnol*. 2013, 31:839-843.
- Pennisi, E., Encode Project Writes Eulogy for Junk DNA. *Science*. 2012, 337(6099):1159-1161. (2 pages).
- Cristea, S., et al., Dissection of Splicing Regulation at an Endogenous Locus by Zinc-Finger Nuclease-Mediated Gene Editing. *PLoS One*. 2011, 6(2):e16961. (5 pages).
- Davis, G. D., and Cui, X., Zinc Finger Nucleases for Genome Editing. *Genetic Engineering & Biotechnology News*. 2010, 30(13). (6 pages).
- De Vries, R., DNA condensation in bacteria: Interplay between macromolecular crowding and nucleoid proteins. *Biochimie*. 2010, 92(12):1715-1721. (7 pages).
- Declaration of Technical Expert Paul Simons, executed Dec. 22, 2015, in relation to U.S. Appl. No. 14/704,551 (77 pages).
- Deltcheva, E., et al., CRISPR RNA maturation by trans-encoded small RNA and host factor RNase III. *Nature*. 2011, 471(7340):602-607 and Supplementary Information (71 pages).
- Deng, D., et al., Structural basis for sequence-specific recognition of DNA by TAL effectors. *Science*. 2012, 335 (6069):720-723.
- Dicarlo, J. E., et al., Genome engineering in *Saccharomyces cerevisiae* using CRISPR-Cas systems. *Nucleic Acids Research*. 2013, 41(7):4336-4343.
- Dickinson, D. J., et al., Engineering the *Caenorhabditis elegans* genome using Cas9-triggered homologous recombination. *Nat. Methods*. 2013, 10:1028-1034.
- Ding, Q., et al., Enhanced efficiency of human pluripotent stem cell genome editing through replacing TALENs with CRISPRs. *Cell Stem Cell*. 2013, 12(4):393-394. (2 pages).

(56)

References Cited

OTHER PUBLICATIONS

- Doyon, J. B., et al., Rapid and efficient clathrin-mediated endocytosis revealed in genome-edited mammalian cells. *Nature Cell Biology*. 2011, 13(3):331-337. (31 pages).
- Doyon, Y., et al., Enhancing zinc-finger-nuclease activity with improved obligate heterodimeric architectures. *Nature Methods*. 2011, 8:74-79. (38 pages).
- Ebina, H., et al., Harnessing the CRISPR/Cas9 system to disrupt latent HIV-1 provirus. *Sci Rep*. 2013, 3:2510. (7 pages).
- Edgar, R., and Qimron, U., The *Escherichia coli* CRISPR system protects from λ lysogenization, lysogens, and prophage induction. *J Bacteriol*. 2010, 192(23):6291-6294.
- Esvelt, K. M. et al., Orthogonal Cas9 proteins for RNA-guided gene regulation and editing. *Nat. Methods*. 2013, 10:1116-1121.
- Fath, S., et al., Multiparameter RNA and Codon Optimization: A Standardized Tool to Assess and Enhance Autologous Mammalian Gene Expression. *PLoS ONE*. 2011, 6(3):e17596. (14 pages).
- Feng, Z., et al., Efficient genome editing in plants using a CRISPR/Cas system. *Cell Res*. 2013, 23(10):1229-1232.
- Fonfara, I., et al., Phylogeny of Cas9 determines functional exchangeability of dual-RNA and Cas9 among orthologous type II CRISPR-Cas systems. *Nucleic Acids Research*. 2014, 42(4):2577-2590. (14 pages).
- Fortier, S., et al., Genome-Wide Interrogation of Mammalian Stem Cell Fate Determinants by Nested Chromosome Deletions. *PLoS Genetics*. 2010, 6(12):e1001241. (14 pages).
- Francis, D. M. and Page, R., Strategies to Optimize Protein Expression in *E. coli*. *Current Protocols in Protein Science*. 2010, 61:5.24.1-5.24.29. (29 pages).
- Friedland, A. E., et al., Heritable genome editing in *C. elegans* via a CRISPR-Cas 9 system. *Nat. Methods*. 2013, 10(8):741-743.
- Fu, Y., et al., High-frequency off-target mutagenesis induced by CRISPR-Cas nucleases in human cells. *Nat Biotechnol*. 2013, 31(9):822-826.
- Fujii, W., et al., Efficient generation of large-scale genome-modified mice using gRNA and CAS9 endonuclease. *Nucleic Acids Research*. 2013, 41(20):e187.
- Gabriel, R., et al., An unbiased genome-wide analysis of zinc-finger nuclease specificity. *Nat Biotechnol*. 2011, 29 (9):816-823.
- Gagnon, J. A., et al., Efficient Mutagenesis by Cas9 Protein-Mediated Oligonucleotide Insertion and Large-Scale Assessment of Single-Guide RNAs. *PLoS ONE*. 2014, 9(5):e98186. Supplementary Information (25 pages).
- Gaj, T., et al., ZFN, TALEN, and CRISPR/Cas-based methods for genome engineering. *Trends in Biotechnology*. 2013, 31(7):397-405. (9 pages).
- Gao, H., et al., Heritable targeting mutagenesis in maize using a designed endonuclease. *The Plant Journal*. 2010, 61 (1):176-187. (13 pages).
- Garneau, J. E., et al., The CRISPR/Cas bacterial immune system cleaves bacteriophage and plasmid DNA. *Nature*. 2010, 468:67-71.
- Gasiunas, G., et al., Cas9-crRNA ribonucleoprotein complex mediates specific DNA cleavage for adaptive immunity in bacteria. *PNAS*. 2012, 109(39):E2579-E2586.
- Genomic Cruise Missiles, *Science*, 2012;338:1526-1527. (12 pages).
- Gilbert, L. A. et al., CRISPR-Mediated Modular RNA-Guided Regulation of Transcription in Eukaryotes. *Cell*. 2013, 154(2):442-451. (10 pages).
- Golic, K. G., RNA-Guided Nucleases: A New Era for Engineering the Genomes of Model and Nonmodel Organisms. *Genetics*. 2013, 195(2):303-308. (6 pages).
- Gonzalez, B., et al, Modular system for the construction of zinc-finger libraries and proteins. *Nat Protoc*. 2010, 5 (4):791-810.
- Goren, M., et al., The bacterial CRISPR/Cas system as analog of the mammalian adaptive immune system. *RNA Biology*. 2012, 9(5):549-554.
- Gottwein, E., et al., Viral microRNA targetome of KSHV-infected primary effusion lymphoma cell lines. *Cell Host & Microbe*. 2011, 10(5):515-526.
- Gratz, S. J., et al., Genome engineering of drosophila with the CRISPR RNA-guided Cas9 nuclease. *Genetics*, 2013, 194(4):1029-1035 and supplementary materials. (17 pages).
- Groisman, E. A., et al., Bacterial Mg²⁺ Homeostasis, Transport, and Virulence. *Annual Reviews of Genetics*. 2013, 47:625-646. (22 pages).
- Guschin, D. Y., et al., A Rapid and General Assay for Monitoring Endogenous Gene Modification. In: Mackay, J., Segal, D. (eds.) *Engineered Zinc Finger Proteins. Methods in Molecular Biology*. 2010, vol. 649. Humana Press, Totowa, NJ.
- Angel, M. and Yanik, M. F., Innate Immune Suppression Enables Frequent Transfection with RNA Encoding Reprogramming Proteins. *PLoS ONE*. 2010, 5(7):e11756. (7 pages).
- Anonymous, Product Information Thermo Scientific TurboFect Transfection Reagent Pub. No. MAN0013147 Rev. Date Aug. 24, 2018. Thermo Scientific, (2018) (2 pages).
- Anonymous, PCT Third Party Observation dated Feb. 19, 2015, in relation to International Application No. PCT/KR2013/009488 filed on Oct. 23, 2013, pp. 1-5.
- Anonymous, Third-Party Observation dated Jul. 18, 2014, in relation to International Application No. PCT/US2013/033106 filed Mar. 20, 2013 (7 pages).
- Anonymous, Third-Party Observation dated Sep. 24, 2014, in relation to International Application No. PCT/US2013/032589 filed Mar. 15, 2013 (8 pages).
- Anonymous, Third-Party Observations dated Jan. 17, 2019, in relation to European Patent Application No. 18158147.1 (Publication No. EP 3372679 A1) (302 pages).
- Anonymous, Third-Party Submissions dated Dec. 22, 2014, in relation to U.S. Appl. No. 14/104,977, filed Dec. 12, 2013, pp. 1-56.
- Anonymous, Third-Party Submission Under 37 C.F.R. §1.290 filed Apr. 6, 2016 in relation to U.S. Appl. No. 14/438,098.
- Artimo, P., et al., ExPASy: SIB bioinformatics resource portal. *Nucleic Acids Research*. 2012, 40:W597-W603. (7 pages).
- ATCC record ATCC 700294, 2019, "*Streptococcus pyogenes* Rosenbach (ATCC 700294)", retrieved from the internet on Apr. 18, 2019 at http://www.lgcstandards-atcc.org/products/all/700294.aspx?geo_country=GB#history (2 pages).
- Barrangou, R., RNA-mediated programmable DNA cleavage. *Nat Biotechnol*. 2012, 30(9):836-838. (3 pages).
- Bassett, A. R., et al., Highly Efficient Targeted Mutagenesis of *Drosophila* with the CRISPR/Cas9 System. *Cell Reports*. 2013, 4(1):220-228 and Supplementary Table.
- Baum, A., et al., Preference of RIG-I for short viral RNA molecules in infected cells revealed by next-generation sequencing. *PNAS*. 2010, 107(37):16303-16308.
- Bedell, V. M., et al., In vivo genome editing using a high-efficiency TALEN system. *Nature*. 2012, 491:114-118. (7 pages).
- Bell, O., et al., Determinants and dynamics of genome accessibility. *Nature Reviews Genetics*. 2011, 12:554-564. (11 pages).
- Bhaya, D., et al., CRISPR-Cas systems in bacteria and archaea: versatile small RNAs for adaptive defense and regulation. *Annual Reviews of Genetics*. 2011;45:273-297. (27 pages).
- Bikard, D., et al., CRISPR Interference Can Prevent Natural Transformation and Virulence Acquisition during In Vivo Bacterial Infection. *Cell Host & Microbe*. 2012, 12(2):177-186.
- Bikard, D., et al., Programmable repression and activation of bacterial gene expression using an engineered CRISPR-Cas system. *Nucleic Acids Research*. 2013, 41(15):7429-7437.
- Boch, J. and Bonas, U., *Xanthomonas AvrBs3* Family-Type III Effectors: Discovery and Function. *Annual Review of Phytopathology*. 2010, 48:419-436.
- Bogerd, H. P., et al., A Mammalian Herpesvirus uses Noncanonical Expression and Processing Mechanisms to Generate Viral MicroRNAs. *Molecular Cell*. 2010, 37(1):135-142.
- Briggs, A.W., et al., Iterative capped assembly: rapid and scalable synthesis of repeat-module DNA such as TAL effectors from individual monomers. *Nucleic Acids Research*. 2012, 40(15):e117. (10 pages).
- Brouns, S. J. J., A Swiss Army Knife of Immunity. *Science*. 2012, 337(6096):808-809.
- Brzostek-Racine, S., et al., The DNA Damage Response Induces IFN. *The Journal of Immunology*. 2011, 187 (10):5336-5345.

(56)

References Cited

OTHER PUBLICATIONS

- Cain, C., CRISPR genome editing. *SciBX Sci. Exch.* 1-3, 2013, doi:10.1038/scibx.2013.77.
- Carlson, D. F., et al., Targeting DNA with fingers and TALENs. *Molecular Therapy—Nucleic Acids.* 2012, 1:e3. (4 pages).
- Carlson, E. D., et al., Cell-free protein synthesis: Applications come of age. *Biotechnology Advances.* 2012, 30 (5):1185-1194.
- Carroll, D., A CRISPR Approach to Gene Targeting. *Molecular Therapy.* 2012, 20(9):1658-1660. (3 pages).
- Carroll, D., Genome Engineering with Zinc-Finger Nucleases. *Genetics.* 2011, 188(4):773-782.
- Carroll, D., Staying on target with CRISPR-Cas. *Nat. Biotechnol.* 2013, 31:807-809.
- Cermak, T., et al., Efficient design and assembly of custom TALEN and other TAL effector-based constructs for DNA targeting. *Nucleic Acids Research.* 2011, 39(12):e82.
- Chang, K., et al., RNAi in Cultured Mammalian Cells Using Synthetic siRNAs. *Cold Spring Harb. Protoc.* 2012, 9:957-961. (9 pages).
- Chang, N., et al., Genome editing with RNA-guided Cas9 nuclease in Zebrafish embryos. *Cell Res.* 2013, 23 (4):465-472 and Supplementary Materials. (8 pages).
- Chapdelaine, P., et al., Meganucleases can restore the reading frame of a mutated dystrophin. *Gene Ther.* 2010, 17:846-858. (13 pages).
- Chen, B. et al., Dynamic Imaging of Genomic Loci in Living Human Cells by an Optimized CRISPR/Cas system. *Cell.* 2013, 155(7):1479-1491.
- Chen, F., et al., High-frequency genome editing using ssDNA oligonucleotides with zinc-finger nucleases. *Nature Methods.* 2011, 8(9):753-755 with Supplementary Information. (31 pages).
- Chen, F., et al., 2012, Methods and Reagents for Modifying Genomes Using RNA-Guided Endonucleases. in relation to U.S. Appl. No. 61/734,256.
- Cho, S. W., et al., Targeted genome engineering in human cells with Cas9 RNA-guided endonucleases. *Nat Biotechnol.* 2013, 31:230-232, and supplementary materials.
- Cho, S. W., et al., Heritable gene knockout in *Caenorhabditis elegans* by direct injection of Cas9-sgRNA ribonucleoproteins. *Genetics.* 2013, 195(3):1177-1180. (5 pages).
- Christian, M., et al., Targeting DNA Double-Strand Breaks with TAL Effector Nucleases. *Genetics.* 2010, 186 (2):757-761 and 2S1-8S1.
- Chugh, A., et al., Cell-penetrating peptides: Nanocarrier for Macromolecule Delivery in Living Cells. *IUBMB Life.* 2010, 62(3):183-193. (11 pages).
- Chylinski, K., et al., The tracrRNA and Cas9 families of type II CRISPR-Cas immunity systems. *RNA Biology.* 2013, 10(5):726-737.
- Clark, K. J., et al., A TALE of Two Nucleases: Gene Targeting for the Masses. *Zebrafish.* 2011, 8(3):147-149.
- Close, D., et al., Expression of Non-Native Genes in a Surrogate Host Organism. 2012. Retrieved from the Internet at <<http://www.intechopen.com>> (33 pages).
- Close, D., et al., The evolution of the bacterial luciferase gene cassette (*lux*) as a real-time bioreporter. *Sensors.* 2012;12(1):732-752. (21 pages).
- Cohen, J., et al., The Emerging Race to Cure HIV Infections. *Science.* 2011, 332(6031):784-789.
- Collins, C. A., and Brown, E. J., Cytosol as battleground: ubiquitin as a weapon for both host and pathogen. *Trends in Cell Biology.* 2010, 20(4):205-213.
- Cong, L., et al., Multiplex Genome Engineering Using CRISPR/Cas Systems. *Science.* 2013, 339(6121):819-823 and Supplementary Materials.
- CNLS Mapper results of *S. pyogenes* Cas9. NLS mapper, URL: http://nls-mapper.iab.keio.ac.jp/cgi-bin/NLS Mapper_ycgi (Apr. 25, 2016) (1 page).
- Cradick, T. J., et al., CRISPR/Cas 9 systems targeting β -globin and CCR5 genes have substantial off-target activity. *Nucleic Acids Research.* 2013, 41(20):9584-9592.
- Pennisi, E., The CRISPR Craze. *Science.* 2013, 341(6148):833-836.
- Perez-Pinera, P., et al., RNA-guided gene activation by CRISPR-Cas9-based transcription factors. *Nat Methods.* 2013, 10(10):973-976.
- Perez-Pinera, P., et al., Advances in Targeted Genome Editing. *Current Opinion in Chemical Biology.* 2012, 16 (3-4):268-277. (1 page).
- Perez-Rodriguez, R., Elucidating a novel mechanism of DNA silencing caused by envelope stress in *Escherichia coli*. Dissertation and Abstract first available to the public on Jun. 17, 2010 (156 pages).
- Pillich, H., et al., Activation of the unfolded protein response by *Listeria monocytogenes*. *Cellular Microbiology.* 2012, 14(6):949-964.
- Primo, M. N., et al., Lentiviral vectors for cutaneous RNA managing. *Experimental Dermatology.* 2012, 21(3):162-170.
- Program and Conference Logistics provided to the attendees of the CRISPR 2012: 5th Annual CRISPR Research Meeting held at the University of California, Berkeley, CA (Jun. 2012). (5 pages).
- PSHooter™ Vector user guide, Invitrogen by Life Technologies, revision date Mar. 29, 2012 (36 pages).
- Qi, L., et al., RNA Processing Enables Predictable Programming of Gene Expression. *Nat Biotechnol.* 2012, 30 (10):1002-1006 and Supplementary Materials.
- Qi, L. S., et al., Repurposing CRISPR as an RNA-Guided Platform for Sequence-Specific Control of Gene Expression. *Cell.* 2013, 152(5):1173-1183.
- Ramirez, C. L., et al., Engineered zinc finger nickases induce homology-directed repair with reduced mutagenic effects. *Nucleic Acid Research.* 2012, 40(12):5560-5568. (9 pages).
- Ran, F. A., et al., Double nicking by RNA-guided CRISPR cas9 for enhanced genome editing specificity. *Cell.* 2013, 154(6):1380-1389.
- Ran, F. A., et al., Genome engineering using the CRISPR-Cas9 system. *Nat. Protoc.* 2013, 8:2281-2308.
- Reyon, D., et al., FLASH assembly of TALENs for high-throughput genome editing. *Nat Biotechnol.* 2012, 30 (5):460-465. (8 pages).
- Richter, H., et al., Exploiting CRISPR/Cas: Interference Mechanisms and Applications. *International Journal of Molecular Sciences.* 2013, 14(7):14518-14531. (14 pages).
- Rios, X., et al., Stable Gene Targeting in Human Cells Using Single-Strand Oligonucleotides with Modified Bases. *PLoS ONE.* 2012, 7(5):e36697. (14 pages).
- Sakurai, K., et al., Silencing of Gene Expression in Cultured Cells Using Small Interfering RNAs. *Curr. Protoc. Cell Biol.* 2010;47:27.1.1-27.1.28. (28 pages).
- Sander, J. D., et al., Targeted gene disruption in somatic zebrafish cells using engineered TALENs. *Nature Biotechnology.* 2011, 29(8):697-698 with Supplementary Information. (27 pages).
- Sanders, R., 2013, "Cheap and easy technique to snip DNA could revolutionize gene therapy," UC Berkeley News Center, Jan. 7, 2013, retrieved from the internet at <<http://newscenter.berkeley.edu/2013/01/07/cheap-and-easy-technique-to-snip-dna-could-revolutionize-gene-therapy/>> pp. 1-3 (3 pages).
- Sanjana, N. E., et al., A Transcription Activator-Like Effector (TALE) Toolbox for Genome Engineering. *Nat Protoc.* 2012, 7(1):171-192. Supplementary Materials (28 pages).
- Sapranaukas, R., et al., The *Streptococcus thermophilus* CRISPR/Cas system provides immunity in *Escherichia coli*. *Nucleic Acids Research.* 2011, 39(21):9275-9282 and Supplementary Materials. (15 pages).
- Schmidt, A., et al., Sensing of viral nucleic acids by RIG-I: from translocation to translation. *European Journal of Cell Biology.* 2012, 91(1):78-85.
- Second Declaration of Dr. Boch dated Apr. 26, 2019 as filed in Opposition to European Patent No. 2825654 (20 pages).
- Segal, D. J., Genome Engineering: Bacteria herald a new era of gene editing. *eLife.* 2013, 2:e00563.
- Shalem, O., et al., Genome-Scale CRISPR-Cas9 Knockout Screening in Human Cells. *Science.* 2013, 343 (6166):84-87.
- Shan, Q., et al., Targeted genome modification of crop plants using a CRISPR-Cas system. *Nat Biotechnol.* 2013, 31 (8):686-688 and Supplementary Materials.

(56)

References Cited

OTHER PUBLICATIONS

- Shen, B., et al., Generation of gene-modified mice via Cas9/RNA-mediated gene targeting. *Cell Res.* 2013, 23:720-723.
- Singapore Written Opinion dated Mar. 18, 2016 in relation to Singapore Patent Application No. 11201503059X.
- Sinkunas, T., et al., Cas3 is a single-stranded DNA nuclease and ATP-dependent helicase in the CRISPR/Cas immune system. *The EMBO Journal.* 2011, 30:1335-1342. (8 pages).
- Strecker, J., et al., Engineering of CRISPR-Cas12b for human genome editing. *Nat Commun.* 2019, 10(1):212.
- Sternberg, S. H., et al., DNA interrogation by the CRISPR RNA-guided endonuclease Cas9. *Biophysical Journal.* 2014, 106(2):62-67. (17 pages).
- Sternberg, S. H., et al., Mechanism of substrate selection by a highly specific CRISPR endonuclease. *RNA* 2012, 18:661-672. (12 pages).
- Streptococcus*—bacterial genus—Microbiology Dictionary, Hardy Diagnostics, available at https://catalog.hardydiagnostics.com/cp_prod/Content/hugo/Streptococcus.htm (last accessed Oct. 25, 2020), (4 pages).
- Sun, C., et al., Functional reconstruction of human eukaryotic translation initiation factor 3 (eIF3). *PNAS.* 2011, 108 (51):20473-20478. (6 pages).
- Sung, Y. H., et al., Mouse genetics: Catalogue and scissors. *BMB Rep.* 2012, 45(12):686-692.
- Sung, Y. H., et al., Highly efficient gene knockout in mice and zebrafish with RNA-guided endonucleases. *Genome Res.* 2014, 24:125-131. (7 pages).
- Svitashev, S., et al., Genome editing in maize directed by CRISPR-Cas9 ribonucleoprotein complexes. *Nature Communications.* 2016, 7:13274. (7 pages).
- Szczepankowska, A., Role of CRISPR/cas system in the development of bacteriophage resistance. *Advances in Virus Research.* 2012, 82:289-338. (50 pages).
- Terns, M. P. and Terns, R. M., CRISPR-Based Adaptive Immune Systems. *Current Opinion in Microbiology.* 2011, 14(3):321-327. (13 pages).
- Tesson, L., et al., Knockout rats generated by embryo microinjection of TALENs. *Nat Biotechnol.* 2011, 29 (8):695-696 and Supplementary Materials. (15 pages).
- Thurman, R. E., et al., The accessible chromatin landscape of the human genome. *Nature.* 2012, 489:75-82. (8 pages).
- Tzur, Y. B., et al., Heritable Custom Genomic Modifications in *Caenorhabditis elegans* via a CRISPR-Cas9 System. *Genetics.* 2013, 195(3):1181-1185. (14 pages).
- Urnov, F. D., et al., Genome editing with engineered zinc finger nucleases. *Nat Rev Genet.* 2010, 11(9):636-646.
- Van Der Oost, J., New Tool for Genome Surgery. *Science.* 2013, 339(6121):768-770.
- Vasu, K., et al., Endonuclease Active Site Plasticity Allows DNA Cleavage with Diverse Alkaline Earth and Transition Metal Ions. *ACS Chemical Biology.* 2011, 6(9):934-942. (9 pages).
- Villion, M., and Moineau, S., The double-edged sword of CRISPR-Cas9 systems. *Cell Res.* 2013, 23:15-17. (4 pages).
- Walsh, R. M., and Hochedlinger, K., A variant CRISPR-Cas9 system adds versatility to genome engineering. *PNAS.* 2013, 110(39):15514-15515.
- Wang, H., et al., One-step generation of mice carrying mutations in multiple genes by CRISPR/Cas-mediated genome engineering. *Cell.* 2013, 153(4):910-918. (17 pages).
- Wang, J., et al., Targeted gene addition to a predetermined site in the human genome using a ZFN-based nicking enzyme. *Genome Res.* 2012, 22(7):1316-1326. (11 pages).
- Wang, T., et al., Genetic Screens in Human Cells Using the CRISPR/Cas9 System. *Science.* 2013, 343(6166):80-84.
- Wee, L., et al., Argonaute divides its RNA Guide into domains with distinct functions and RNA-binding properties. *Cell.* 2012, 151(5):1055-1067. (23 pages).
- Weeks, A., et al., Structural and Biochemical Studies of a Fluoroacetyl-CoA-Specific Thioesterase Reveal a Molecular Basis for Fluorine Selectivity. *Biochemistry.* 2010, 49(43):9269-9279 and Supplementary Materials. (21 pages).
- Welch, M., et al., Chapter 3: Designing Genes for Successful Protein Expression. Eds: Voigt, C., *Methods in Enzymology.* Academic Press. 2011, 498:43-66. (24 pages).
- Wente, S. R. and Rout, M. P., The Nuclear Pore Complex and Nuclear Transport. *Cold Spring Harb Perspect Biol.* 2010, 2:a000562. (21 pages).
- Whisnant, A. W., et al., In depth analysis of the interaction of HIV-1 with cellular microRNA biogenesis and effector mechanisms. *mBio.* 2013, 4:10.1128/mbio.00193-13.
- White, T. B. and Lambowitz, A. M., The Retrohoming of Linear Group II Intron RNAs in *Drosophila melanogaster* Occurs by Both DNA Ligase 4-Dependent and -Independent Mechanisms. *PLoS Genetics.* 2012, 8(2):e1002534. (16 pages).
- Wiedenheft, B., et al., RNA-guided complex from a bacterial immune system enhances target recognition through seed sequence interactions. *PNAS.* 2011, 108(25):10092-10097. (7 pages).
- Wiedenheft, B., et al., Structures of the RNA-guided surveillance complex from a bacterial immune system. *Nature.* 2011, 477:486-489. (5 pages).
- Wiedenheft, B., et al., RNA-Guided Genetic Silencing Systems in Bacteria and Archaea. *Nature.* 2012, 482:331-338.
- Wieland, M., et al., Engineering of ribozyme-based riboswitches for mammalian cells. *Methods.* 2012, 56(3):351-357. (7 pages).
- Wilen, C. B., et al., Engineering HIV-Resistant Human CD4+ T Cells with CXCR4-Specific Zinc-Finger Nucleases. *PLoS Pathogens.* 2011, 7(4):e1002020. (15 pages).
- Wilusz, J. E., et al., tRNAs marked with CCACCA are targeted for degradation. *Science.* 2011, 334(6057):817-821.
- Woo, J. W., et al., DNA-free genome editing in plants with preassembled CRISPR-Cas9 ribonucleoproteins. *Nature Biotechnology.* 2015, 33:1162-1164. (4 pages).
- Wood, A. J., et al., Targeted Genome Editing Across Species Using ZFNs and TALENs. *Science.* 2011, 333 (6040):307 and Supplementary Materials. (27 pages).
- Wu, S. et al., Establishment of a PEG-mediated protoplast transformation system based on DNA and CRISPR/Cas9 ribonucleoprotein complexes for banana. *BMC Plant Biol.* 2020, 20:425. (10 pages).
- Wu, Z., et al., Effect of Genome Size on AAV Vector Packaging. *Molecular Therapy.* 2010, 18(1):80-86. (7 pages).
- Xu, K., The Next Generation Biotechnology for Apple Improvement and Beyond: The CRISPR/Cas9 Story. *New York Fruit Quarterly.* 2013, 21(4):19-22. (4 pages).
- Yamano, S., et al., Comparison of Transfection Efficiency of Nonviral Gene Transfer Reagents. *Mol Biotechnol.* 2010, 46:287-300. (14 pages).
- Yang, H., et al., One-Step Generation of Mice Carrying Reporter and Conditional Alleles by CRISPR/Cas-Mediated Genome Engineering. *Cell.* 2013, 154(6):1370-1379 and Supplementary Materials.
- Yarris, 2012, "Programmable DNA scissors found for bacterial immune system", downloaded from <http://newscenter.lbl.gov/2012/06/28/programmabledna.scissors/> (4 pages).
- Yi, Y., et al., Current Advances in Retroviral Gene Therapy. *Current Gene Therapy.* 2011, 11(3):218-228.
- Yu, Z., et al., Highly Efficient Genome Modifications Mediated by CRISPR/Cas9 in *Drosophila*. *Genetics.* 2013, 195 (1):289-291.
- Zaret, K. S., and Carroll, J. S., Pioneer transcription factors: establishing competence for gene expression. *Genes & Dev.* 2011, 25:2227-2241.
- Zhang, F., et al., Efficient construction of sequence-specific TAL effectors for modulating mammalian transcription. *Nat Biotechnol.* 2011, 29(2):149-153 and Supplementary Materials. (20 pages).
- Zhang, Y., et al., Processing-Independent CRISPR RNAs Limit Natural Transformation in *Neisseria meningitidis*. *Molecular Cell.* 2013, 50(4):488-503. (16 pages).
- Zhang, Y. et al., A highly efficient rice green tissue protoplast system for transient gene expression and studying light/chloroplast-related processes. *Plant Methods* 2011, 7(1):30. (14 pages).

(56)

References Cited

OTHER PUBLICATIONS

- Zou, J., et al., Oxidase-deficient neutrophils from X-linked chronic granulomatous disease iPS cells: functional correction by zinc finger nuclease-mediated safe harbor targeting. *Blood*. 2011, 117(21):5561-5572. (13 pages).
- Zuris, J. A., et al., Efficient Delivery of Genome-Editing Proteins In Vitro and In Vivo. *Nat. Biotechnol.* 2015, 33 (1):73-80. (26 pages).
- Handel, E-M., et al., Versatile and Efficient Genome Editing in Human Cells by Combining Zinc-Finger Nucleases with Adeno-Associated Viral Vectors. *Human Gene Therapy*. 2011, 23(3):321-329. (9 pages).
- Haurwitz, R. E., et al., Sequence- and structure-specific RNA processing by a CRISPR endonuclease. *Science*. 2010, 329(5997):1355-1358. (4 pages).
- PCT International Search Report dated Jan. 27, 2014, in relation to International Application No. PCT/KR2013/009488 filed Oct. 23, 2013, pp. 1-5.
- Hocine, S., et al., RNA processing and export. *Cold Spring Harb Perspect Biol.* 2010, 2:a000752.
- Holt, N., et al., Zinc finger nuclease-mediated CCR5 knockout hematopoietic stem cell transplantation controls HIV-1 in vivo. *Nat Biotechnol.* 2010, 28(8):839-847. (26 pages).
- Horvath, P., and Barrangou, R., RNA-guided genome editing a la carte. *Cell Res.* 2013, 23:733-734.
- Horvath, P., and Barrangou, R., CRISPR/Cas, the immune system of bacteria and archaea. *Science*. 2010, 327 (5962):167-170. (4 pages).
- Hoshijima, K., et al., Highly efficient CRISPR-Cas9-based methods for generating deletion mutations and F0 embryos that lack gene function in zebrafish. *Developmental Cell*. 2019, 51(5):645-657.e4. (35 pages).
- Hruscha, A., et al., Efficient CRISPR/Cas9 genome editing with low off-target effects in zebrafish. *Development*. 2013, 140(24):4982-4987. Supplementary Information (12 pages).
- Hsu, P. D., et al., DNA targeting specificity of RNA-guided Cas9 nucleases. *Nat Biotechnol.* 2013, 31(9):827-832.
- Hu, P., et al., Comparison of various nuclear localization signal-fused Cas9 proteins and Cas9 mRNA for genome editing in zebrafish. *G3 Genes/Genomes/Genetics*. 2018, 8(3):823-831. (9 pages).
- Huang, P., et al., Heritable gene targeting in zebrafish using customized TALENs. *Nature Biotechnology*. 2011, 29 (8):699-700. (2 pages).
- Hwang, W. Y., et al., Efficient genome editing in zebrafish using a CRISPR-Cas system. *Nat Biotechnol.* 2013, 31:227-229 and Supplementary Materials.
- Hwang, W. Y., et al., Heritable and Precise Zebrafish Genome Editing Using a CRISPR-Cas System. *PLoS ONE*. 2013, 8(7):e68708 and Supplementary Materials. (14 pages).
- Jani, B. and Fuchs R., In Vitro Transcription and Capping of Gaussia Luciferase mRNA Followed by HeLa Cell Transfection. *J. Vis. Exp.* 2012, 61:3702. (9 pages).
- Japanese Office Action dated Nov. 24, 2015 for Japanese Patent Application No. 2015-538033, pp. 1-10 (with English Translation).
- Jensen, N. M., et al., An update on targeted gene repair in mammalian cells: methods and mechanisms. *J. Biomed. Sci.* 2011;18:10. (14 pages).
- Jeyarajan, S., et al., 2010, Plasmid DNA delivery into MDA-MB-453 cells mediated by recombinant Her-NLS fusion protein. *International Journal of Nanomedicine*. 2010, 5:725-733. (9 pages).
- Jiang, F., et al., 2011, Structural basis of RNA recognition and activation by innate immune receptor RIG-I. *Nature*. 2011, 479:423-429.
- Jiang, W., et al., Demonstration of CRISPR/Cas9/sgRNA-mediated targeted gene modification in *Arabidopsis*, tobacco, sorghum and rice. *Nucleic Acids Research*. 2013, 41(20):e188.
- Jiang, W., et al., RNA-guided editing of bacterial genomes using CRISPR-Cas systems. *Nat. Biotechnol.* 2013, 31:233-239.
- Jiang, W., et al., CRISPR-assisted editing of bacterial genomes. *Nat Biotechnol.* 2013, 31(3):233-239.
- Jinek, M., et al., A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity. *Science*. 2012, 337(6096):816-821 and Supplementary Materials. (7 pages).
- Jinek, M., et al., Methods and Compositions for RNA-Directed Site-Specific DNA Modification. U.S. Appl. No. 61/652,086, filed May 25, 2012.
- Jinek, M., et al., RNA-programmed genome editing in human cells. *eLife*. 2013, 2:e00471.
- Jore, M. M., et al., Structural basis for CRISPR RNA-guided DNA recognition by Casade. *Nat. Struct. Mol. Biol.* 2011, 18(5):529-536. (9 pages).
- Joung, J. K. and Sander J. D., TALENs: A widely applicable technology for targeted genome editing. *Nat. Rev. Mol. Cell Biol.* 2013, 14:49-55. (7 pages).
- Kang, H. C. and Bae, Y. H., Co-delivery of small interfering RNA and plasmid DNA using a polymeric vector incorporating endosomolytic oligomeric sulfonamide. *Biomaterials*. 2011, 32(21):4914-4924. (11 pages).
- Karginov, F. V. and Hannon, G. J., The CRISPR System: Small RNA-Guided Defense in Bacteria and Archaea. *Molecular Cell*. 2010, 37(1):7-19. (13 pages).
- Karvelis, T., et al., crRNA and tracrRNA guide Cas9-mediated DNA interference in *Streptococcus thermophilus*. *RNA Biology*, 2013, 10(5):841-851.
- Katic, I. and Großhans, H., Targeted Heritable Mutation and Gene Conversion by Cas9-CRISPR in *Caenorhabditis elegans*. *Genetics*. 2013, 195(3):1173-1176. (4 pages).
- Kim, E., et al., Precision genome engineering with programmable DNA-nicking enzymes. *Genome Res.* 2012, 22:1327-1333.
- Kim, H., et al., Surrogate reporters for enrichment of cells with nuclease-induced mutations. *Nat Methods*. 2011, 8:941-943.
- Kim, S., et al., Highly efficient RNA-guided genome editing in human cells via delivery of purified Cas9 ribonucleoproteins. *Genome Res.* 2014, 24:1012-1019. (9 pages).
- Kim, S., et al., 2018, CRISPR RNAs trigger innate immune responses in human cells. *Genome Res.* 2018, 28: 367-373. (8 pages).
- Kim, T. K., and Eberwine, J. H., Mammalian cell transfection: the present and the future. *Anal Bioanal Chem.* 2010, 397:3173-3178. (6 pages).
- Kolomeisky, A. B., Physics of protein—DNA interactions: mechanisms of facilitated target search. *Phys. Chem. Chem. Phys.* 2011, 13:2088-2095. (8 pages).
- Kondo, S., et al., Highly Improved Gene Targeting by Germline-Specific Cas9 Expression in *Drosophila*. *Genetics*. 2013, 195(3):715-721.
- Koo, T., et al., Measuring and Reducing Off-Target Activities of Programmable Nucleases Including CRISPR-Cas9. *Molecules and Cells*. 2015, 38(6):475-481. (7 pages).
- Kouranova, E., et al., CRISPRs for optimal targeting: delivery of CRISPR components as DNA, RNA, and protein into cultured cells and single-cell embryos. *Human Gene Therapy*. 2016, 27(6):464-475. (12 pages).
- Lambowitz, A. M. and Zimmerly, S., Group II Introns: Mobile Ribozymes that Invade DNA. *Cold Spring Harb Perspect Biol.* 2011, 3:a003616. (19 pages).
- Lane, J., et al., Targeting RHOc by way of ribozyme transgene in human breast cancer cells and its impact on cancer invasion. *World J Oncol.* 2010, 1(1):7-13. (7 pages)
- Lane, J., et al., Targeting RHOc by way of ribozyme transgene in human breast cancer cells and its impact on cancer invasion. *World J Oncol.* 2010, 1(1):7-13. (7 pages).
- Larson, D. R., et al., Real-time observation of transcription initiation and elongation on an endogenous yeast gene. *Science*. 2011, 332(6028):475-478.
- Larsen, H. O., et al., Nonviral transfection of leukemic primary cells and cells lines by siRNA—a direct comparison between Nucleofection and Accell delivery. *Experimental Hematology*. 2011, 39:1081-1089. (9 pages).
- Larson, M. H. et al., CRISPR interference (CRISPRi) for sequence-specific control of gene expression. *Nat. Protoc.* 2013, 8:2180-2196.
- Ledford, H., Targeted gene editing enters clinic. *Nature*. 2011, 471:16. (1 page).

(56)

References Cited

OTHER PUBLICATIONS

Lee, H. Y., et al., Targeted chromosomal deletions in human cells using zinc finger nucleases. *Genome Res.* 2010, 20:81-89.

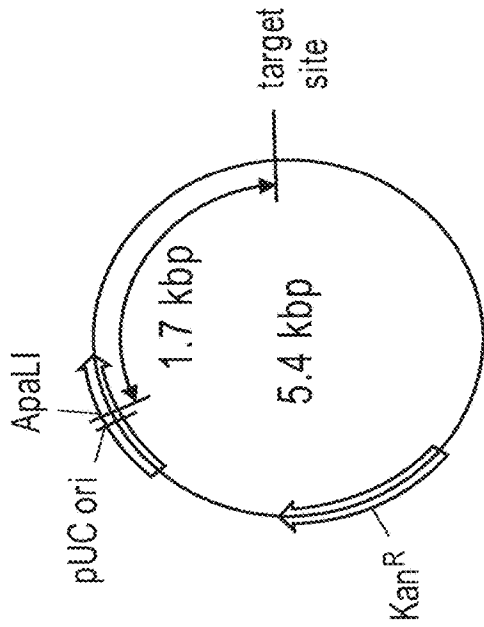
Lee, H. J., et al., Targeted chromosomal duplications and inversions in the human genome using zinc finger nucleases. *Genome Res.* 2012, 22:539-548.

Lee, J-S., et al., RNA-guided genome editing in *Drosophila* with the purified Cas9 protein. *G3 Genes|Genomes|Genetics.* 2014, 4(7):1291-1295. (5 pages).

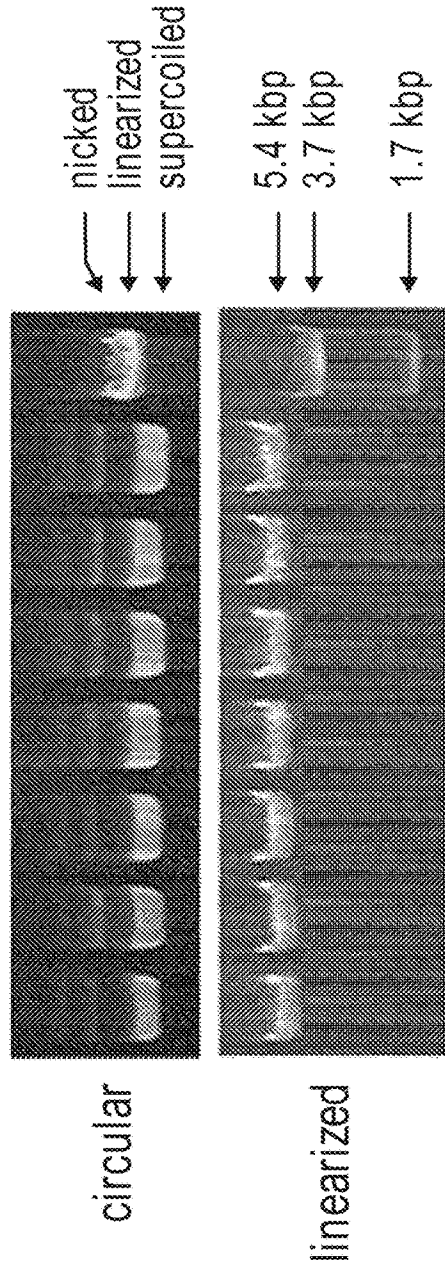
Notice of Allowance and Fees Due dated Feb. 4, 2015, U.S. Appl. No. 14/226,274, filed Mar. 26, 2014, pp. 1-22.

* cited by examiner

FIG. 1B



| | | | | | | | | | | | | | | | | | | | | | |
|------------------|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| Cas9: | - | + | - | + | - | + | - | + | - | + | - | + | - | + | - | + | - | + | - | + | |
| guide RNA: | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - |
| target sequence: | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - |



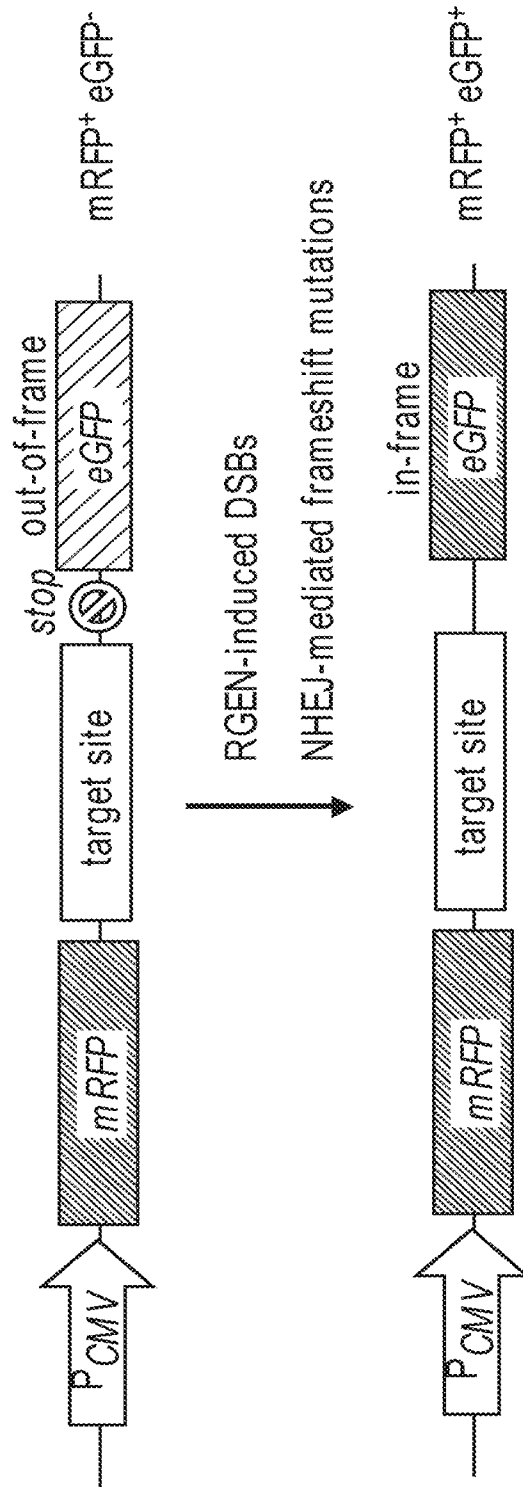


FIG. 2A

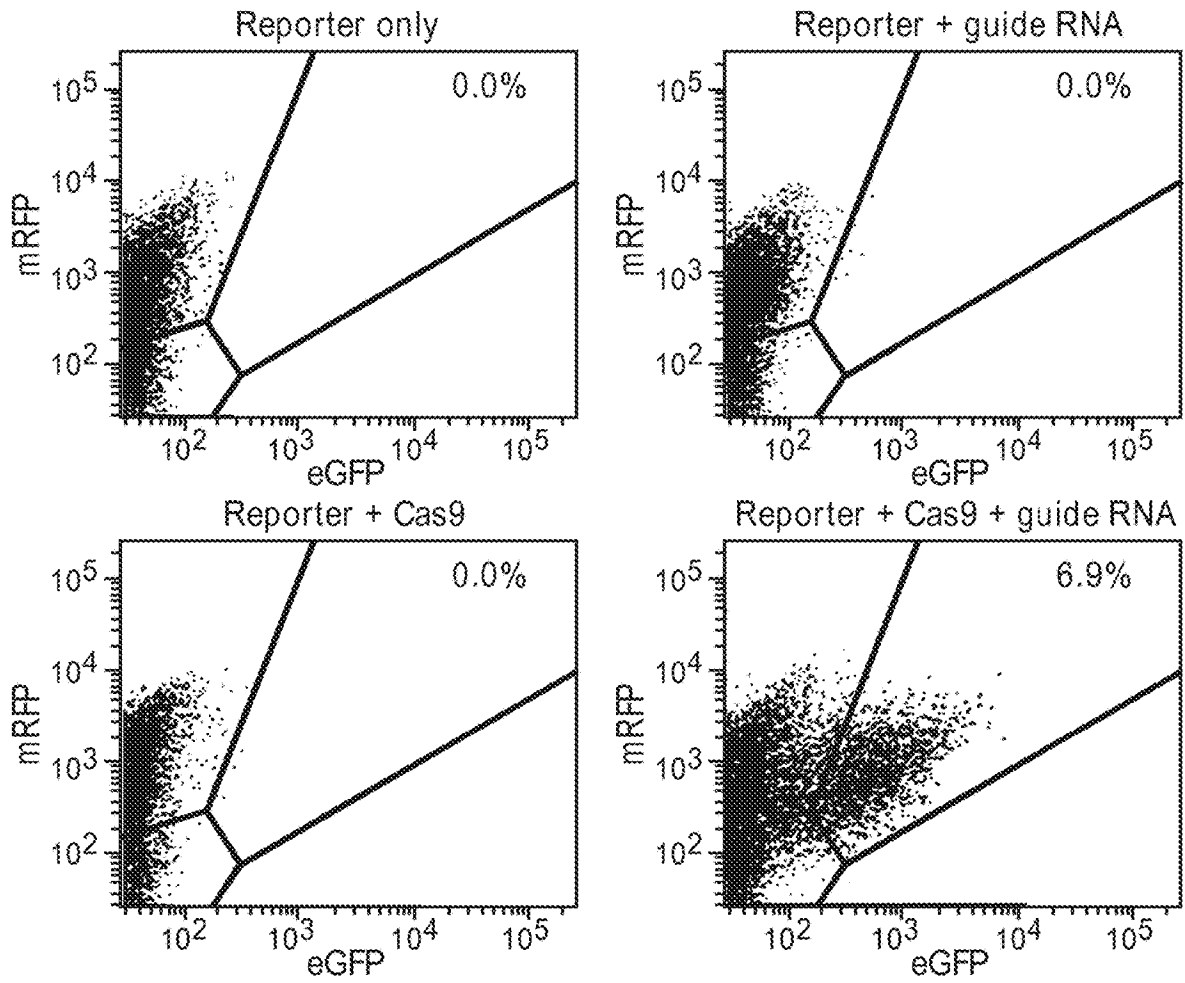


FIG. 2B

CCR5

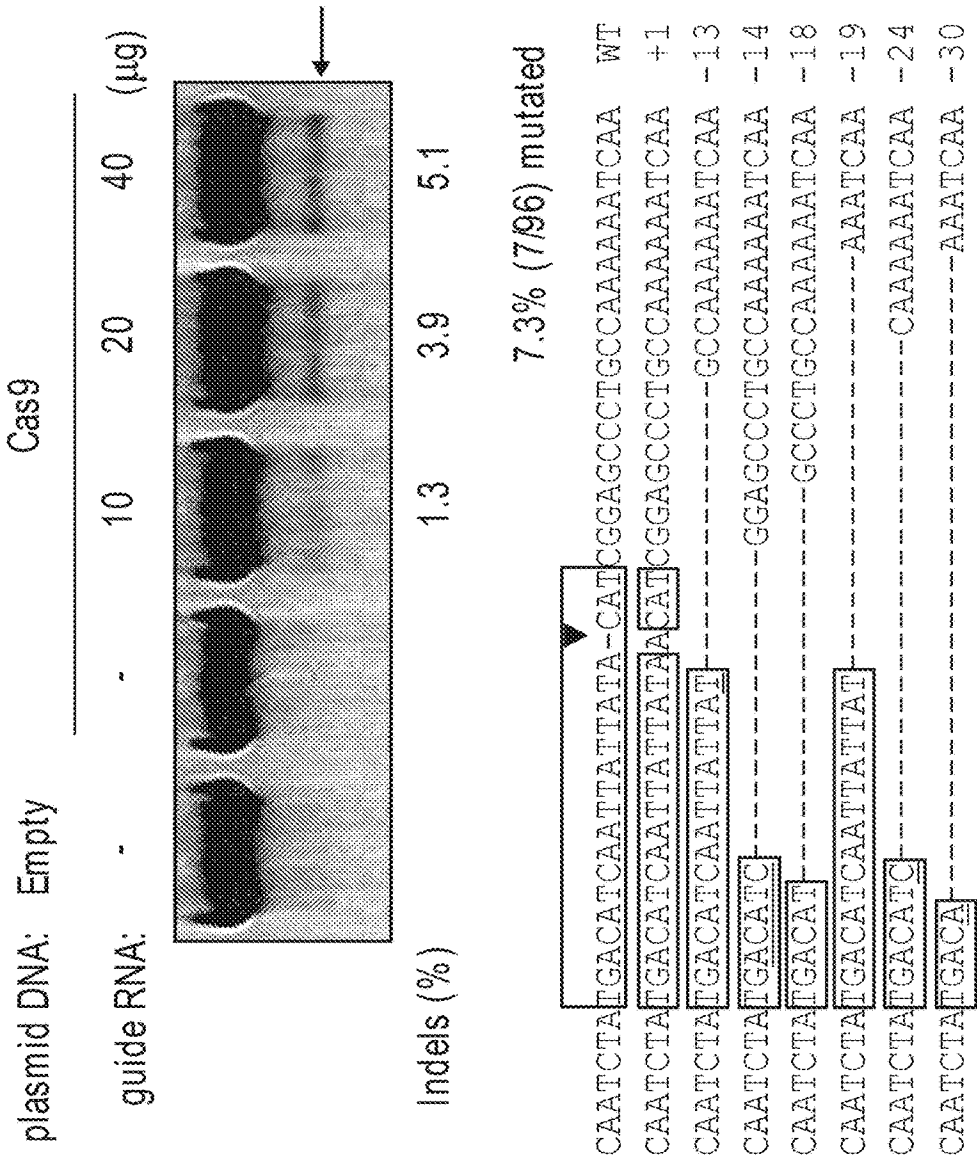
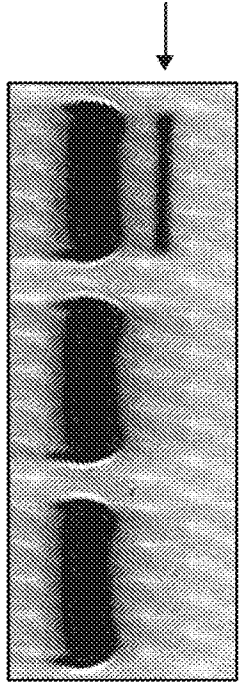


FIG. 3A

C4BPB

plasmid DNA: Empty Cas9

guide RNA: - +



Indels (%)

14

8.3% (4/48) mutated

TATGTGCAATGACCACTACATCCT--CAAAGGGCAGCAATCGGAGCCAG WT
 TATGTGCAATGACCACTACATCCTT-CAAAGGGCAGCAATCGGAGCCAG +1
 TATGTGCAATGACCACTACATCCTCTCAAAGGGCAGCAATCGGAGCCAG +2
 TATGTGCAATG AATG-----GAGCCAG -30
 TATGTGCAATGAC-----180 bp-----180

FIG. 3B

| | | | | | | | |
|----------------------|---|-----------|------|-------|-------|-------|------------|
| TGACATCAATTATTATACAT | ▼ | CGG | CCR5 | | | | |
| TGACATCAATTATTATA | g | AT | g | GA | ADCY5 | | |
| TGACATCA | c | TTATTAT | g | CAT | g | GG | KCNJ6 |
| TGACAT | a | AATTATT | c | TACAT | g | GG | CNTNAP2 |
| TGA | a | ATCAATTAT | c | ATA | g | ATCGG | Chr. 5 N/A |

FIG. 4A

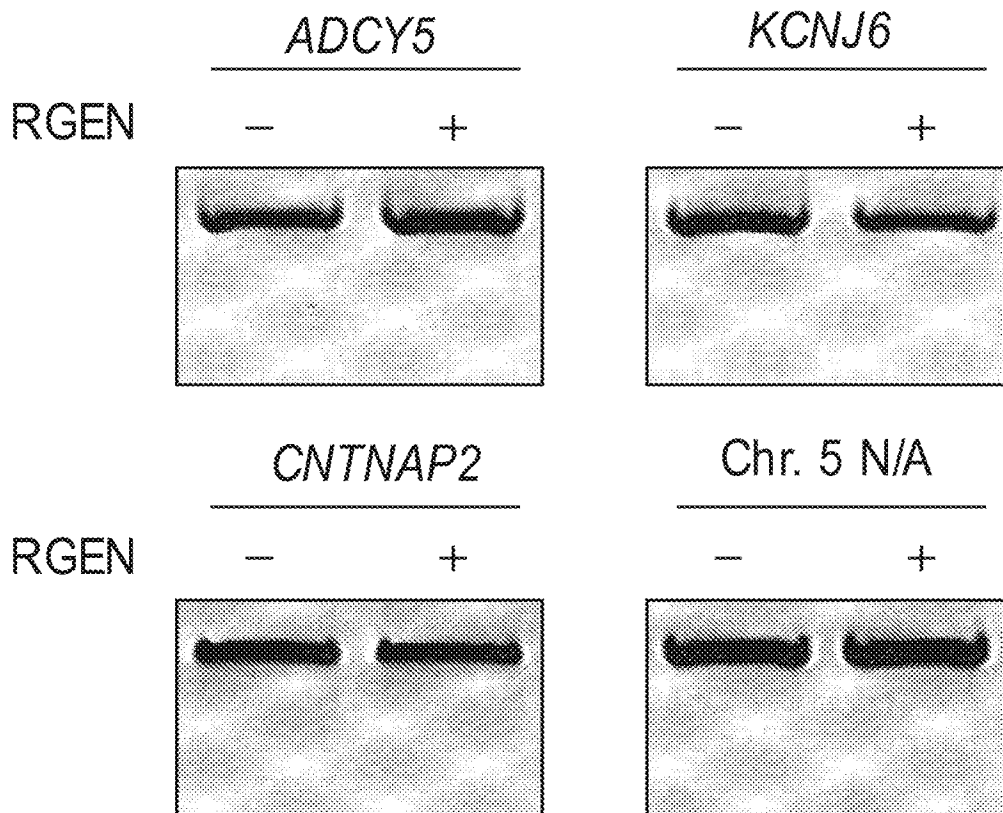


FIG. 4B

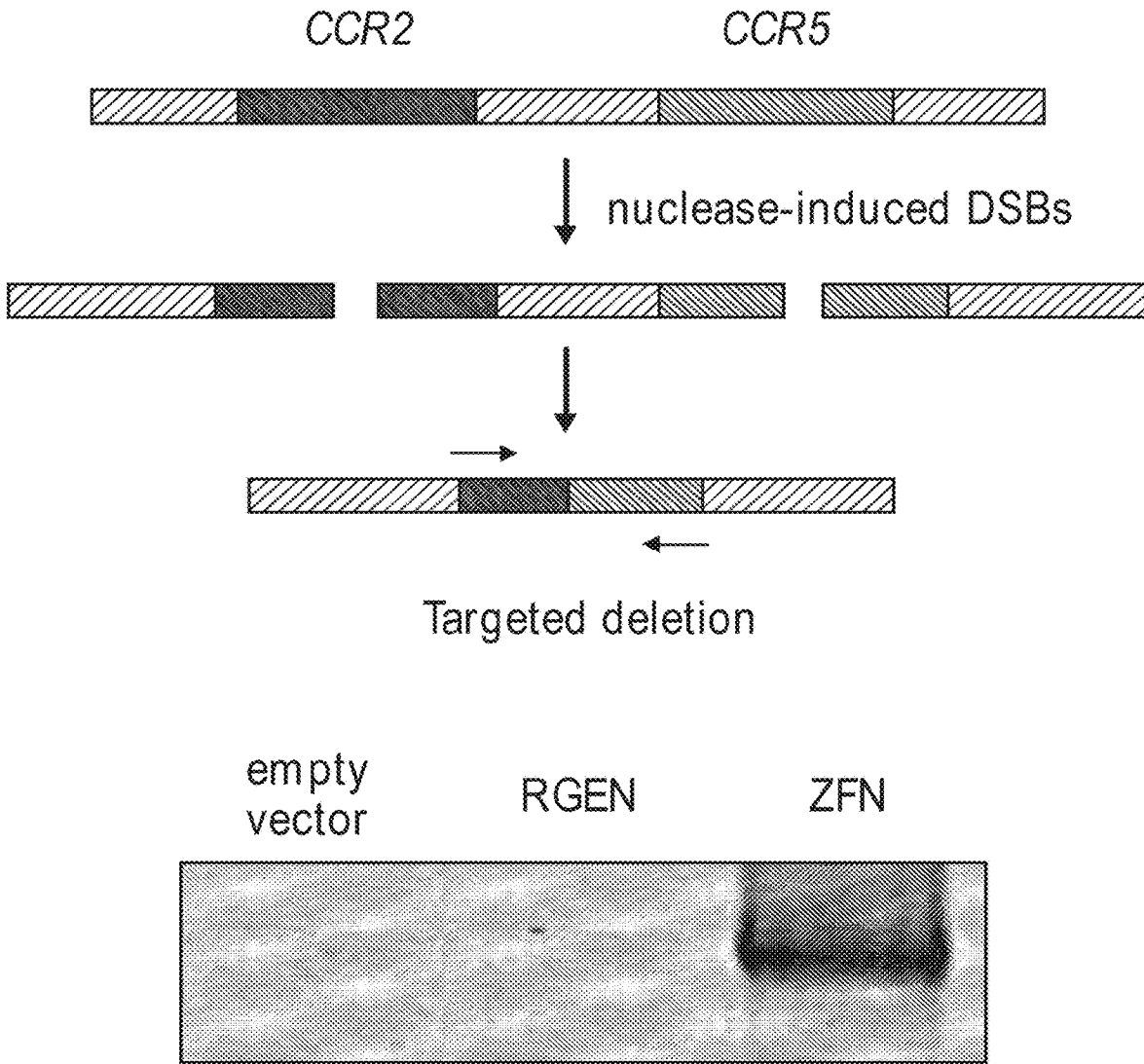


FIG. 4C

Foxn1

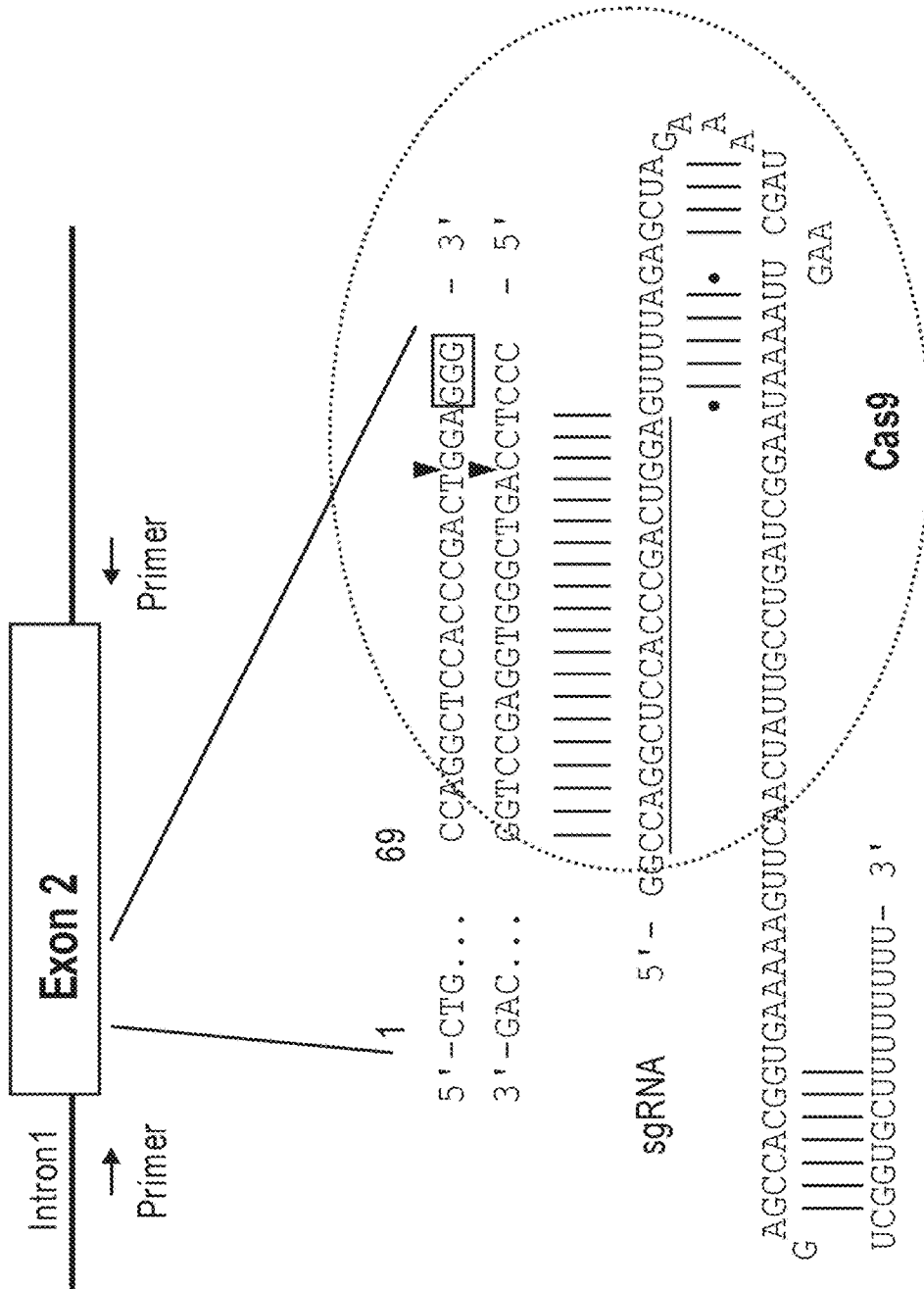


FIG. 5A

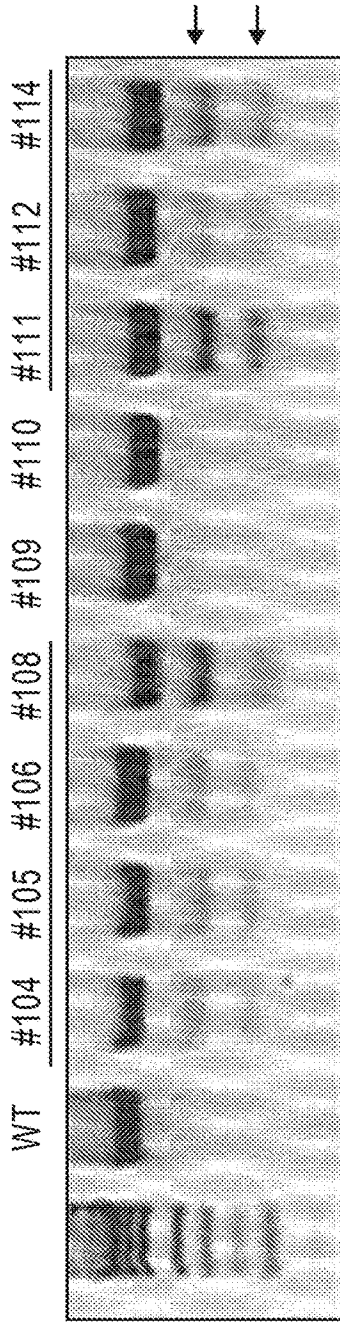


FIG. 5B

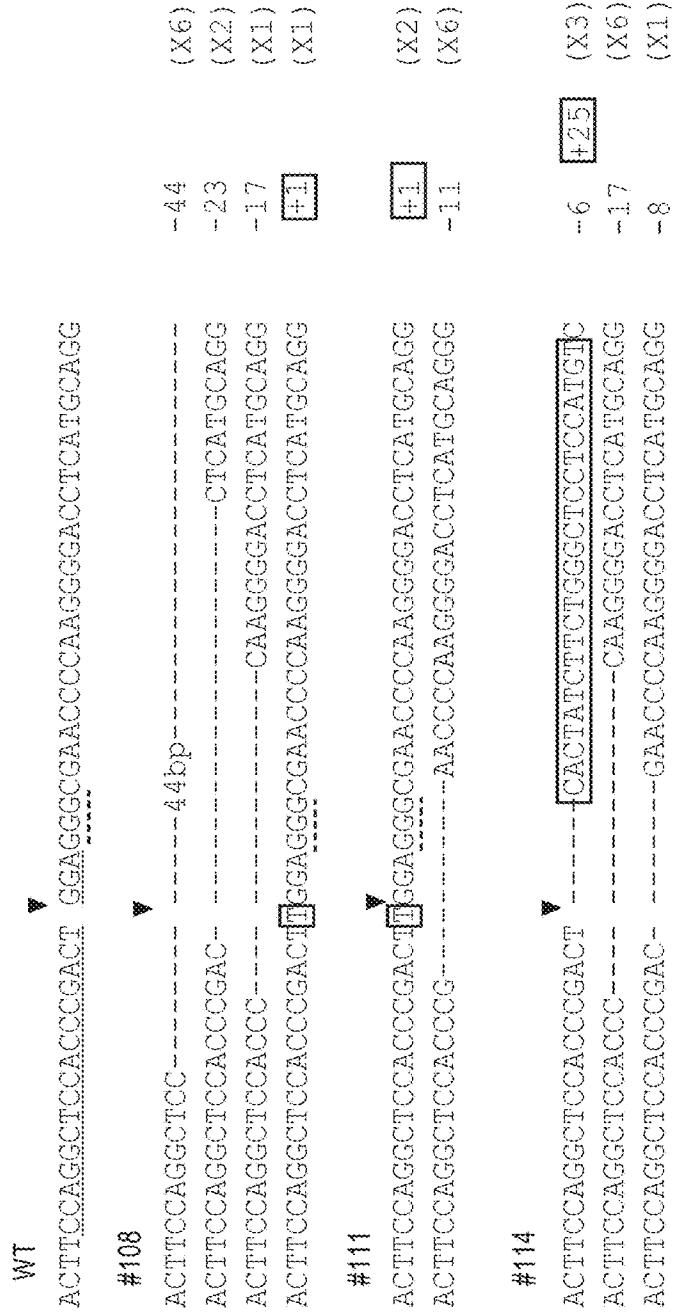


FIG. 5C

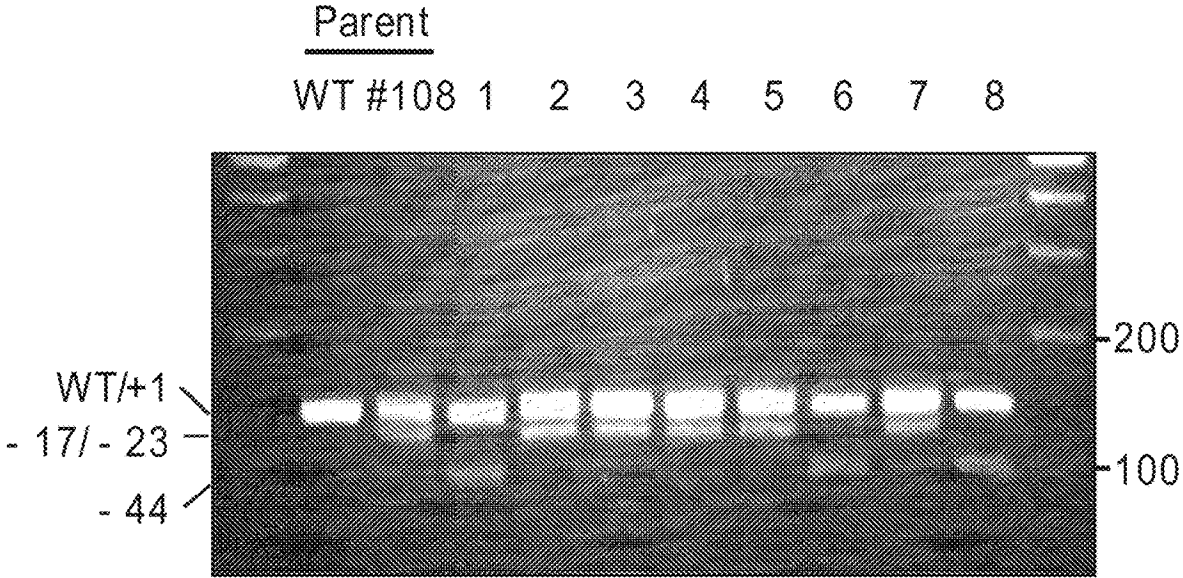


FIG. 5D

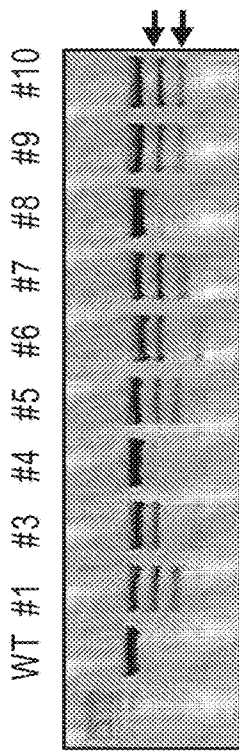


FIG. 6A

| Cas9 mRNA (ng/μl) | sgRNA (ng/μl) | Tested embryos | Mutants (%) |
|-------------------|---------------|----------------|-------------|
| 10 | 1 | 27 | 9 (33) |
| 10 | 10 | 49 | 28 (57) |
| 10 | 100 | 45 | 41 (91) |

FIG. 6B

| sgRNA (ng/μl) | L | P | G | S | T | R | L | E | G | E | P | Q | G | D | L | M | Q | A | Mutants | | |
|---------------|-----|-----|-----|-----|-----|-----|-----|-----|------|-----|-----|-----|-----|-----|-----|-----|-----|-----|---------|--------|---------|
| 100 | A | CTT | CCA | GGC | TCC | ACC | CGA | CT | G | GAA | CCC | CAA | GGG | GAC | CTC | ATG | CAG | GCT | CC WT | | |
| | #1 | A | CTT | CCA | GGC | TCC | ACC | CGA | -- | -- | A | CCC | CAA | GGG | GAC | CTC | ATG | CAG | GCT | CC Δ11 | |
| | #3 | A | CTT | CCA | GGC | TCC | ACC | CGA | -- | -- | A | CCC | CAA | GGG | GAC | CTC | ATG | CAG | GC- | -- | Δ11+Δ17 |
| | #2 | A | CTT | CCA | GGC | TCC | ACC | CGA | C- | -- | -- | -- | -- | -- | -- | -- | -- | -- | -- | -- | Δ57 |
| | #5 | A | CTT | CCA | GGC | TCC | ACC | CGA | C- | GAA | CCC | CAA | GGG | GAC | CTC | ATG | CAG | GCT | CC | Δ11 | |
| | #11 | A | CTT | CCA | GGC | TCC | ACC | CGA | C- | -- | -- | CAA | GGG | GAC | CTC | ATG | CAG | GCT | CC | Δ17 | |
| | #9 | A | CTT | CCA | GGC | TCC | ACC | CGA | CTTG | GAG | GAA | CCC | CAA | GGG | GAC | CTC | ATG | CAG | GCT | CC | +1 |
| | #11 | A | CTT | CCA | GGC | TCC | A-- | -- | -- | GGT | GAA | CCC | CAA | GGG | GAC | CTC | ATG | CAG | GCT | CC | Δ12 |
| | #9 | A | CTT | CCA | GGC | --- | --- | --- | --- | GGT | GAA | CCC | CAA | GGG | GAC | CTC | ATG | CAG | GCT | CC | Δ72 |
| | #9 | A | CTT | CCA | GGC | --- | --- | --- | --- | -- | -- | AA | GGG | GAC | CTC | ATG | CAG | GCT | CC | Δ25 | |
| | #9 | A | CTT | CCA | GGC | T-- | --- | --- | --- | -- | -- | AA | GGG | GAC | CTC | ATG | CAG | GCT | CC | Δ24 | |

FIG. 6C

Pronucleus injection

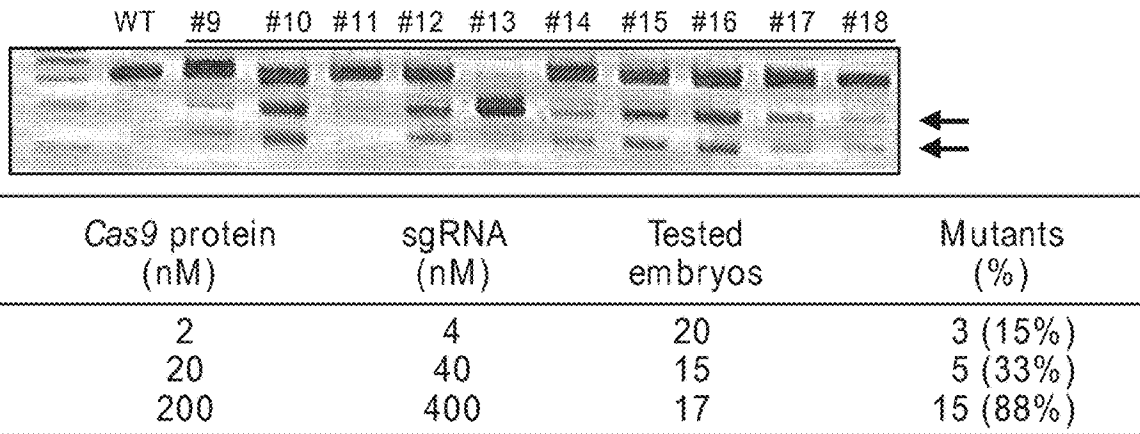


FIG. 7A

Intra-cytoplasmic injection

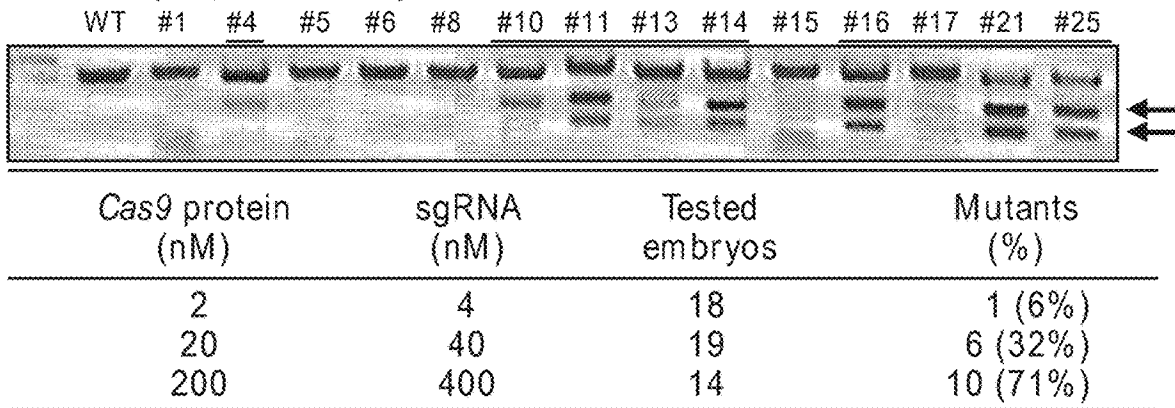


FIG. 7B

| Sequence | Indels | Embryo no. |
|--|--------|------------|
| ACTTCCAGGCTCCACCCGACTGGAGGGCGGAACCCCAAGGGACCTCATGCAG | WT | |
| ACTTCCAGGCGAACCC-----AAGGGACCTCATGCAG | Δ18 | 2 |
| ACTTCCAGGCTCCAC-----AAGGGACCTCATGCAG | Δ20 | 1 |
| ACTTCCAGGCTCCACCC-----AAGGGACCTCATGCCC | Δ19 | 1 |
| ACTTCCAGGCTCCACCC-----CAAGGGACCTCATGCAG | Δ17 | 1 |
| ACTTCCAGGCTCCACCCGA-----ACCCCAAGGGACCTCATGCAG | Δ11 | 3 |
| ACTTCCAGGCTCCACCCGAA--GGAGGGCGAACCCCAAGGGACCTCATGCAG | Δ3+1 | 1 |
| ACTTCCAGGCTCCACCCGACT--AGGGCGAACCCCAAGGGACCTCATGCAG | Δ2 | 1 |
| ACTTCCAGGCTCCACCCGACTGGAGGGCGAACCCCAAGGGACCTCATGCAG | +1 | 1 |
| ACTTCCAGGCTCCACCCGACTGGAGGGCGAACCCCAAGGGACCTCATGCAG | +1 | 10 |
| ACTTCCAGGCTCCACCCGA-----GGCGAACCCCAAGGGACCTCATGCAG | Δ6 | 1 |
| ACTTCCAGGCTCCACCCGA-----GGCGAACCCCAAGGGACCTCATGCAG | Δ5 | 2 |
| ACTTCCAGGCTCCACCC-----TCATGCAG | Δ28 | 1 |
| -----AGGGCGAACCCCAAGGGACCTCATGCAG | Δ126 | 1 |
| Total | | 26 |

FIG. 7C

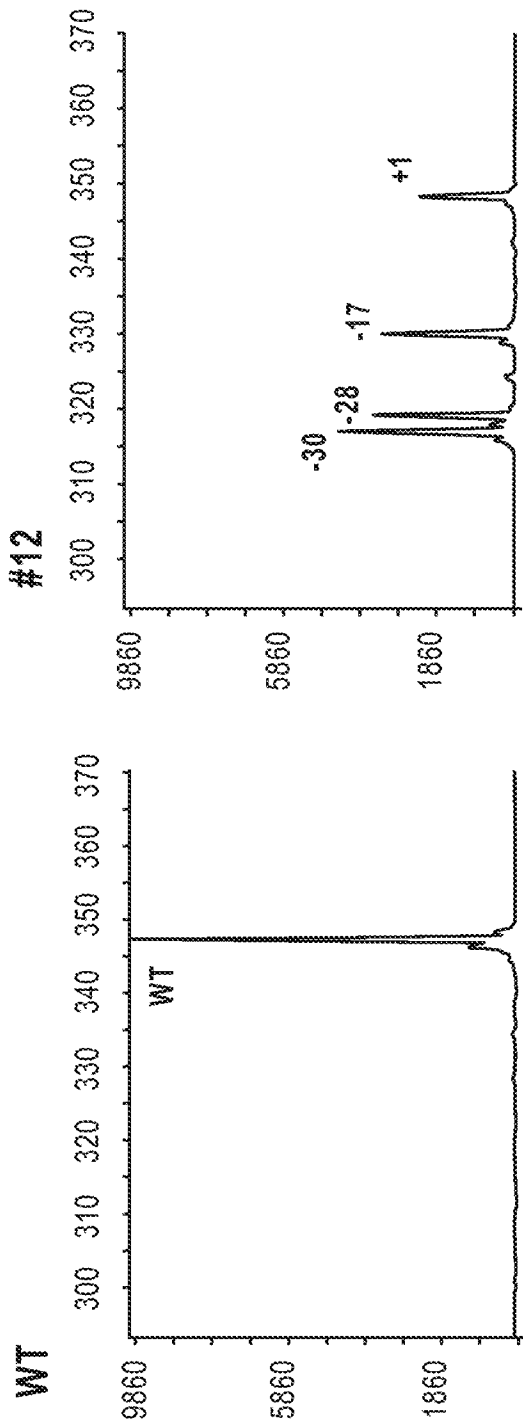


FIG. 8A

FIG. 8B

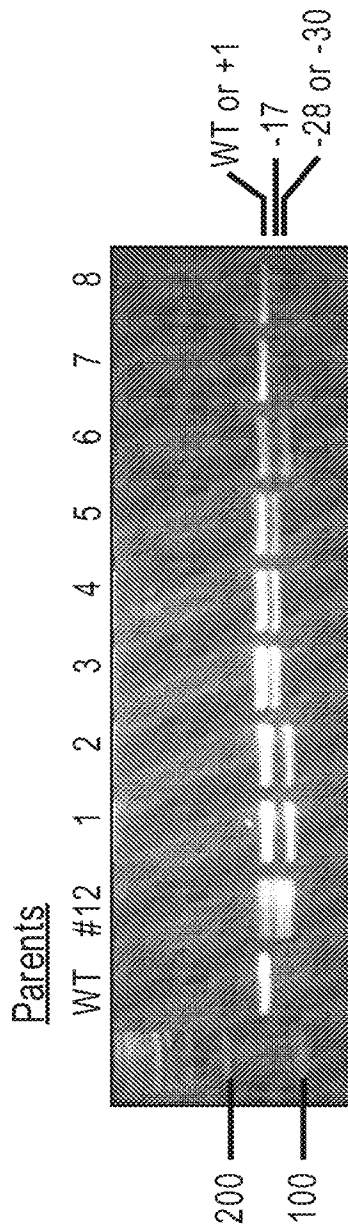


FIG. 8C

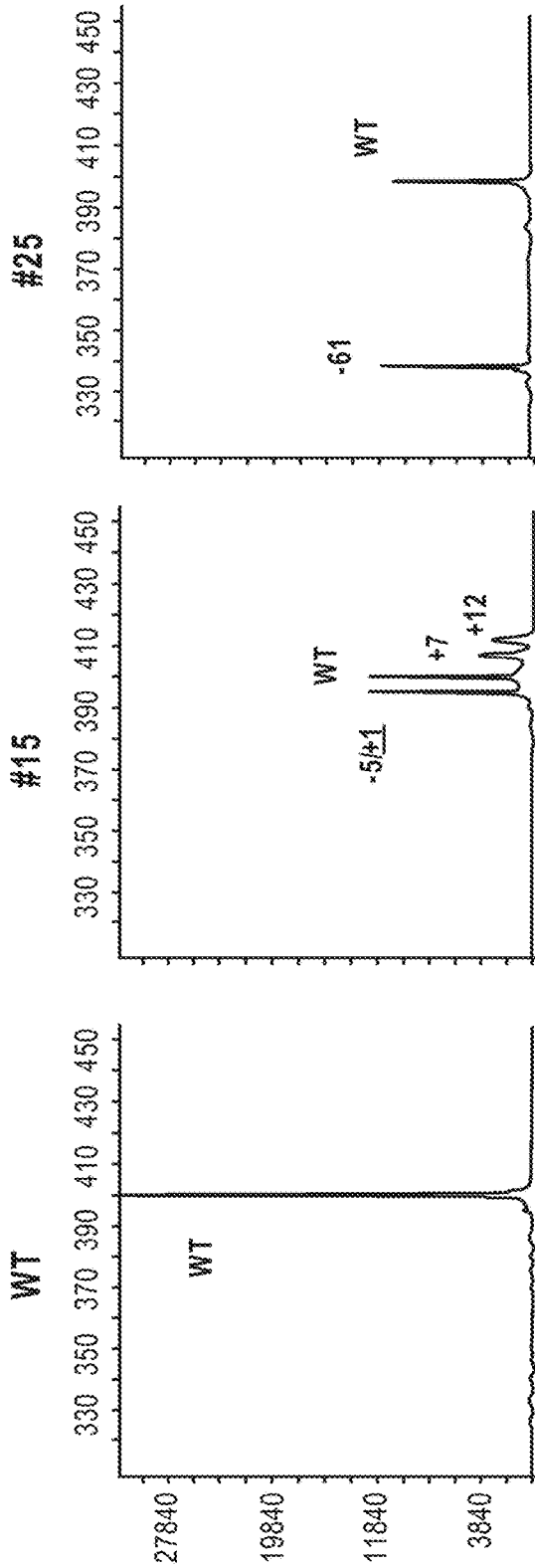


FIG. 9A

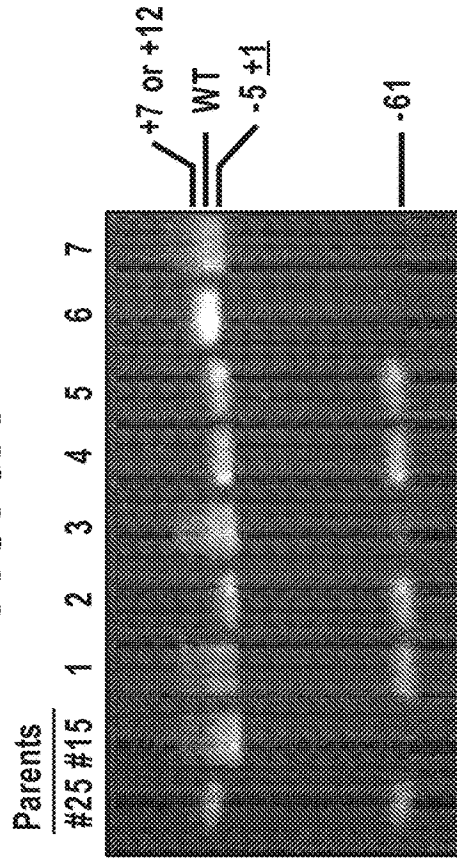
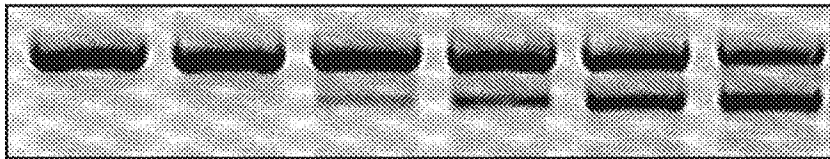


FIG. 9B

CCR5 #4

| | | | | | | | |
|--------------|----------|----------|-----------|---------|---------|--------|---------|
| sgRNA | - | 100 (29) | 10(2.9) | 30(8.8) | 100(29) | 50(15) | μg (μM) |
| Cas9 protein | 225 (14) | - | 22.5(1.4) | 75(4.5) | 225(14) | - | μg (μM) |
| Cas9 plasmid | - | - | - | - | - | 5 | μg |



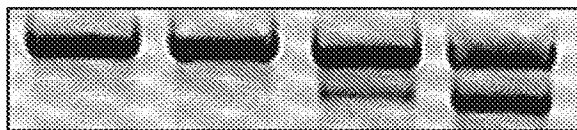
4.8 18 38 47 indel (%)

| | | | | | | | |
|--------------|----------|----------|-----------|---------|---------|--------|---------|
| sgRNA | - | 100 (29) | | | | 50(15) | μg (μM) |
| Cas9 protein | 225 (14) | - | 22.5(1.4) | 75(4.5) | 225(14) | - | μg (μM) |
| Cas9 plasmid | - | - | - | - | - | 5 | μg |



12 30 33 43 indel (%)

| | | | | | |
|--------------|---------|--------|---------|---------|---------|
| crRNA | - | 40(29) | 20(15) | μg (μM) | |
| tracrRNA | - | 80(29) | 40(15) | μg (μM) | |
| Cas9 protein | 225(14) | - | 225(14) | - | μg (μM) |
| Cas9 plasmid | - | - | 5 | μg | |



9.4 42 indel (%)

FIG. 10A

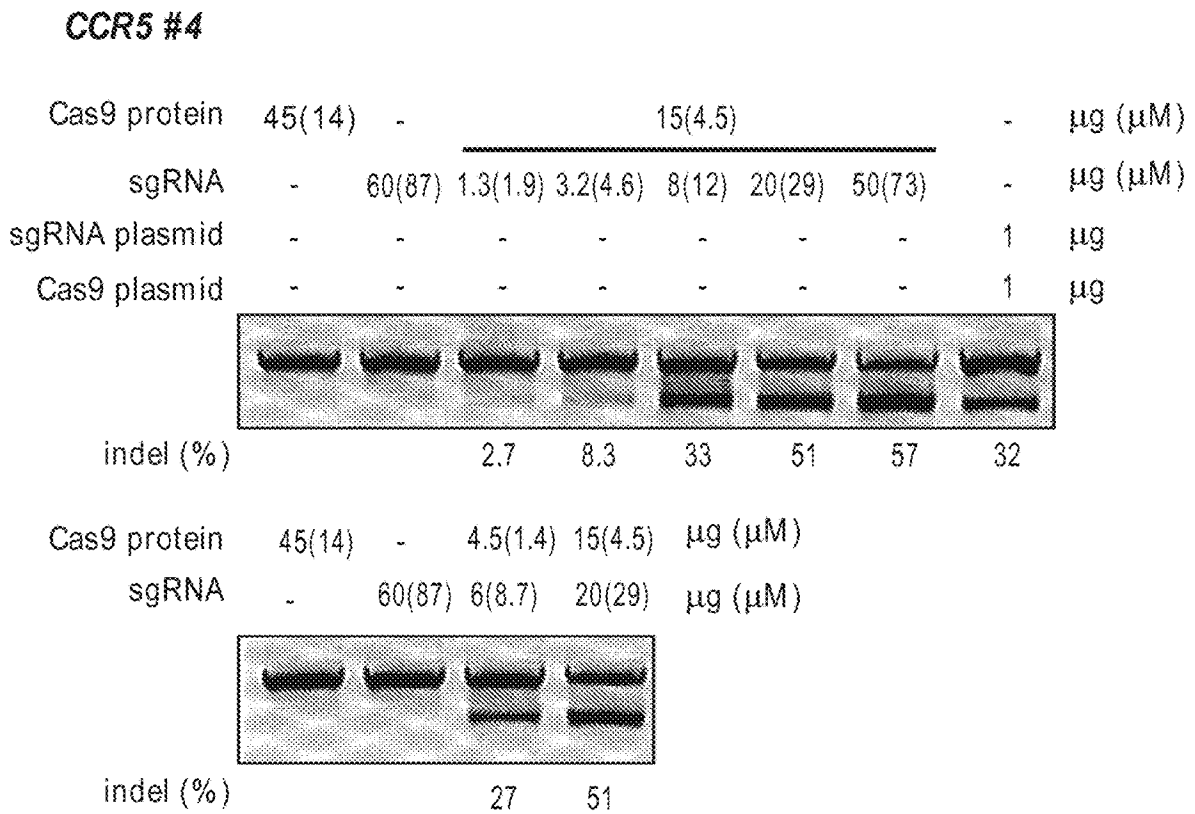


FIG. 10B

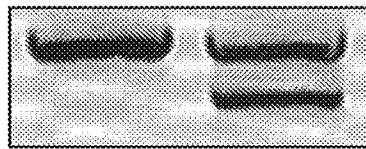
CCR5

CAATCTATGACATCAATTATTATA-CATCGGAGCCCTGCCAAAAAATCAA WT
 CAATCTATGACATCAATTATTAT-----CGGAGCCCTGCCAAAAAATCAA -4
 CAATCTATGACATCAATTAT-----CATCGGAGCCCTGCCAAAAAATCAA -4
 CAATCTATGACATCAATTAT-----CGGAGCCCTGCCAAAAAATCAA -7
 CAATCTATGACATCAATTATTAT--CATCGGAGCCCTGCCAAAAAATCAA -1
 CAATCTATGACATCAATTATTATAACATCGGAGCCCTGCCAAAAAATCAA +1
 CAATCTATGACAA-----GAGCCCTGCCAAAAAATCAA -17, +1

FIG. 10C

ABCC11

| | | | |
|--------------|---|---------|---------|
| Cas9 protein | - | 15(4.5) | μg (μM) |
| sgRNA | - | 20(29) | μg (μM) |



Indel (%) 35

FIG. 10D

ABCC11

TTCTCAAGGCAGCATCATACTTCCCCCACGGTGGGACAGCTGCCCTCCCTGG WT
 TTCTCAAGGCAGCATCATACTTCC-----CTGGGACAGCTGCCCTCCCTGG -6
 TTCTCAAGGCAGCATCATACTTC---CACGGTGGGACAGCTGCCCTCCCTGG -3
 TTCTCAAGGCAGC-----TGCCCTCCCTGG -29
 TTCTCAAGGCAGCATCATACTT-----CCCTCCCTGG -20
 TTCTCAAGGCAGCATCATACTT-----CCCTCCCTGG -20
 TTCTC----- -256

FIG. 10E

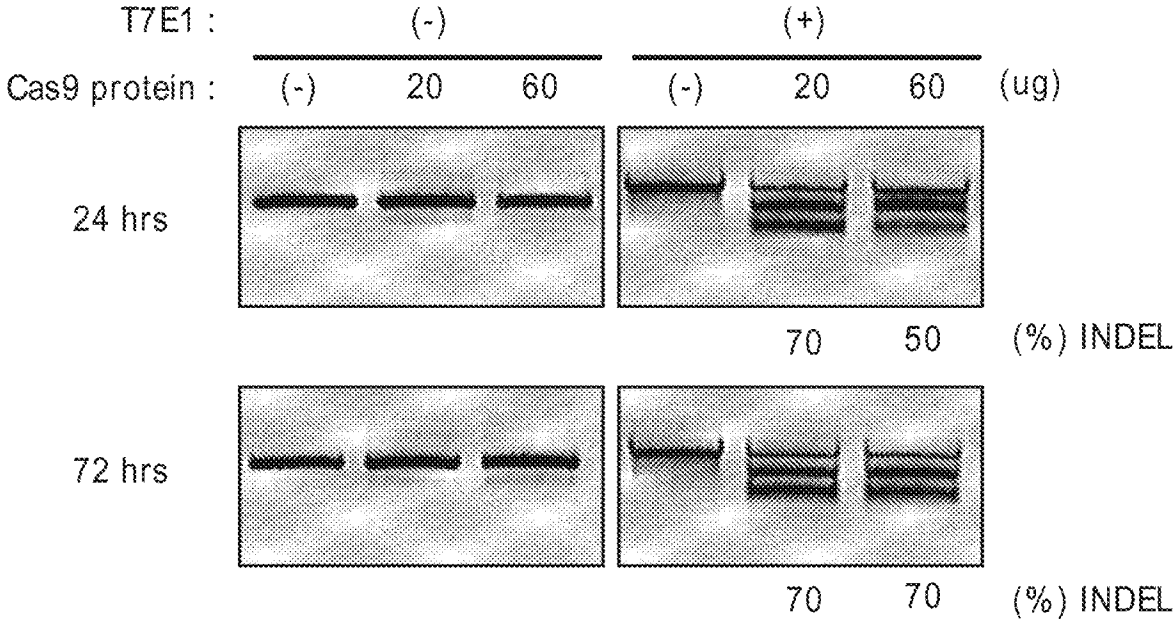


FIG. 11

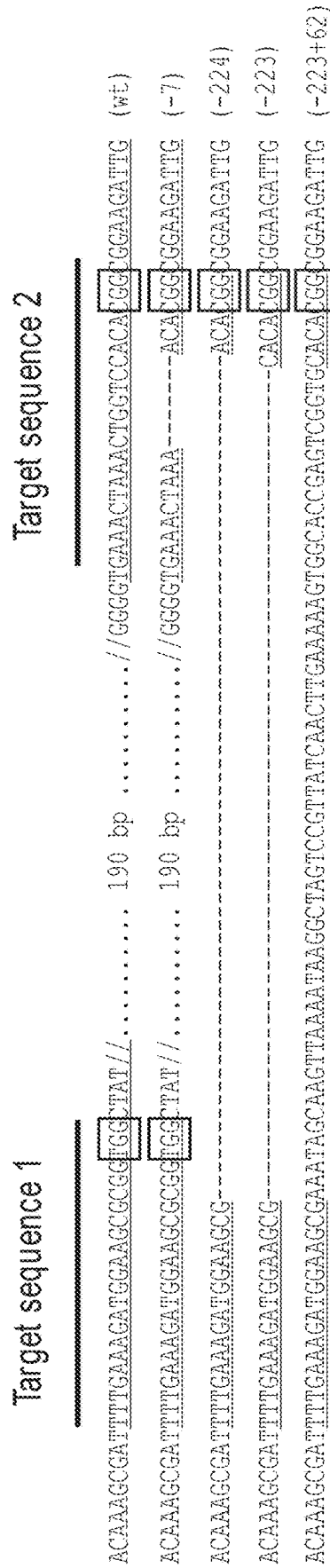


FIG. 12

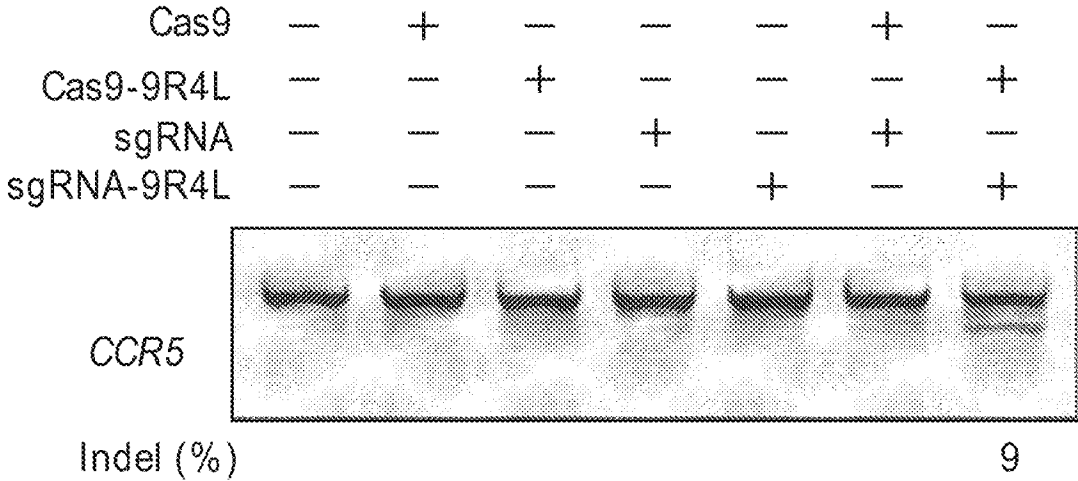


FIG. 13

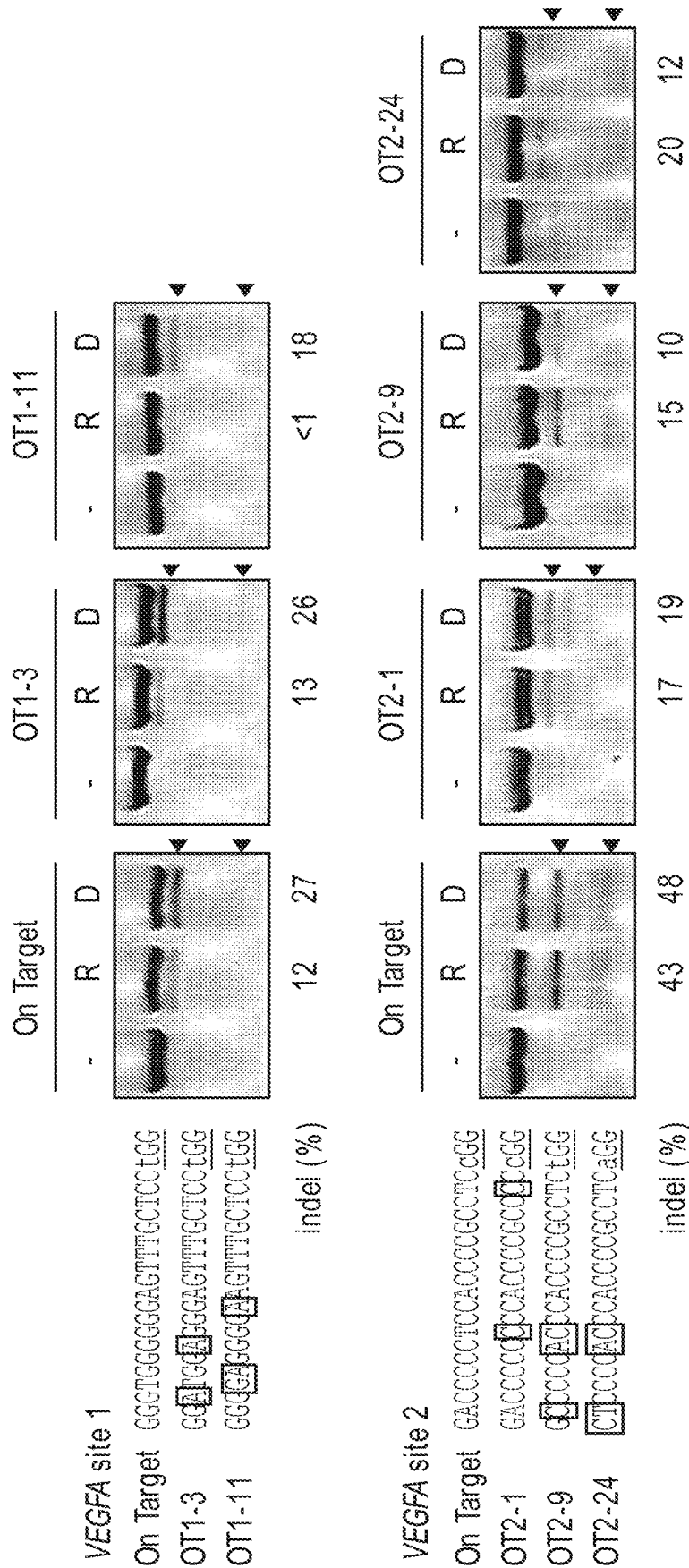


FIG. 14A

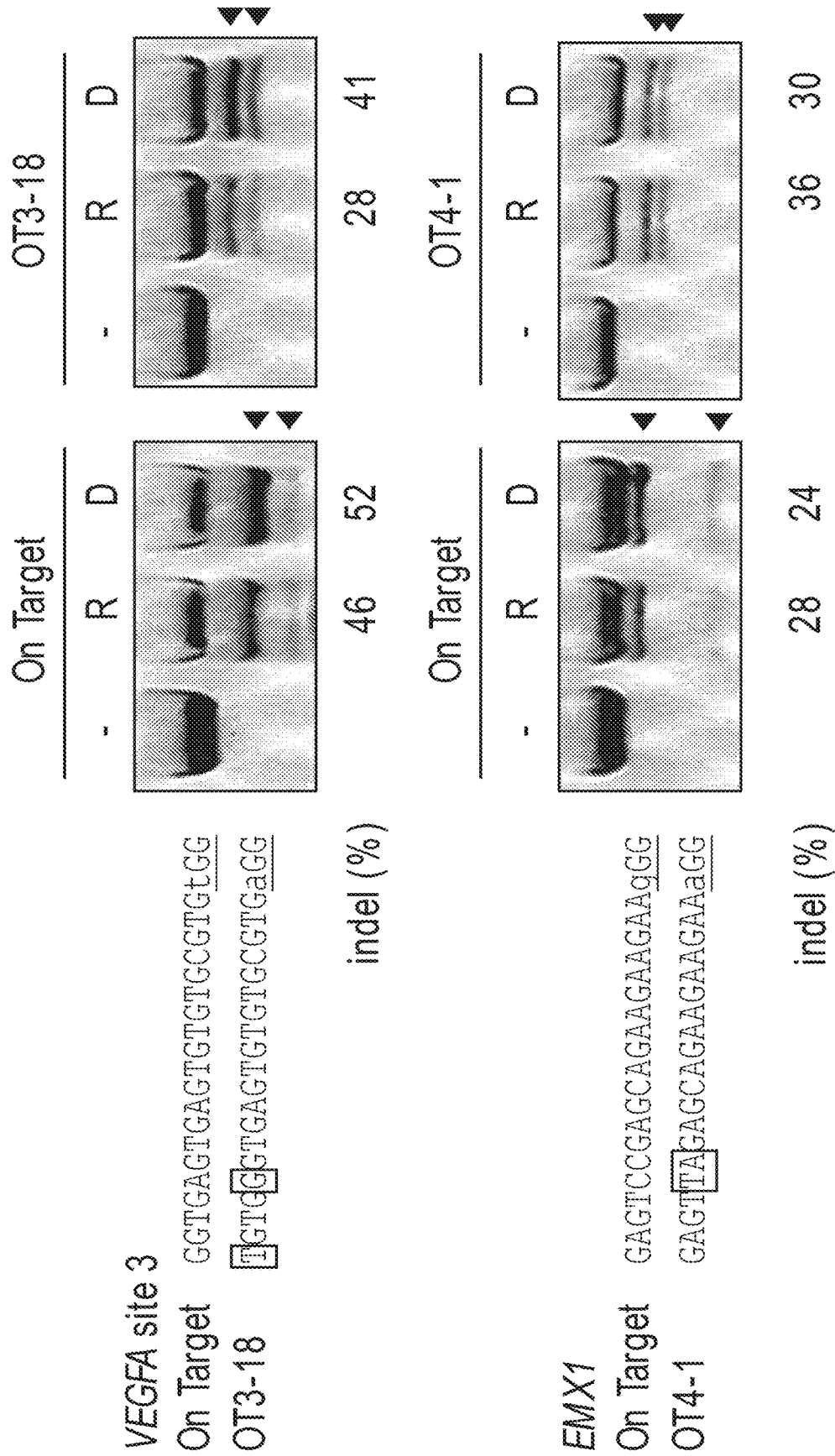


FIG. 14B

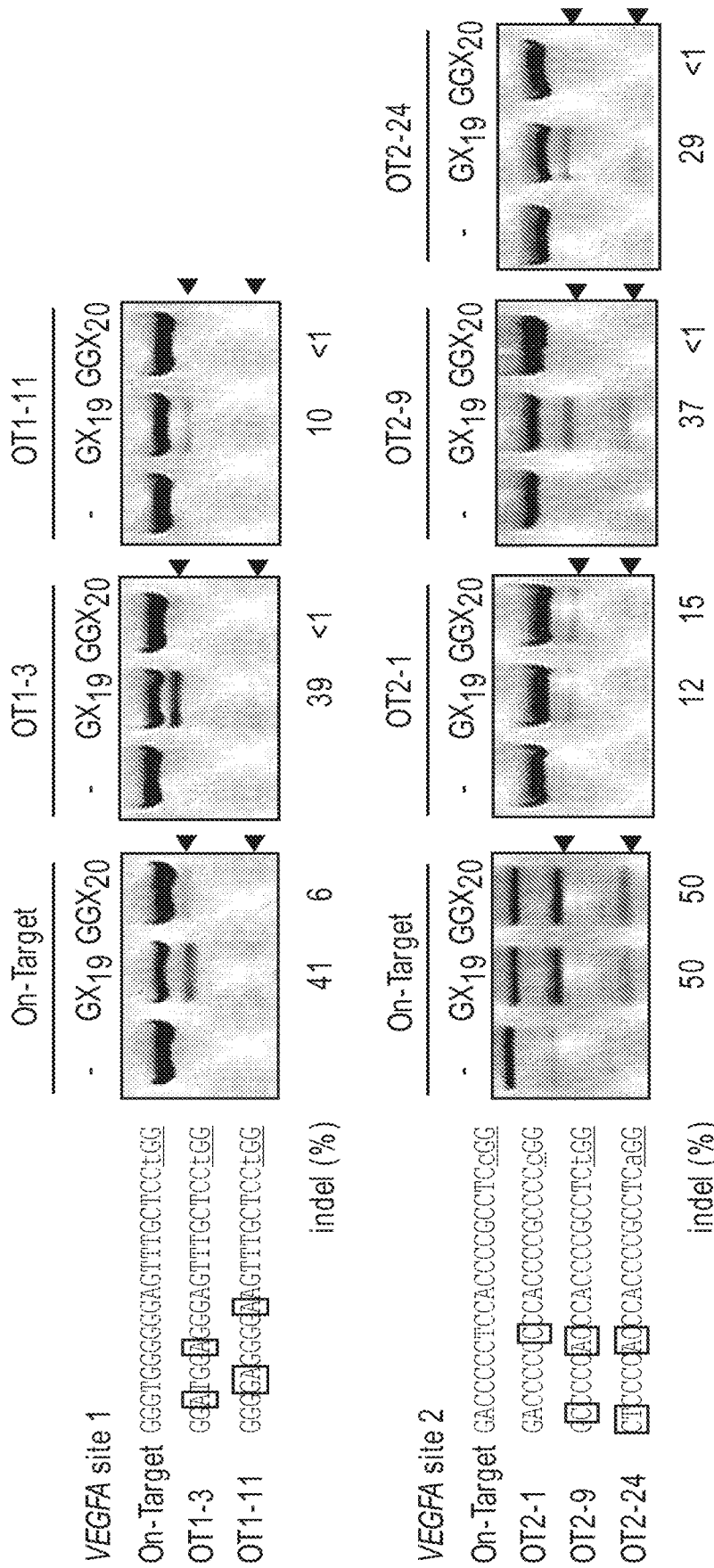


FIG. 15A

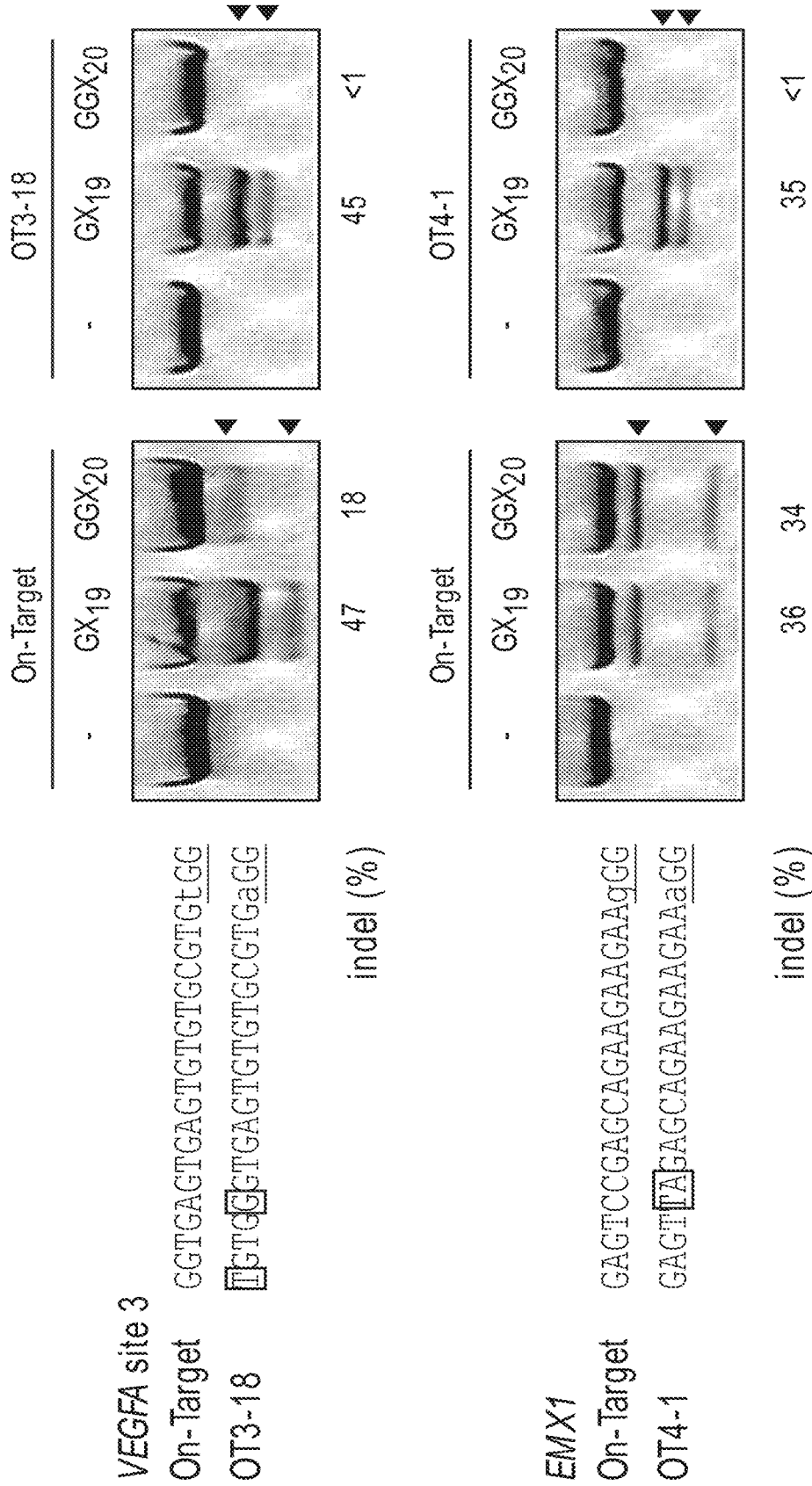


FIG. 15B

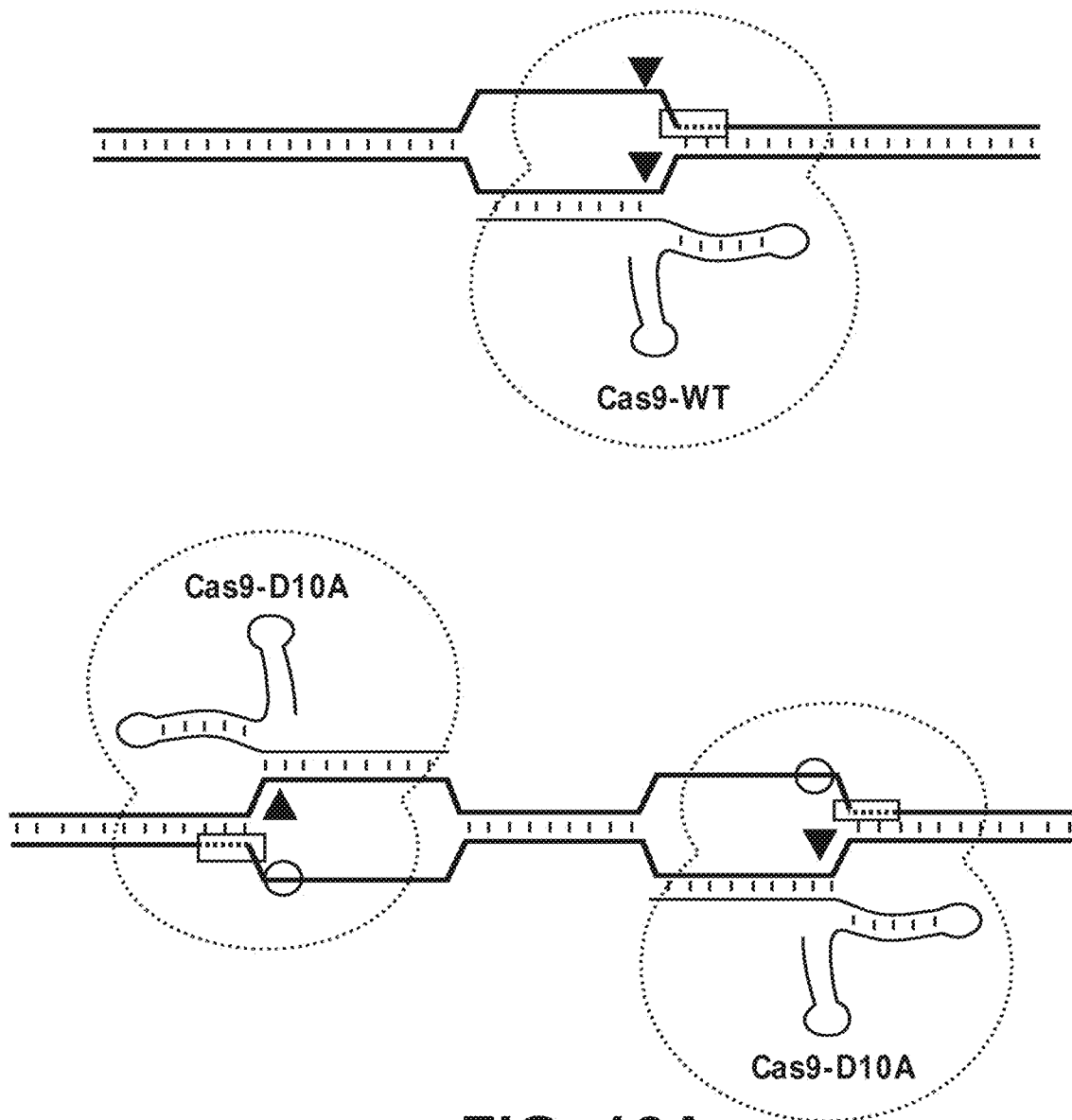


FIG. 16A

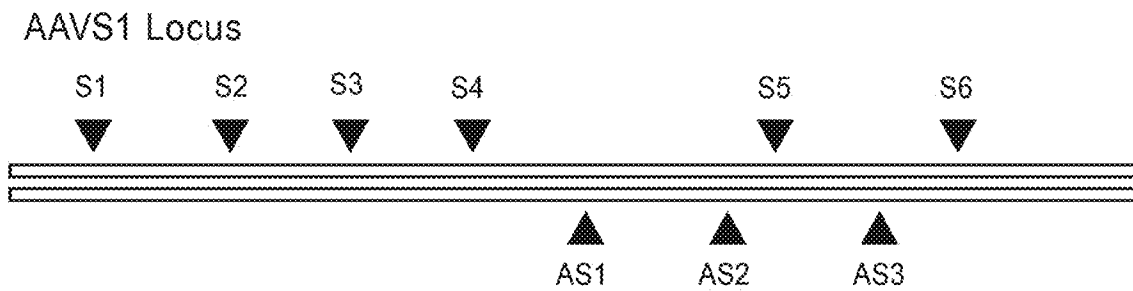


FIG. 16B

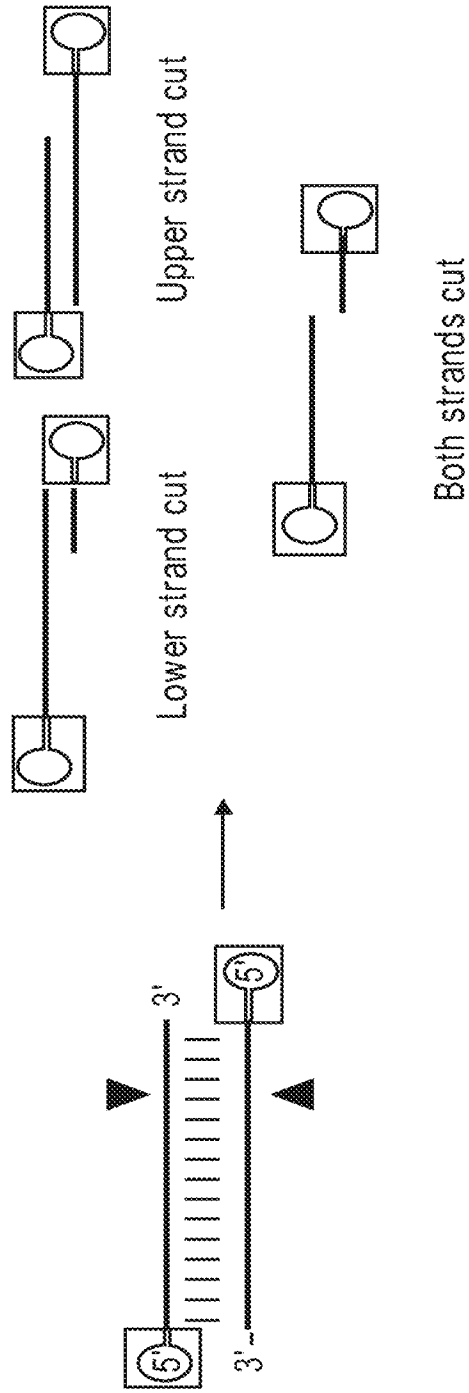


FIG. 16C

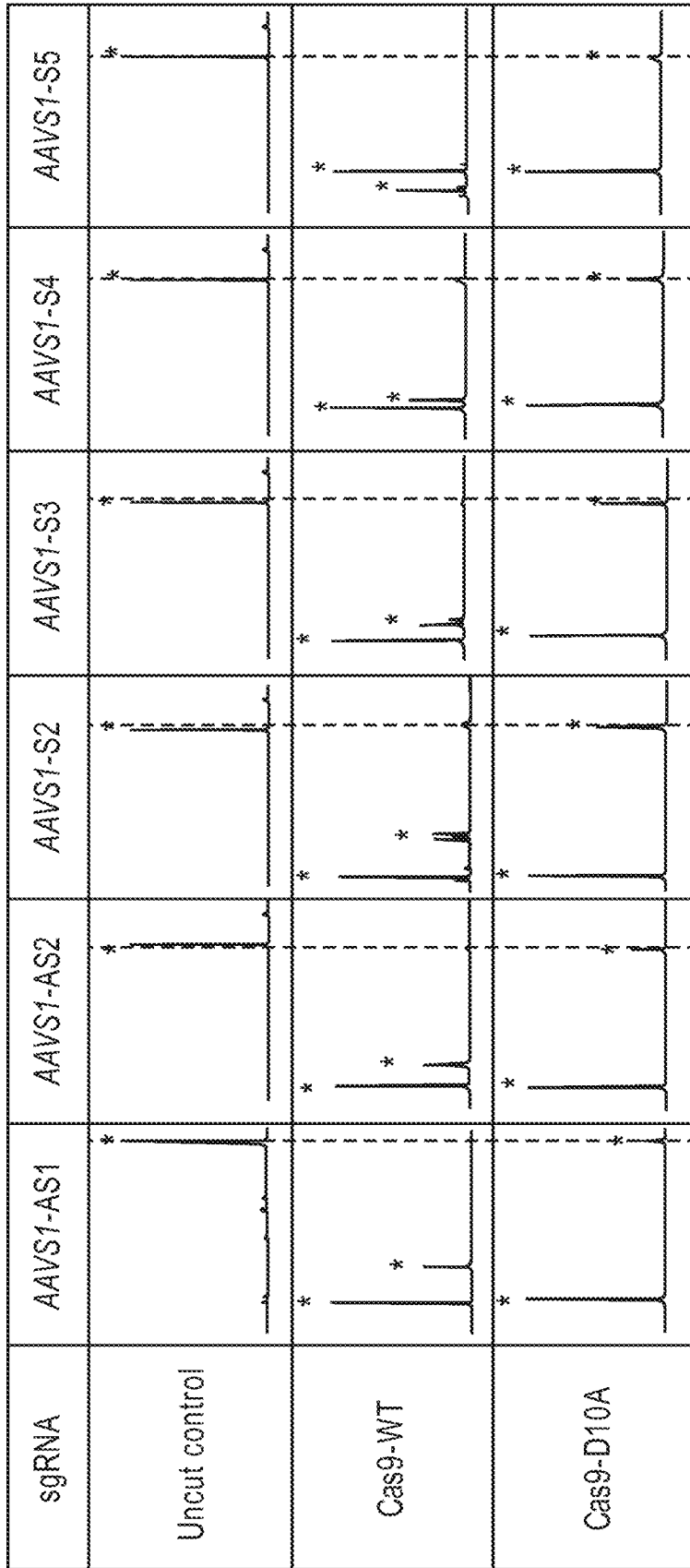


FIG. 16D

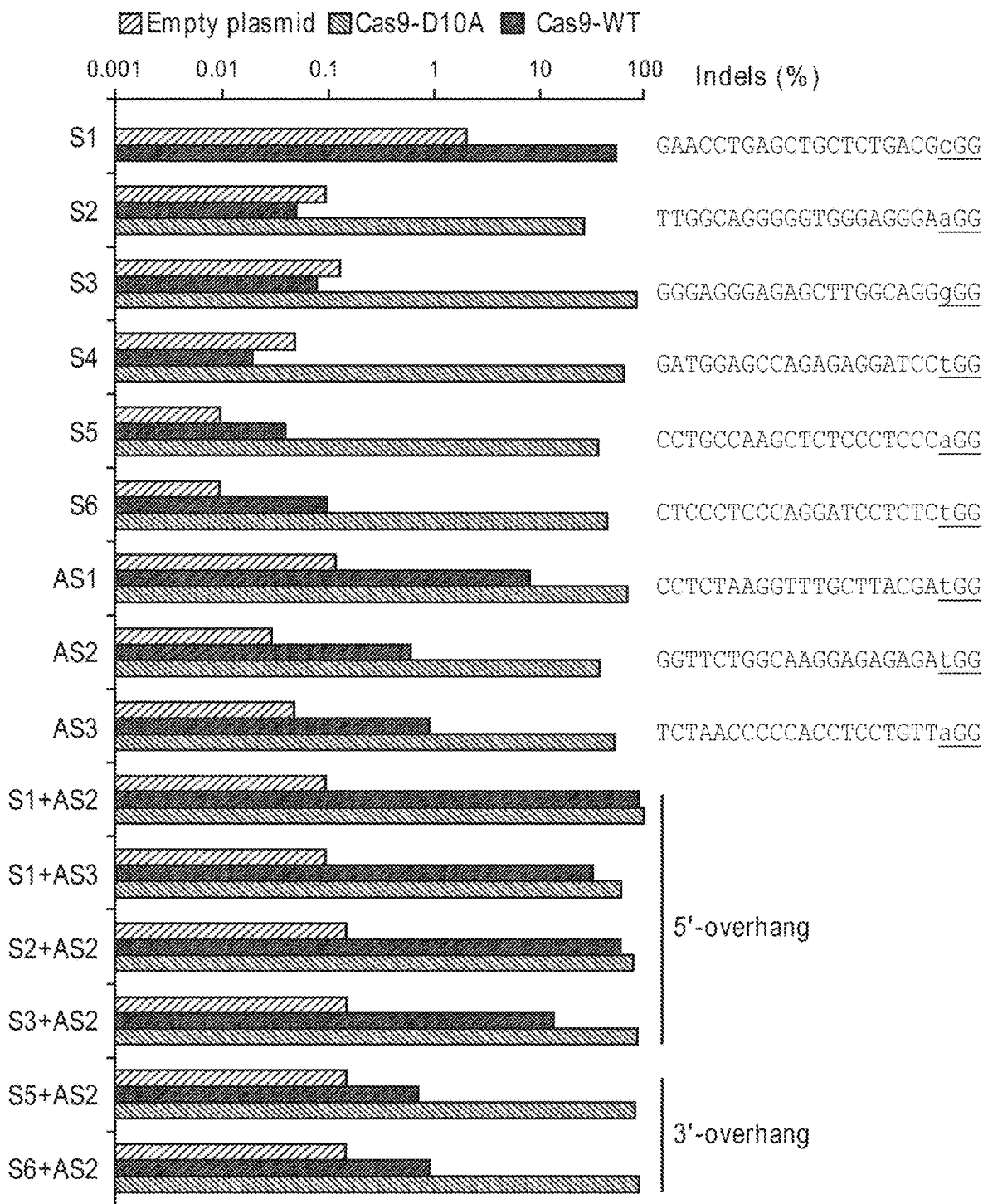


FIG. 17A

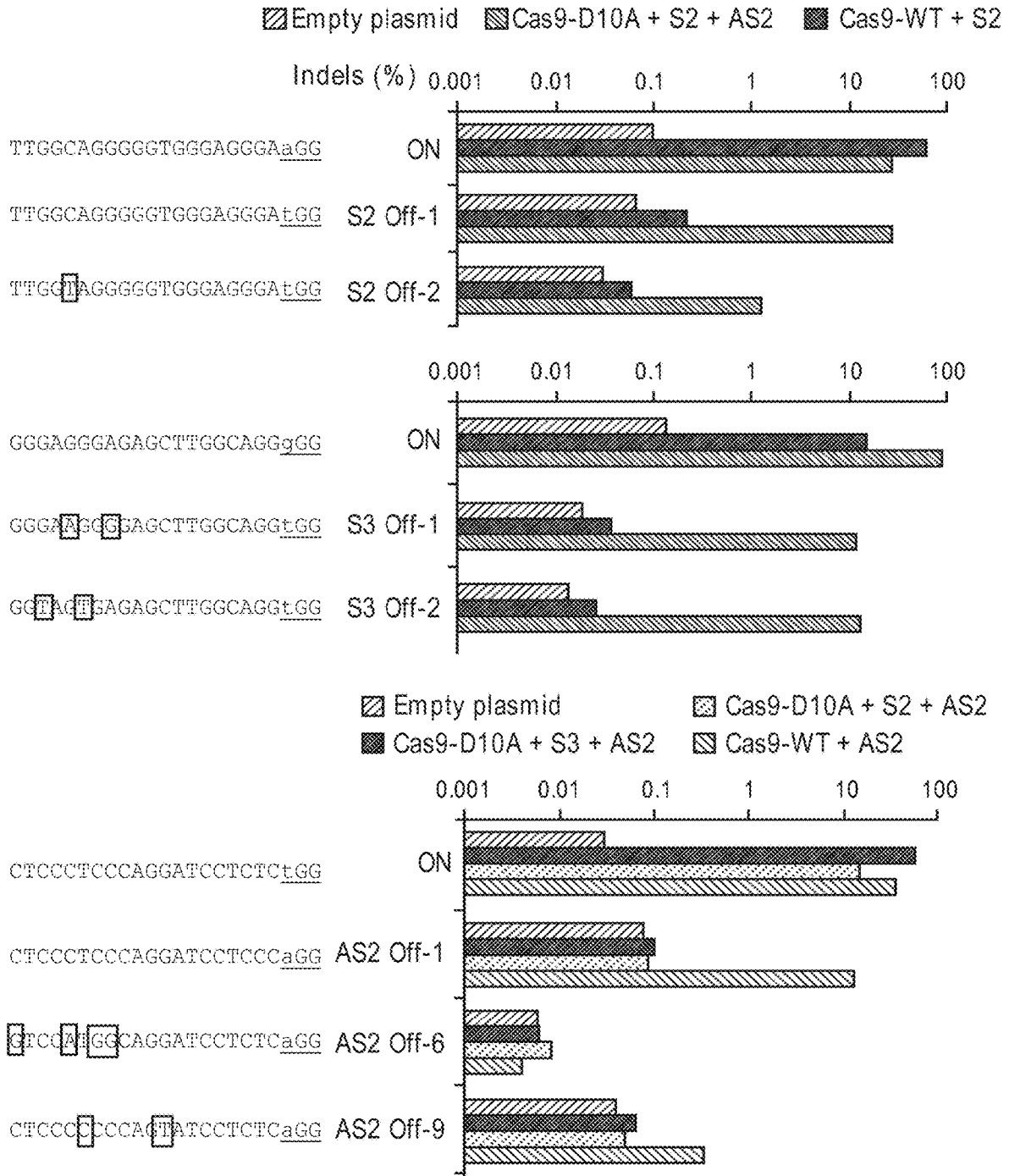


FIG. 17B

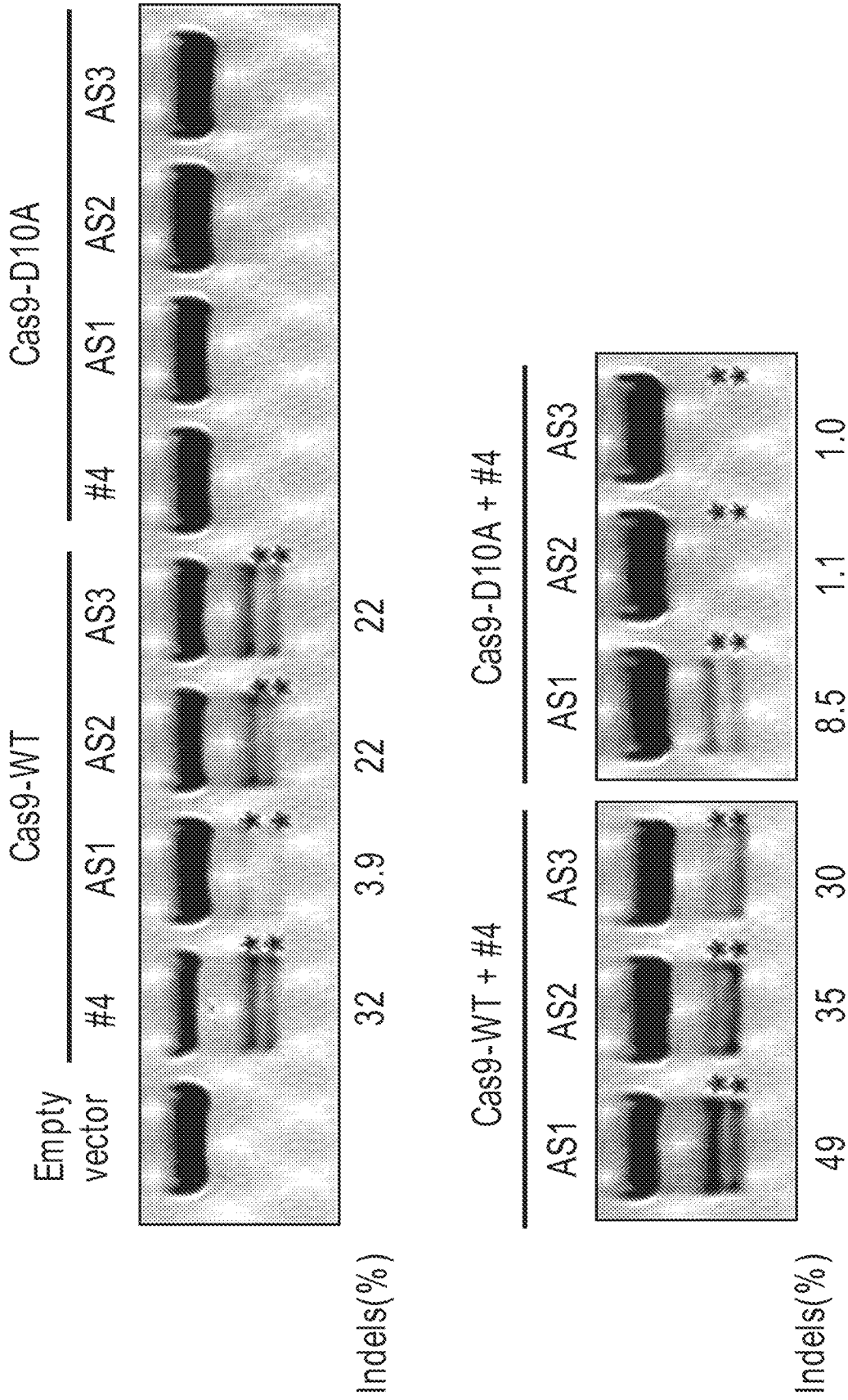


FIG. 18B

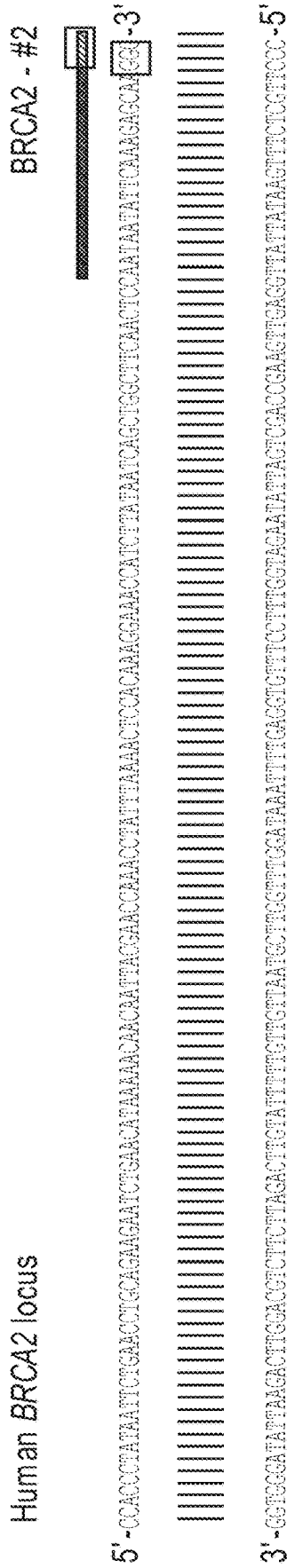


FIG. 18C

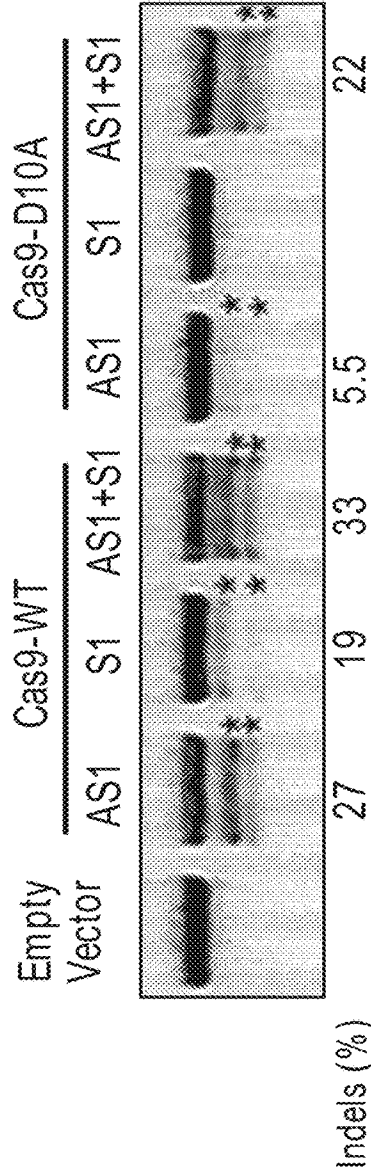


FIG. 18D

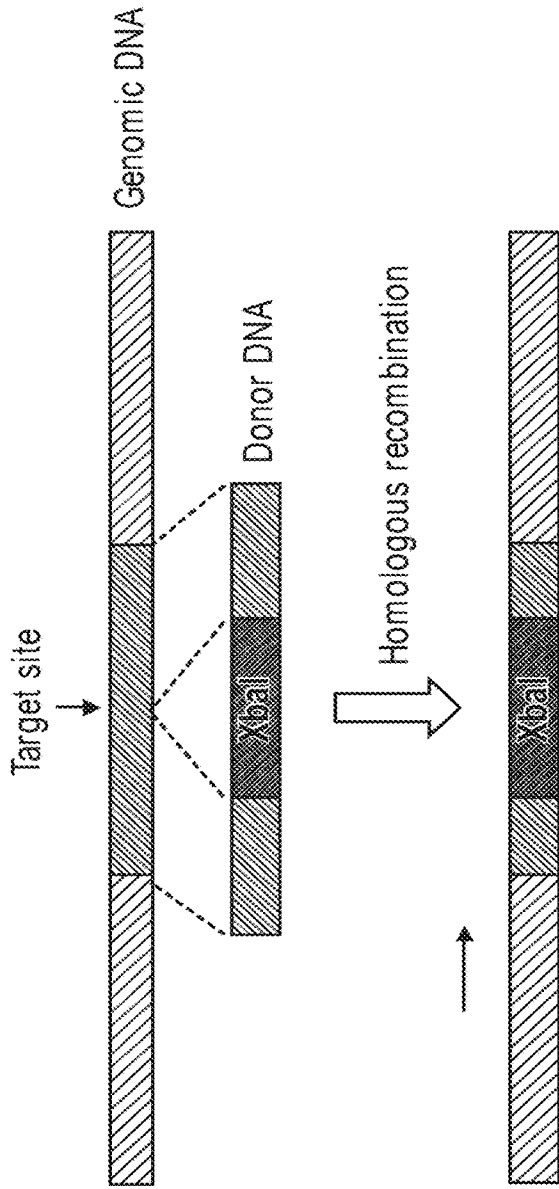


FIG. 19A

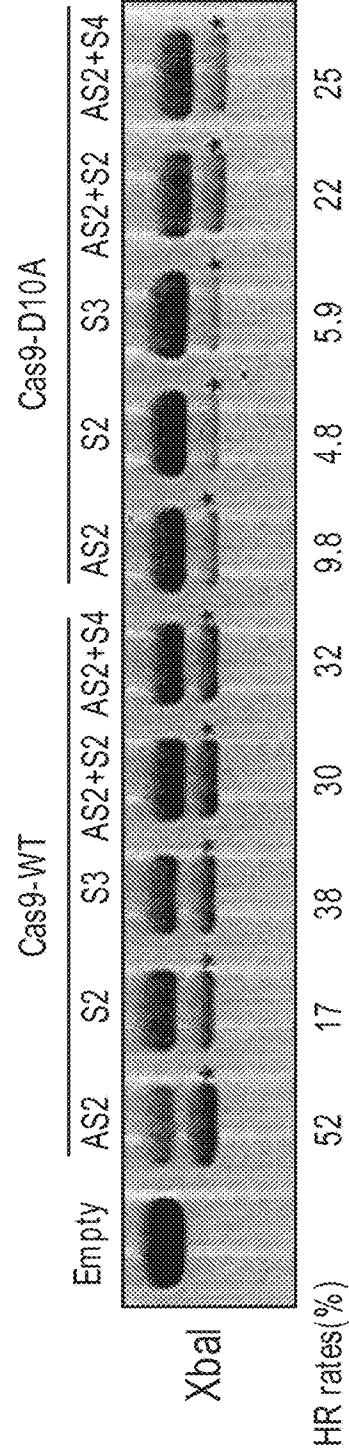


FIG. 19B

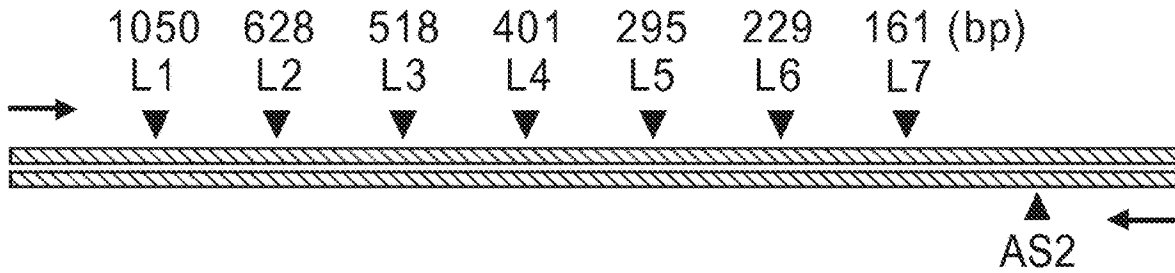


FIG. 20A

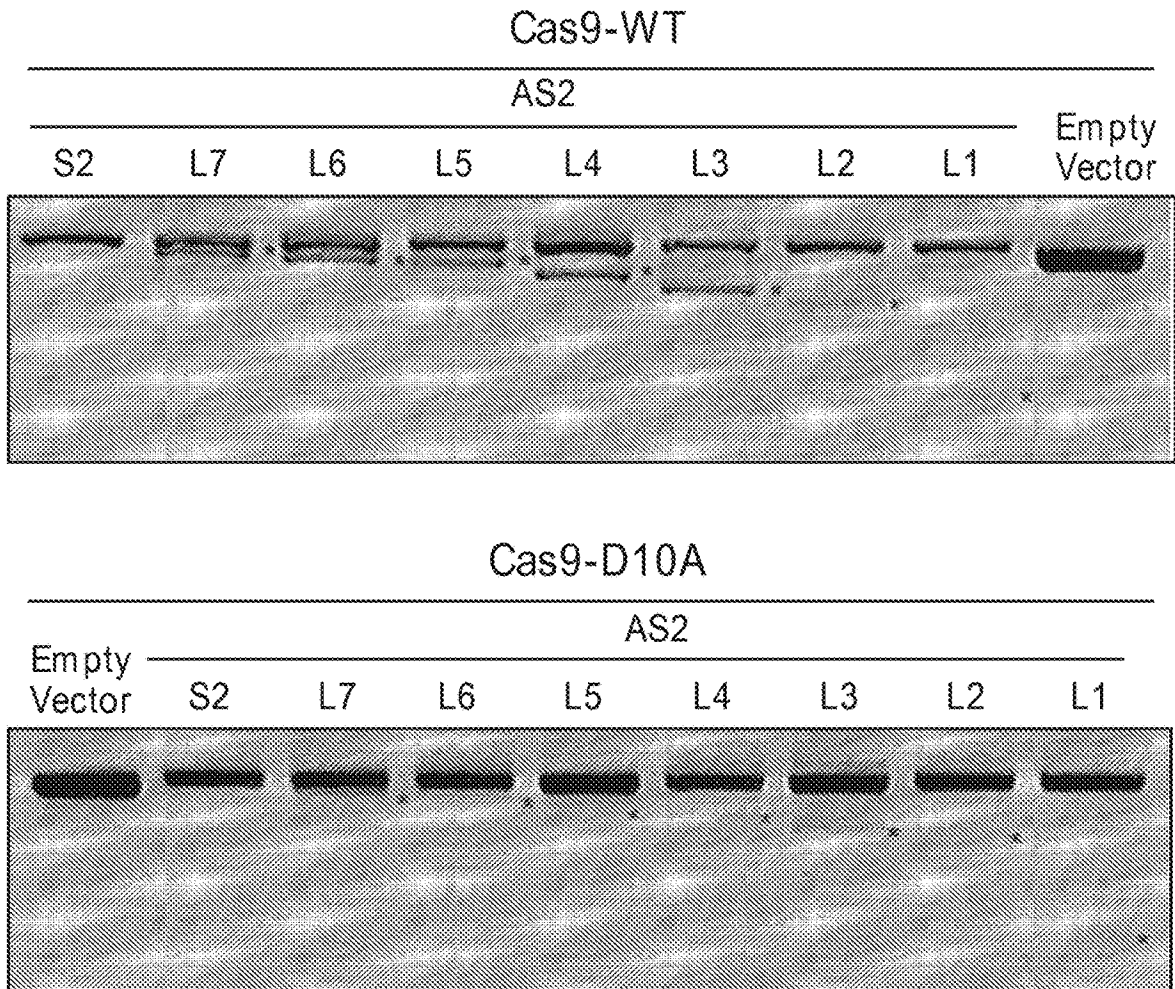


FIG. 20B

Cas9-WT AS2 + L1 (~1050bp deletion)

```

ggccgggaatcaagagctc[CC]AGAGACAGTGCACCAAGCAAGCAAGCCTTt...//...agctctccctcccaagga[CTCTCTG]tccatcgttaagcaaaccttagaggttcttggcaaggagagagatg WT
ggccgggaatcaagagctc[CC]AG-----TGACCAACCAAGCCT-----gtaagcaaaccttagaggttcttggcaaggagagagatg
ggccgggaatcaagagctc[CC]AG-----gaa
ggccgggaatcaagagctc[CC]AGAA-----CCCTCTG]tccatcgttaagcaaaccttagaggttcttggcaaggagagagatg
ggccgggaatcaagagctc[CC]-----taacag
ggccgggaatcaaga-----cgt[CTG]tccatcgttaagcaaaccttagaggttcttggcaaggagagagatg
gg-----tccatcgttaagcaaaccttagaggttcttggcaaggagagagatg
ggccgggaatcaagagctc[CC]AGAA-----CTCTCTG]tccatcgttaagcaaaccttagaggttcttggcaaggagagagatg

```

Cas9-D10A AS2 + L1 (~1050bp deletion)

```

ggccgggaatcaagagctc[CC]AGAGACAGTGCACCAAGCAAGCCTTt...//...agctctccctcccaagga[CTCTCTG]tccatcgttaagcaaaccttagaggttcttggcaaggagagagatg WT
ggccgggaatcaagagctc[CC]AGAGACAGTGCACCAAGCAAGC-----gtaagcaaaccttagaggttcttggcaaggagagagatg
gg-----tccatcgttaagcaaaccttagaggttcttggcaaggagagagatg
ggccgggaatcaagagctc[CC]A-----tctccatcgttaagcaaaccttagaggttcttggcaaggagagagatg
ggccgggaatcaagagctc[CC]AGAA-----CT[CTG]tccatcgttaagcaaaccttagaggttcttggcaaggagagagatg
gg-----tccatcgttaagcaaaccttagaggttcttggcaaggagagagatg
ggccgggaatcaagagctc[CC]AGAGACAGTGCACCAAGCAAGC-----atata
ggccgggaatcaagagctc-----tccatcgttaagcaaaccttagaggttcttggcaaggagagagatg X2

```

FIG. 20C

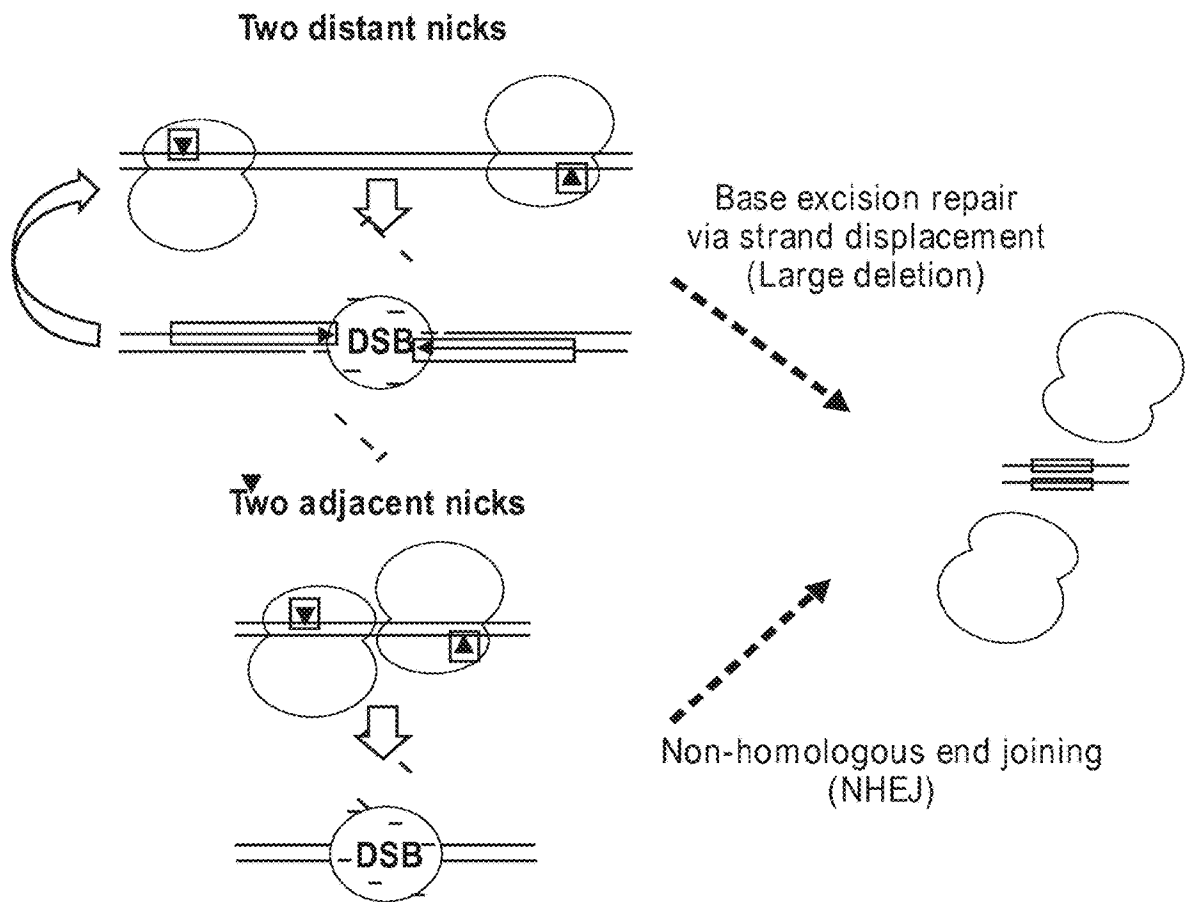


FIG. 20D

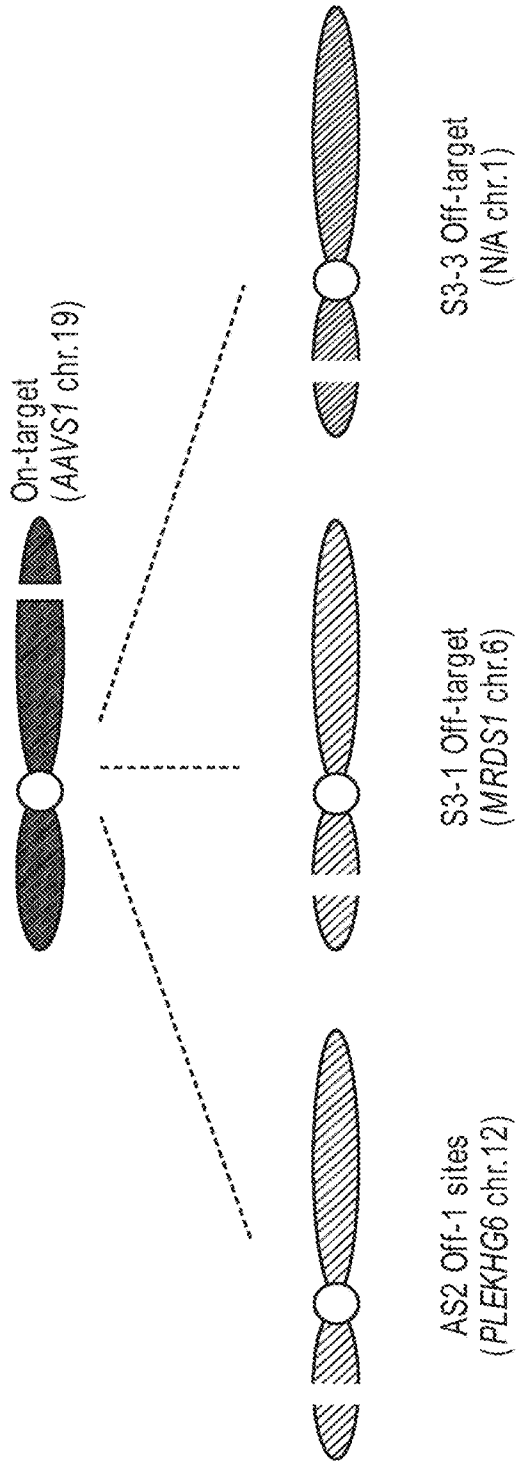


FIG. 21A

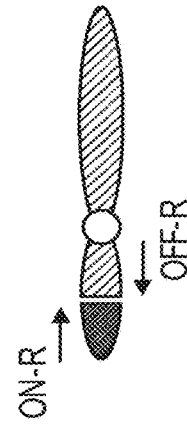
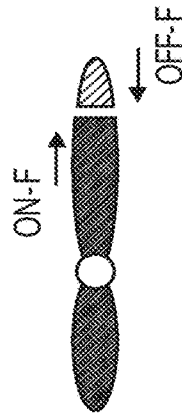


FIG. 21B

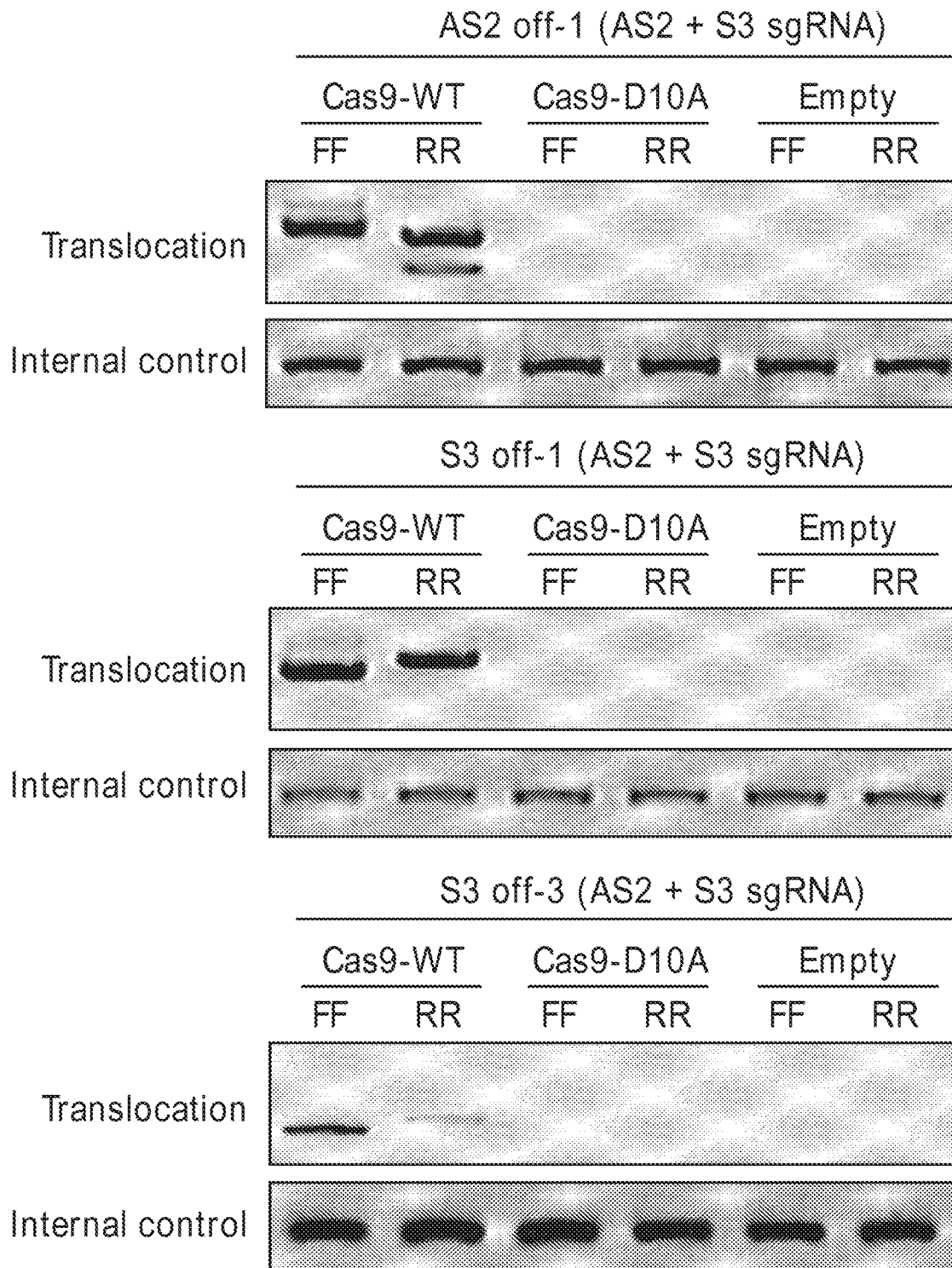


FIG. 21C

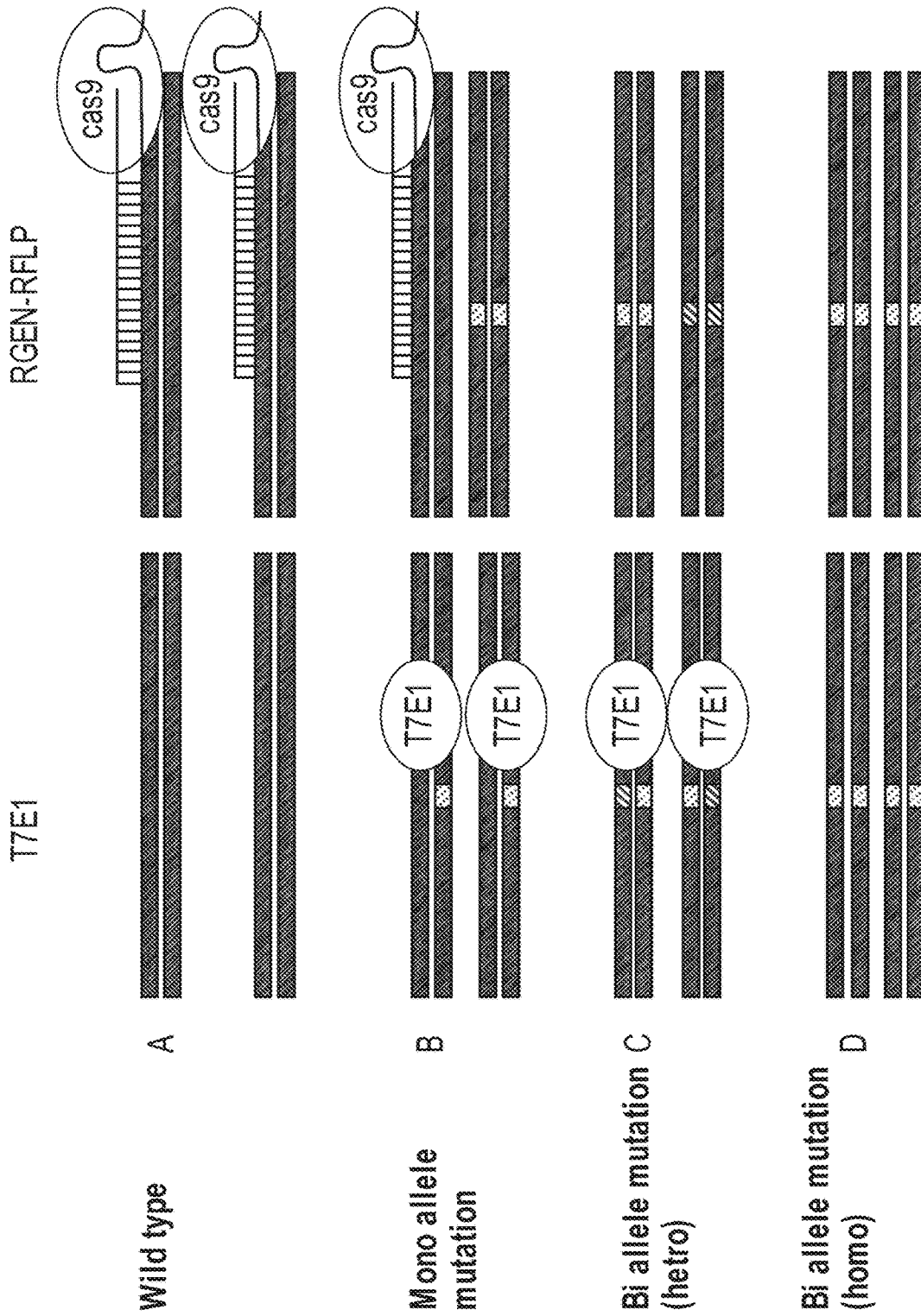


FIG. 22A

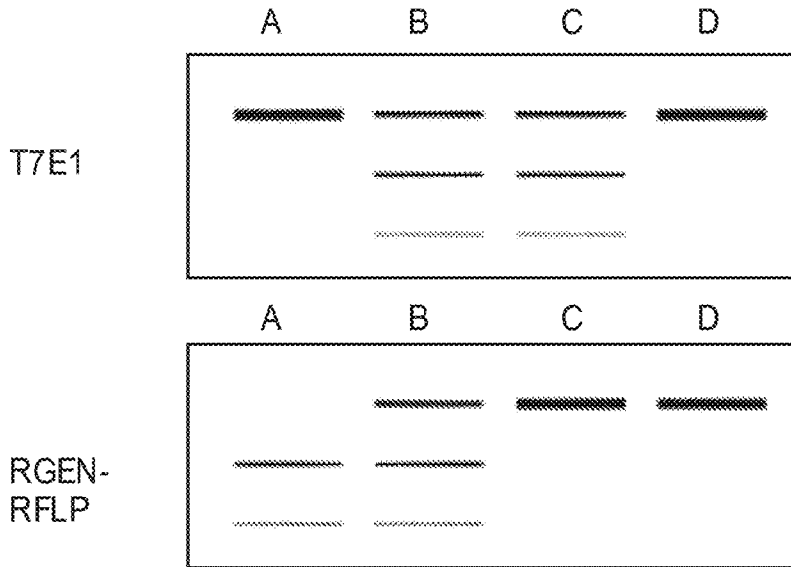


FIG. 22B

Plasmid target sequence

| | |
|---------------------------------------|----|
| AATGACCACTACATCCT---CAAGGG | WT |
| AATGACCACTACATCCT <u>T</u> ---CAAGGG | I1 |
| AATGACCACTACATCCT <u>TT</u> ---CAAGGG | I2 |
| AATGACCACTACATCCT <u>TTT</u> CAAGGG | I3 |
| AATGACCACTACATCCT-----AAGGG | D1 |
| AATGACCACTACATCCT-----AGGG | D2 |
| AATGACCACTACATCCT-----GGG | D3 |

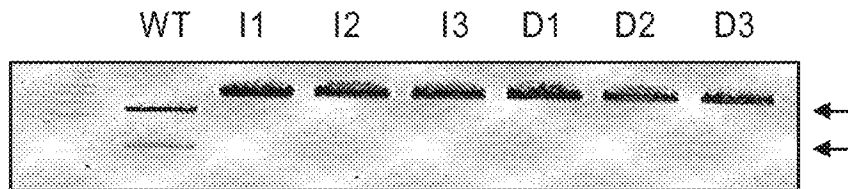


FIG. 23

#1 (+/-)

TATGTGCAATGACCACTACATCCT---CAAAGGGCAGCAATCGGAG WT
TATGTGCAATGACCACTACATCCTCCTCAAAGGGCAGCAATCGGAG +3

#2 (+/-)

TATGTGCAATGACCACTACATCCTCAAAGGGCAGCAATCGGAG WT
TATGTGCAATGACCACTACATC-----AATCGGAG -12

#5 (+/-)

TATGTGCAATGACCACTACATCCTCAAAGGGCAGCAATCGGAG WT
TATGTGCAATGACCACTACAT-----CAGCAATCGGAG -9

#6 (+/-)

TATGTGCAATGACCACTACATCCTCAAAGGGCAGCAATCGGAG WT
TATGTGCAATGACCACTACATCC-----AGCAATCGGAG -8

#12 (-/-)

-----CAGCAATCGG -36
TATGTGCAATGACCACTACATCCT-----CAAAGGGCAGCAATCGG +1
TATGTGCAATGACCACTACATCCT-----CAAAGGGCAGCAATCGG +1
TATGTGCAATGACCACTACATCCT/67bp/CAAAGGGCAGCAATCGG +67

#28 (-/-)

TATGTGCAATGACCACTACATCCTCAAAGGGCAGCAATCGG +1
TATGTGCAATGACCACTACAT---|-----GGCAGCAATCGG -7, +1
TATGTGCAATGACCACTACAT|----- -94

FIG. 24A

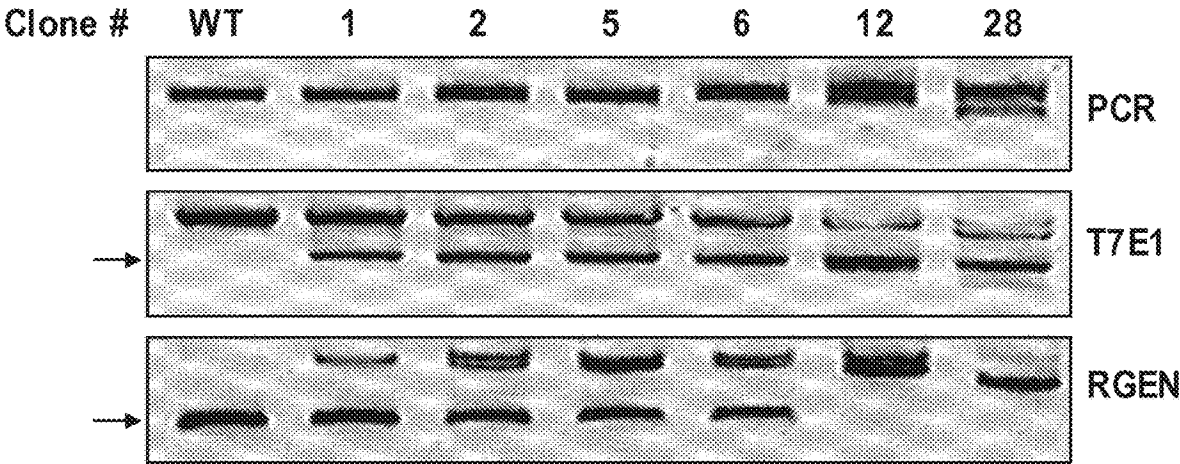


FIG. 24B

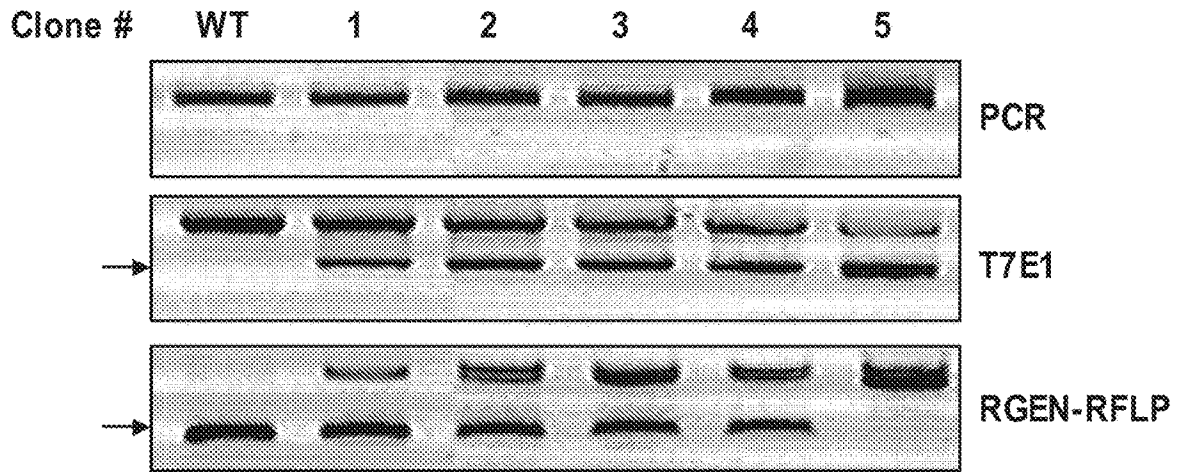


FIG. 25A

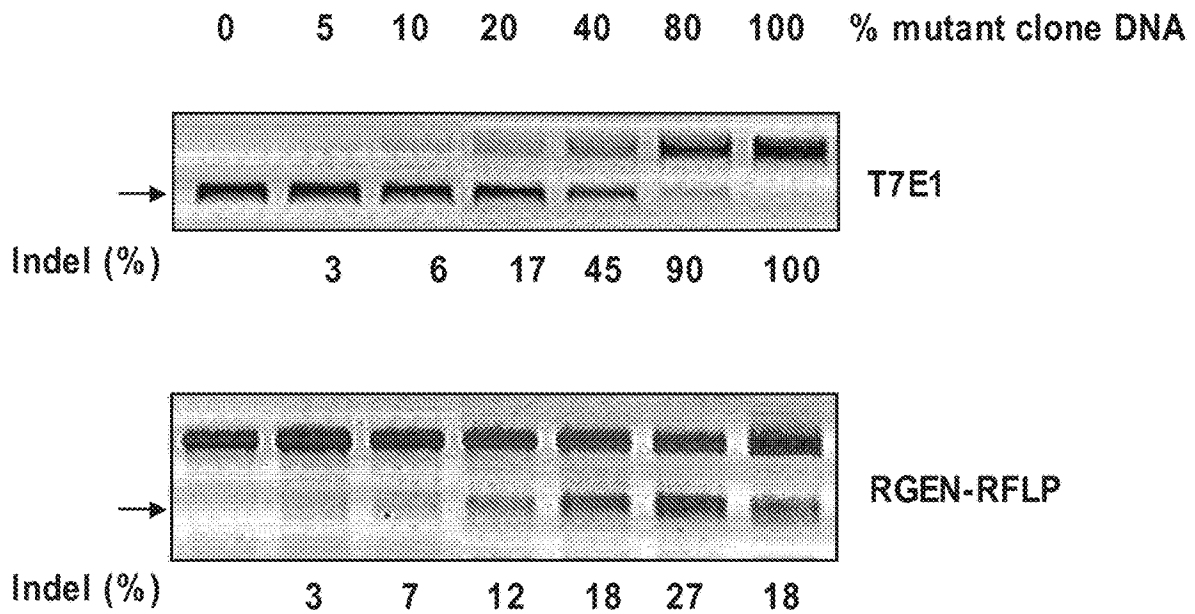


FIG. 25B

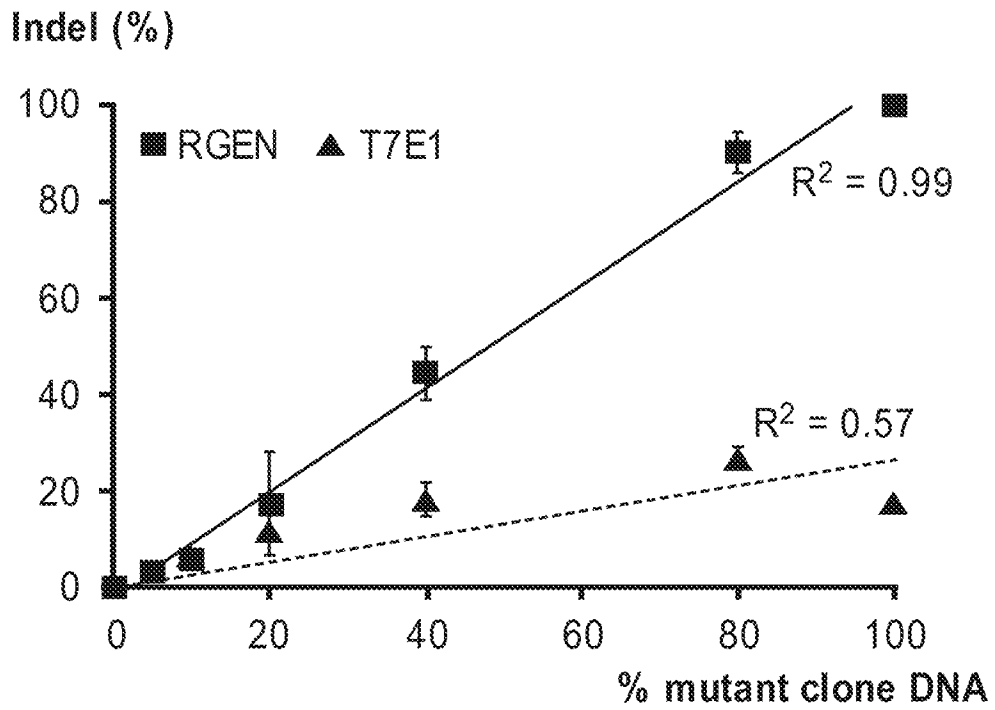
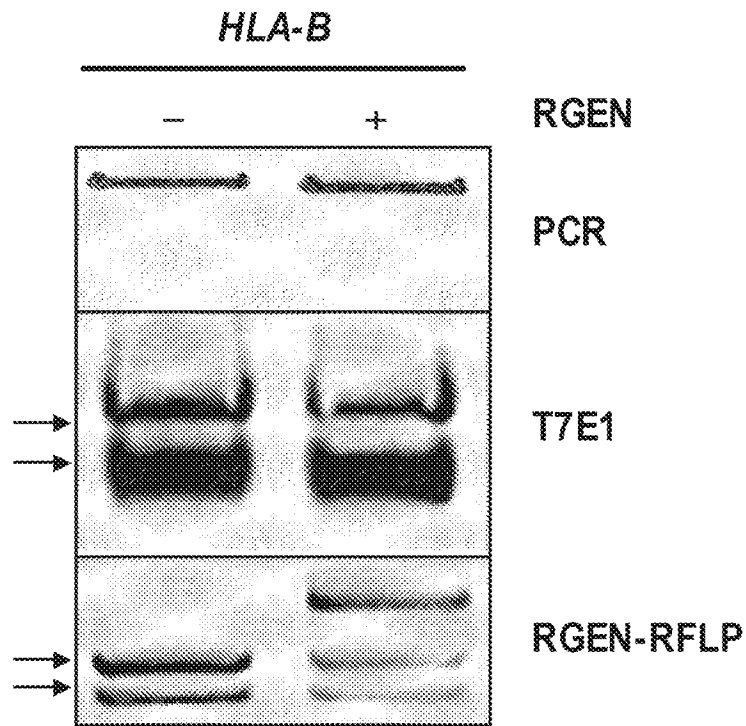


FIG. 25C

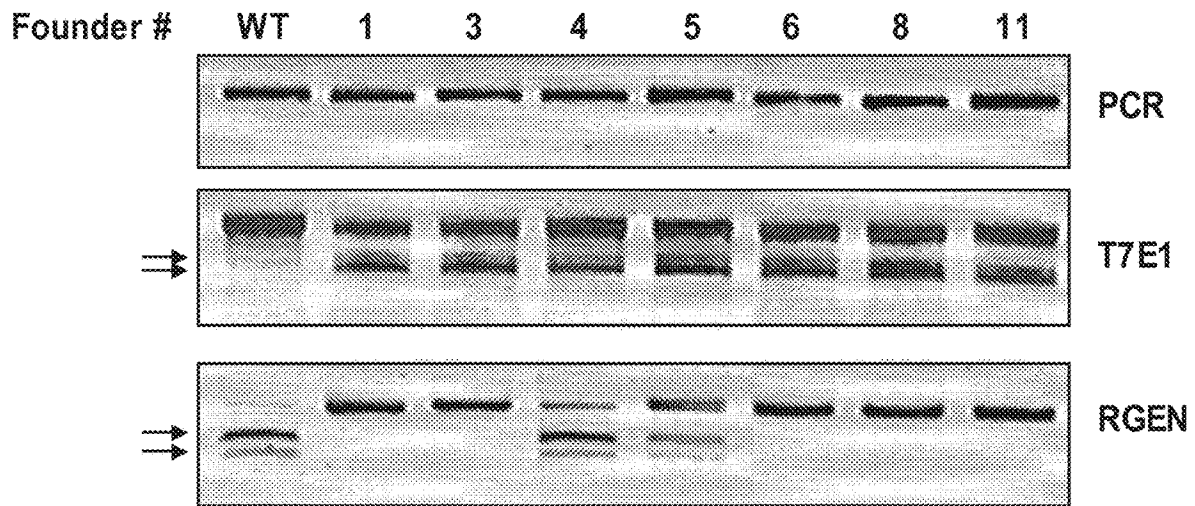


FIG. 26B

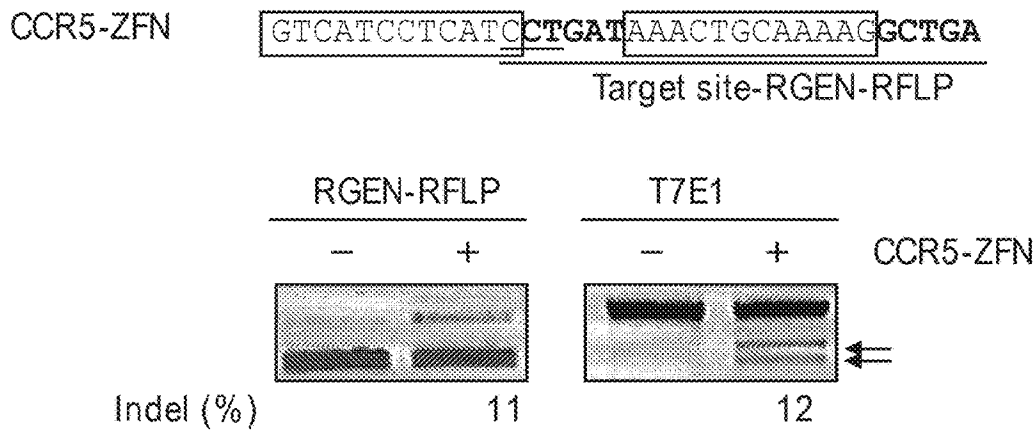


FIG. 27

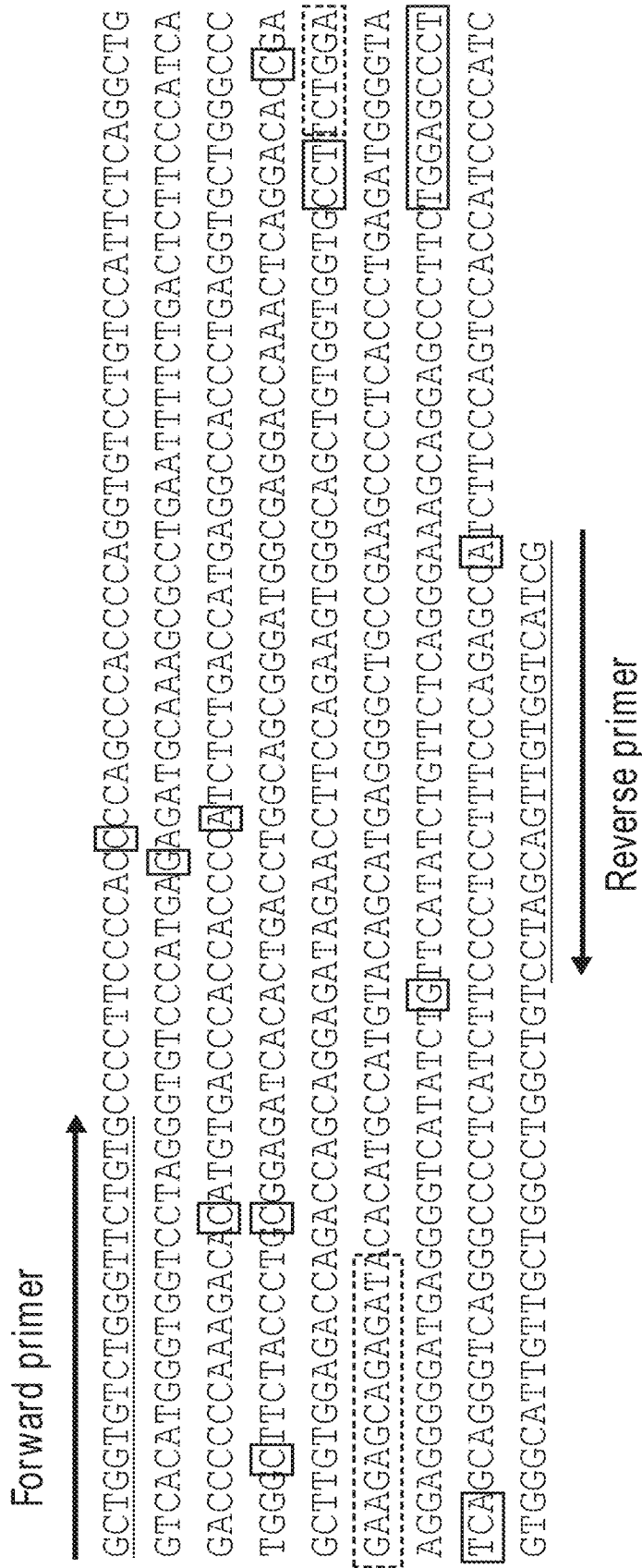
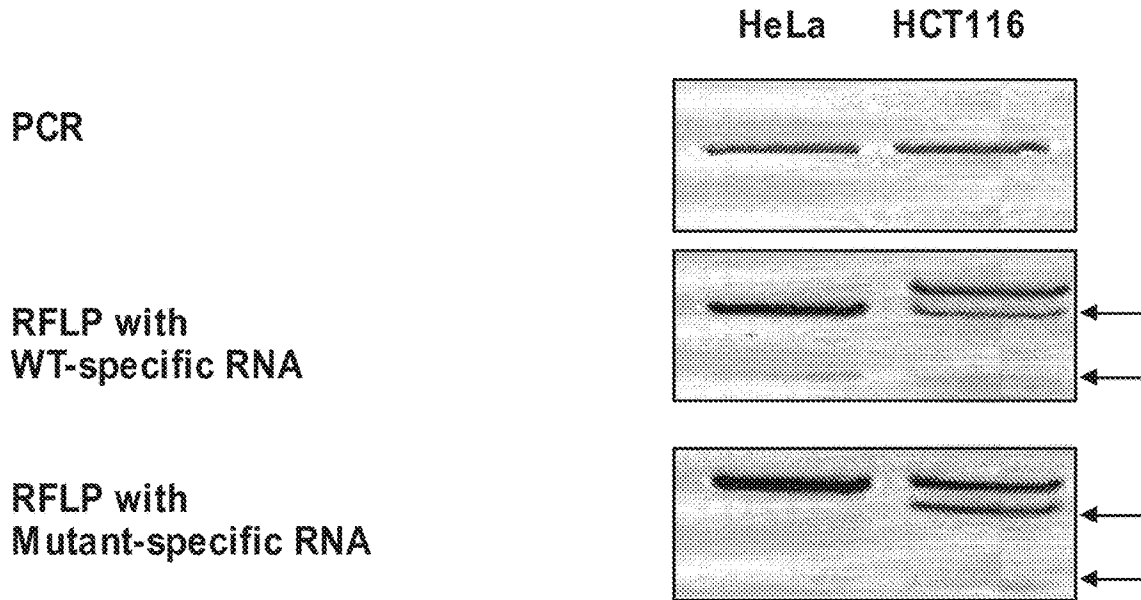


FIG. 28



HeLa

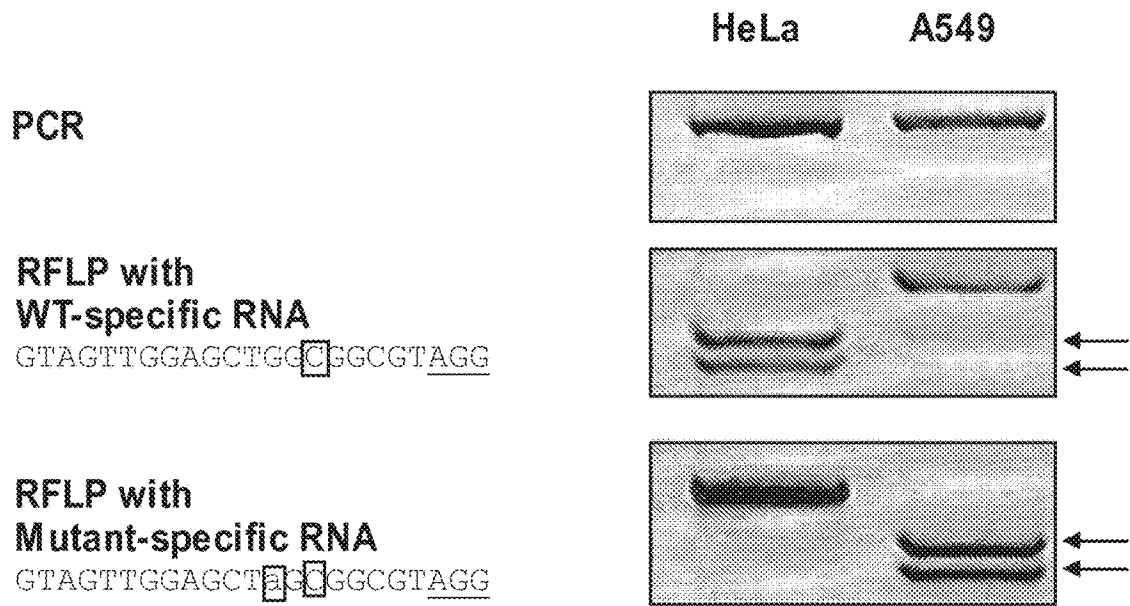
ACTACCACAGCTCCTTCTCTGAGTGG wild-type

HCT116

ACTACCACAGCTCCTTCTCTGAGTGG wild-type

ACTACCACAGCTCCT---CTGAGTGG c.133-135 del TCT

FIG. 29A



HeLa

GTAGTTGGAGCTGGTGGCGTAGG wild-type

A549

GTAGTTGGAGCT**G**GTGGCGTAGG c.34G>A

FIG. 29B

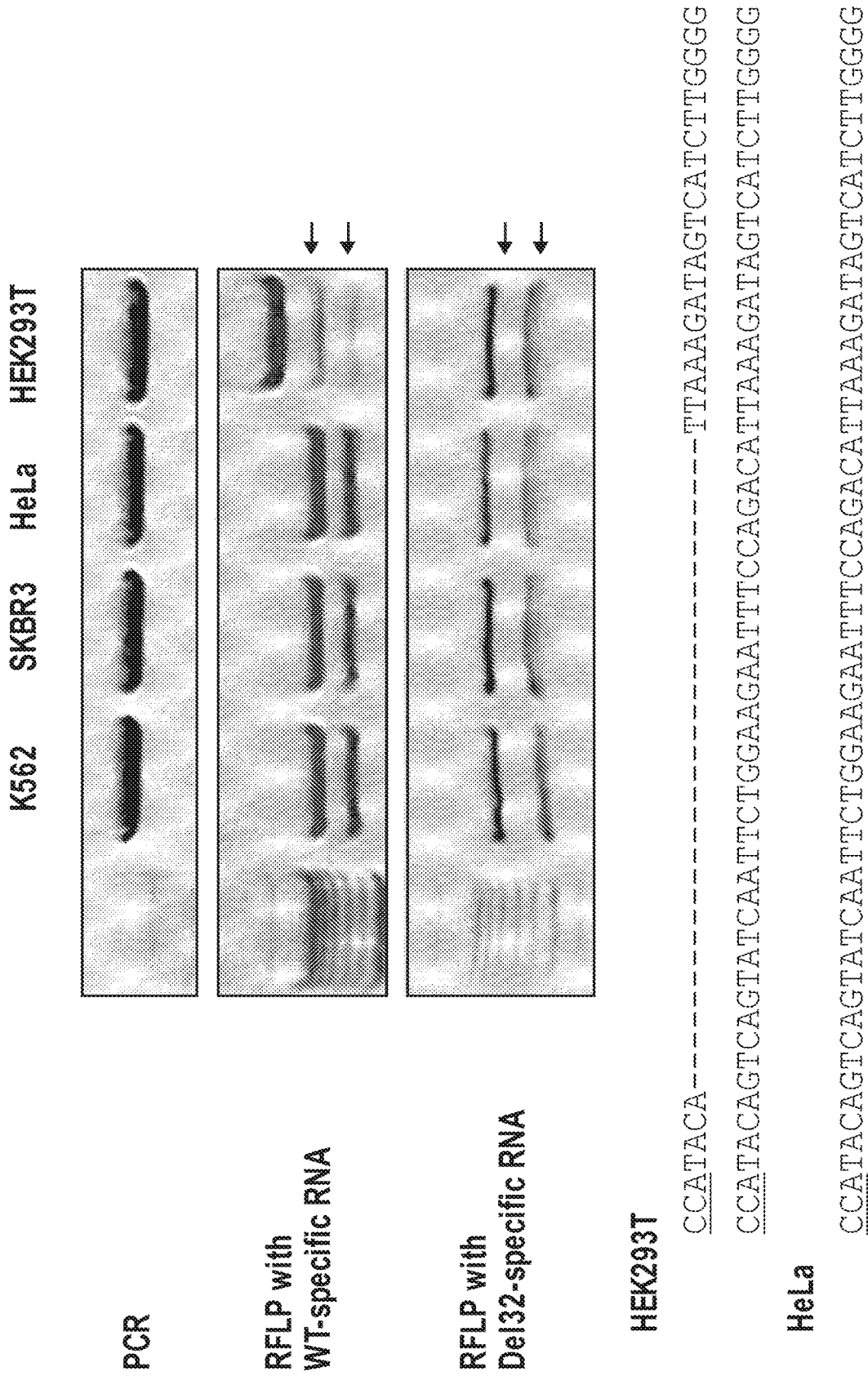


FIG. 30A

CCR5 WT
 ccatacagtcagtatcaattctggaagaatttccagacattaaagatagtcattct

CCR5 WT target site

CCR5 Δ 32
 ccataca-----ttaagatagtcattct

CCR5 Δ 32 on-target site

CCR5 Δ 32 Δ 32 off-target site

FIG. 30B

WT Δ 32 Linearized plasmid

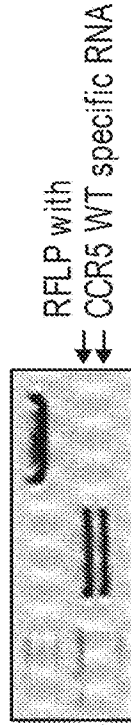
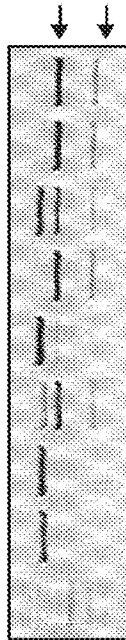


FIG. 30C

181 127 1/9
 on off on off on off on off
 Dilution (Cas9 0.1ug/crRNA 25ng / tracrRNA 60ng)
 Plasmid



On agatgactatctttaatgctcgg
 Off agatgactatctttaatgctcgg

FIG. 30D

KRAS

HeLa

GTAGTTGGAGCTGGTGGCGTAGG Wild-type

A549

GTAGTTGGAGCTaGTGGCGTAGG c.34G>A

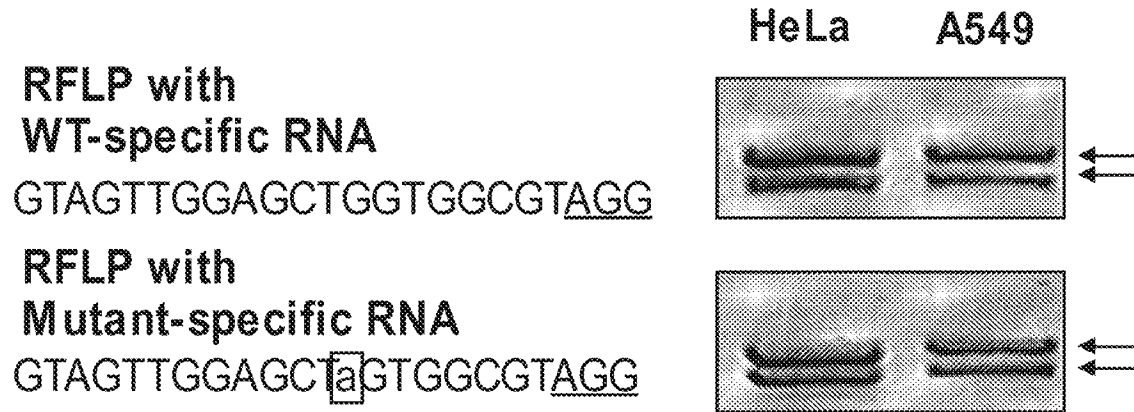


FIG. 31A

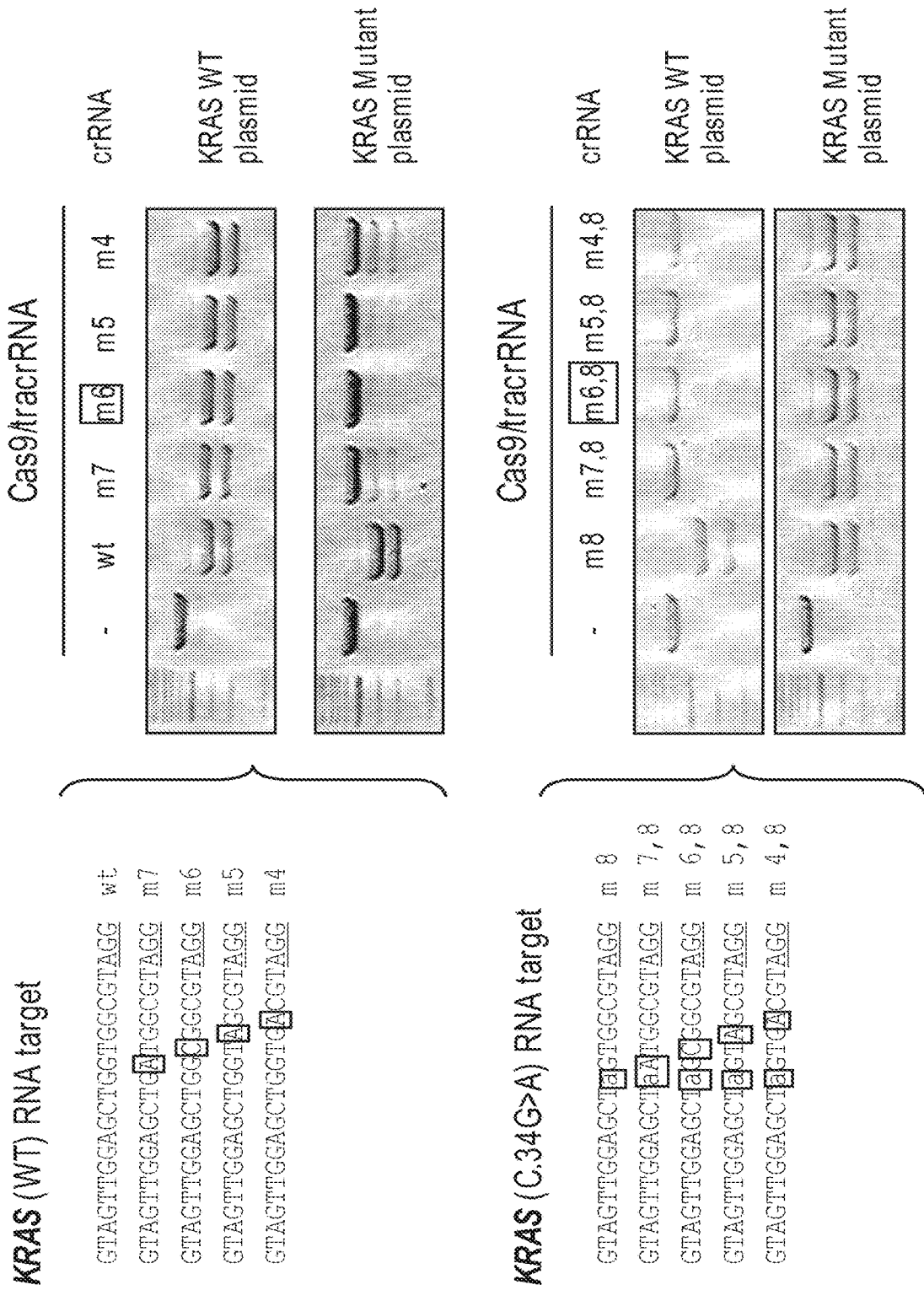


FIG. 31B

PIK3CA

HeLa

CAAATGAATGATGCACATCATGG Wild-type

HCT116

CAAATGAATGATGCACATCATGG Wild-type

CAAATGAATGATGCACAGTCATGG C.3140A>G

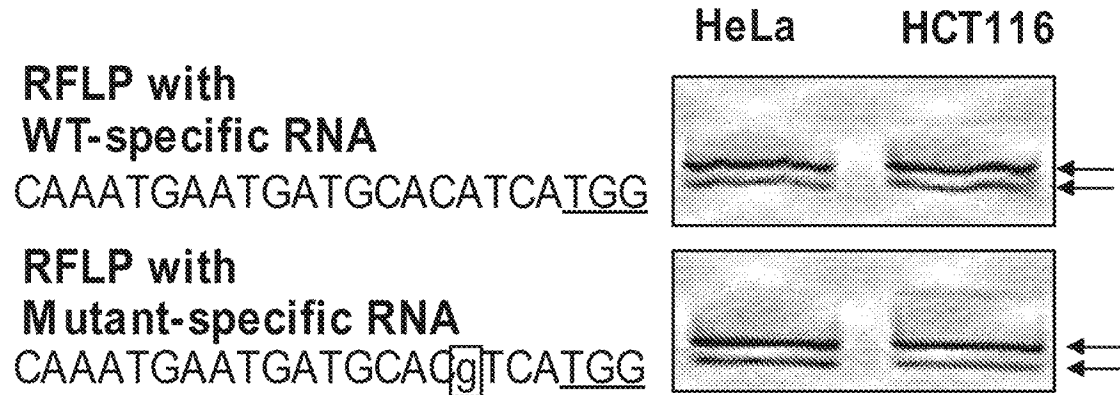


FIG. 32A

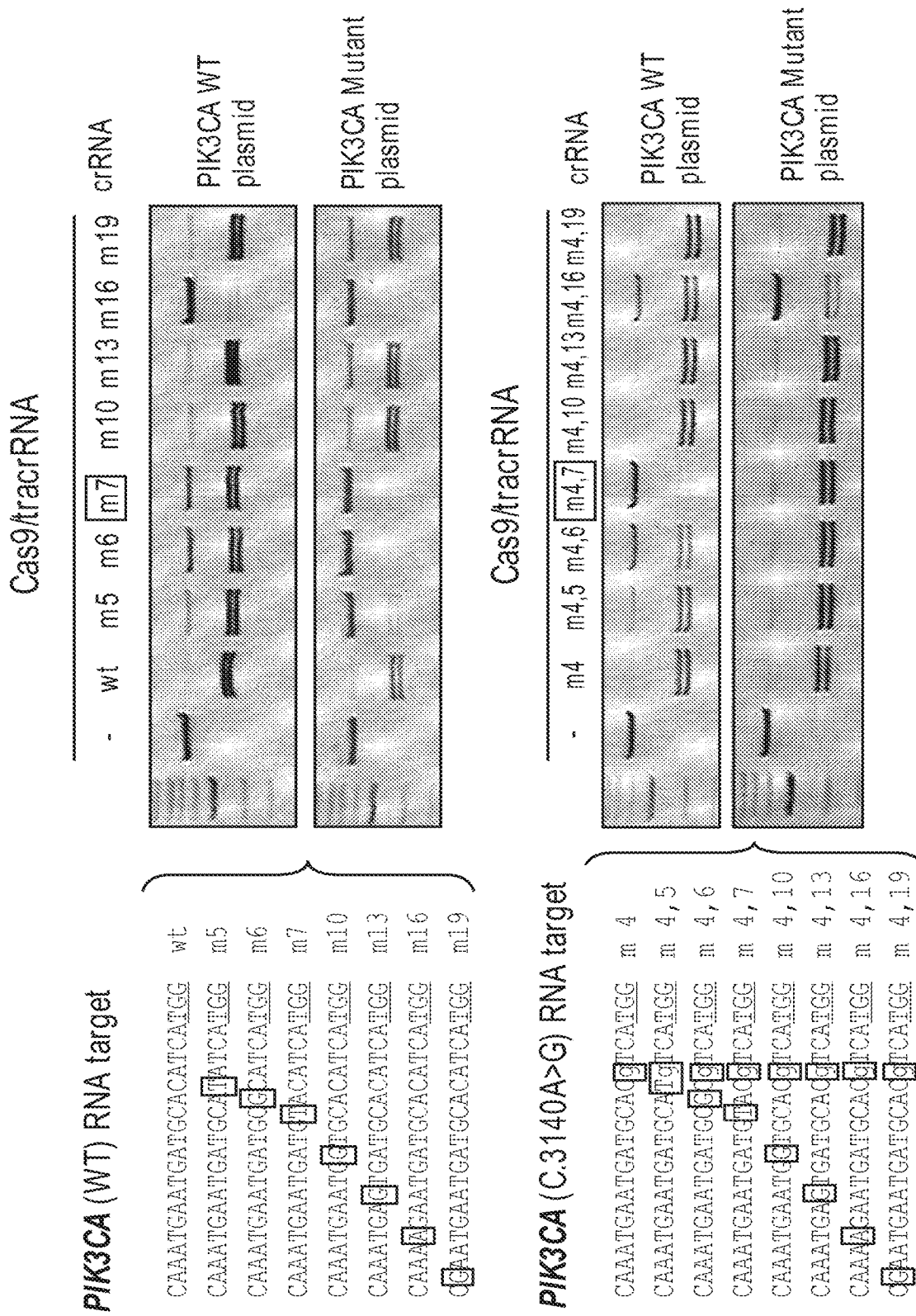


FIG. 32B

HeLa

ATCATAGGTCGTCATGCTTATGG Wild-type

HT1080

ATCATAGGTCGTCATGCTTATGG Wild-typ

ATCATAGGT[]GTCATGCTTATGG c.394C>T

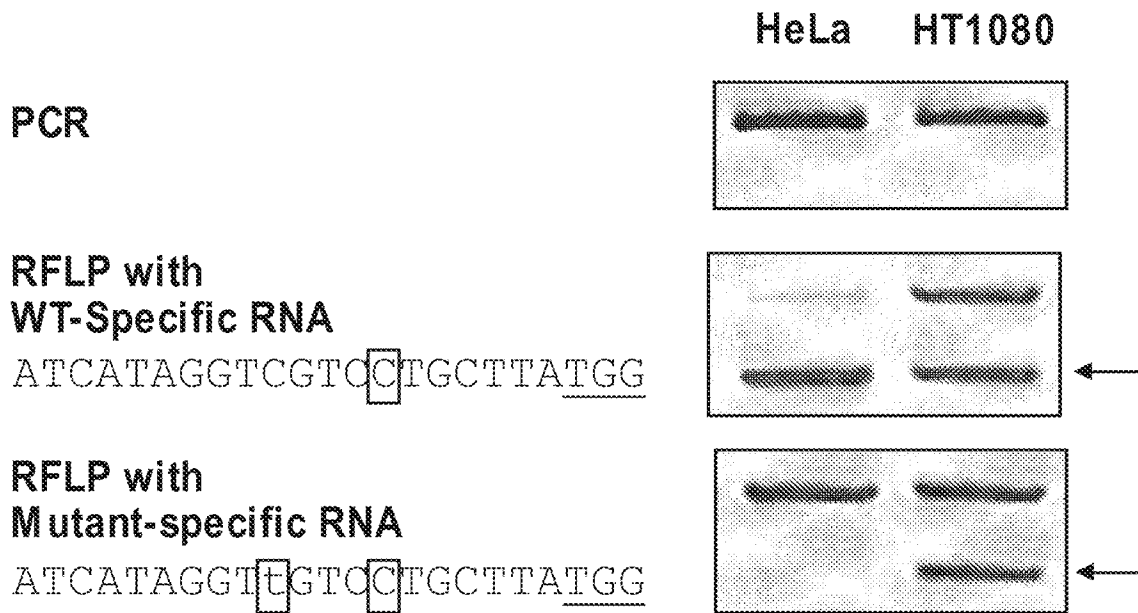


FIG. 33A

PIK3CA

HeLa

CAAATGAATGATGCACATCATGG Wild-type

HCT116

CAAATGAATGATGCACATCATGG Wild-type

CAAATGAATGATGCACgTCATGG C.3140A>G

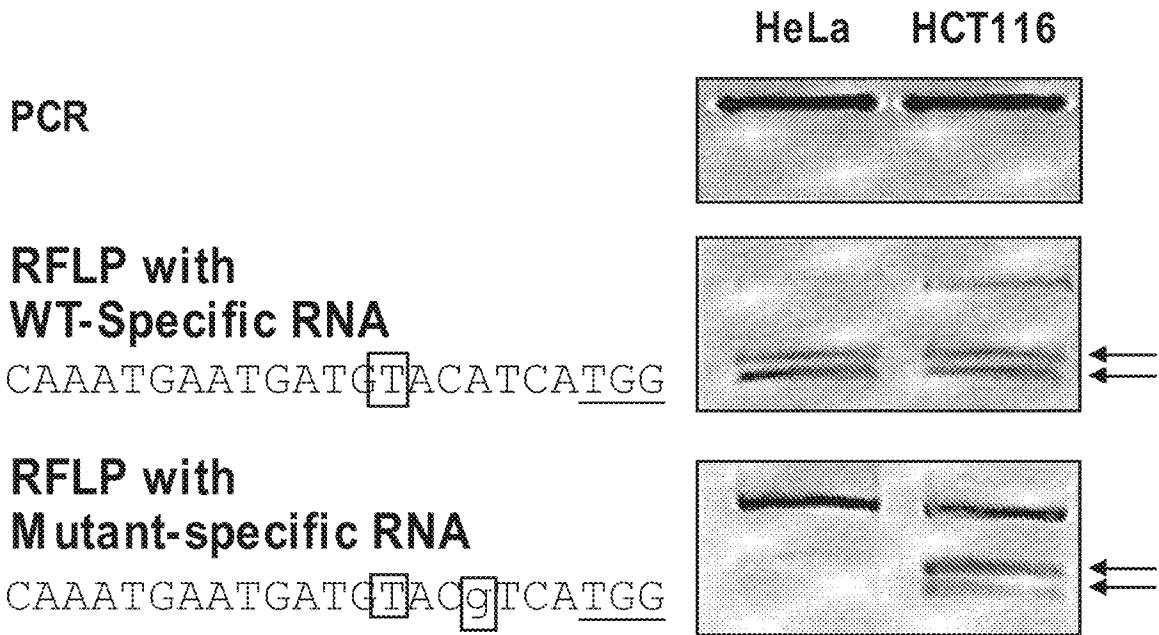


FIG. 33B

NRAS

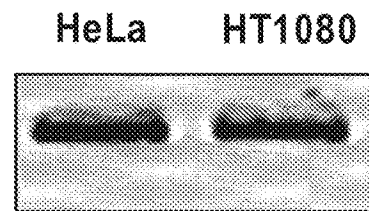
HeLa

CTGGACAAGAAGAGTACAGTGCC Wild-type

HT1080

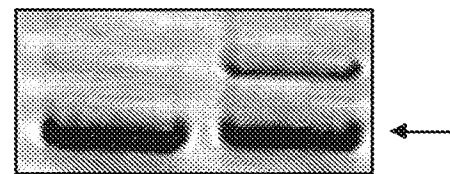
CTGGACAAGAAGAGTACAGTGCC Wild-type
CTGGGA^aAAGAAGAGTACAGTGCC c.181C>A

PCR



**RFLP with
WT-Specific RNA**

CTGGACAAGAAGAGTACAGTGCC



**RFLP with
Mutant-specific RNA**

CTGGGA^aAAGAAGAGTACAGTGCC

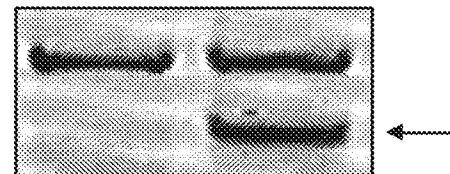


FIG. 33C

BRAF

HeLa

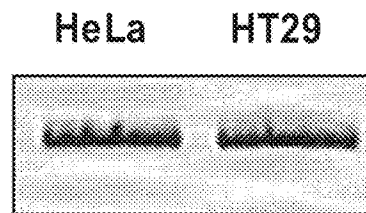
ACTCCATCGAGATTTCACTGTAG Wild-type

HT29

ACTCCATCGAGATTTCACTGTAG Wild-type

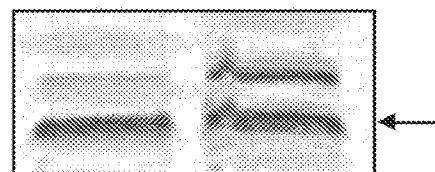
ACTCCATCGAGATTTCTCTGTAG (c.1799T>A)

PCR



**RFLP with
WT-Specific RNA**

ACTCCATCGAGATTTCACTGTAG



**RFLP with
Mutant-specific RNA**

ACTCCATCGAGATTTCTCTGTAG

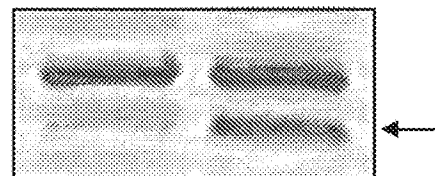


FIG. 33D

1

**COMPOSITIONS FOR INDUCING
MODIFICATIONS OF TARGET
ENDOGENOUS NUCLEIC ACID SEQUENCES
IN NUCLEUSES OF EUKARYOTIC CELLS**

CROSS REFERENCE TO RELATED
APPLICATIONS

The present application is a continuation application of U.S. application Ser. No. 18/932,745 filed Oct. 31, 2024, which is a continuation application of U.S. application Ser. No. 17/004,338 filed Aug. 27, 2020, which is a continuation application of U.S. application Ser. No. 14/685,568 filed Apr. 13, 2015, which is a continuation of PCT/KR2013/009488 filed Oct. 23, 2013, which claims priority to U.S. Provisional Application No. 61/837,481 filed on Jun. 20, 2013, U.S. Provisional Application No. 61/803,599 filed Mar. 20, 2013, and U.S. Provisional Application No. 61/717,324 filed Oct. 23, 2012, the entire contents of each aforementioned application are incorporated herein by reference.

SEQUENCE LISTING

The instant application contains a Sequence Listing which has been submitted electronically in XML file format and is hereby incorporated by reference in its entirety. Said XML copy, created on Jan. 13, 2025, is named 00161_SL.xml and is 495,164 bytes in size.

TECHNICAL FIELD

The present invention relates to targeted genome editing in eukaryotic cells or organisms. More particularly, the present invention relates to a composition for cleaving a target DNA in eukaryotic cells or organisms comprising a guide RNA specific for the target DNA and Cas protein-encoding nucleic acid or Cas protein, and use thereof.

BACKGROUND ART

CRISPRs (Clustered Regularly Interspaced Short Palindromic Repeats) are loci containing multiple short direct repeats that are found in the genomes of approximately 40% of sequenced bacteria and 90% of sequenced archaea. CRISPR functions as a prokaryotic immune system, in that it confers resistance to exogenous genetic elements such as plasmids and phages. The CRISPR system provides a form of acquired immunity. Short segments of foreign DNA, called spacers, are incorporated into the genome between CRISPR repeats, and serve as a memory of past exposures. CRISPR spacers are then used to recognize and silence exogenous genetic elements in a manner analogous to RNAi in eukaryotic organisms.

Cas9, an essential protein component in the Type II CRISPR/Cas system, forms an active endonuclease when complexed with two RNAs termed CRISPR RNA (crRNA) and trans-activating crRNA (tracrRNA), thereby slicing foreign genetic elements in invading phages or plasmids to protect the host cells. crRNA is transcribed from the CRISPR element in the host genome, which was previously captured from such foreign invaders. Recently, Jinek et al. (1) demonstrated that a single-chain chimeric RNA produced by fusing an essential portion of crRNA and tracrRNA could replace the two RNAs in the Cas9/RNA complex to form a functional endonuclease.

CRISPR/Cas systems offer an advantage to zinc finger and transcription activator-like effector DNA-binding pro-

2

teins, as the site specificity in nucleotide binding CRISPR-Cas proteins is governed by a RNA molecule instead of the DNA-binding protein, which can be more challenging to design and synthesize.

5 However, until now, a genome editing method using the RNA-guided endonuclease (RGEN) based on CRISPR/Cas system has not been developed.

Meanwhile, Restriction fragment length polymorphism (RFLP) is one of the oldest, most convenient, and least expensive methods of genotyping that is still used widely in molecular biology and genetics but is often limited by the lack of appropriate sites recognized by restriction endonucleases.

10 Engineered nuclease-induced mutations are detected by various methods, which include mismatch-sensitive T7 endonuclease I (T7E1) or Surveyor nuclease assays, RFLP, capillary electrophoresis of fluorescent PCR products, Dideoxy sequencing, and deep sequencing. The T7E1 and Surveyor assays are widely used but are cumbersome. Furthermore, these enzymes tend to underestimate mutation frequencies because mutant sequences can form homoduplexes with each other and cannot distinguish homozygous bi-allelic mutant clones from wildtype cells. RFLP is free of these limitations and therefore is a method of choice. Indeed, RFLP was one of the first methods to detect engineered nuclease-mediated mutations in cells and animals. Unfortunately, however, RFLP is limited by the availability of appropriate restriction sites. It is possible that no restriction sites are available at the target site of interest.

DISCLOSURE OF INVENTION

Technical Problem

35 Until now, a genome editing and genotyping method using the RNA-guided endonuclease (RGEN) based on CRISPR/Cas system has not been developed.

Under these circumstances, the present inventors have made many efforts to develop a genome editing method based on CRISPR/Cas system and finally established a programmable RNA-guided endonuclease that cleave DNA in a targeted manner in eukaryotic cells and organisms.

40 In addition, the present inventors have made many efforts to develop a novel method of using RNA-guided endonucleases (RGENs) in RFLP analysis. They have used RGENs to genotype recurrent mutations found in cancer and those induced in cells and organisms by engineered nucleases including RGENs themselves, thereby completing the present invention.

Solution to Problem

55 It is an object of the present invention to provide a composition for cleaving target DNA in eukaryotic cells or organisms comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is another object of the present invention to provide a composition for inducing targeted mutagenesis in eukaryotic cells or organisms, comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

60 It is still another object of the present invention to provide a kit for cleaving a target DNA in eukaryotic cells or organisms comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

3

It is still another object of the present invention to provide a kit for inducing targeted mutagenesis in eukaryotic cells or organisms comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a method for preparing a eukaryotic cell or organism comprising Cas protein and a guide RNA comprising a step of co-transfecting or serial-transfecting the eukaryotic cell or organism with a Cas protein-encoding nucleic acid or Cas protein, and a guide RNA or DNA that encodes the guide RNA.

It is still another object of the present invention to provide a eukaryotic cell or organism comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a method for cleaving a target DNA in eukaryotic cells or organisms comprising a step of transfecting the eukaryotic cells or organisms comprising a target DNA with a composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a method for inducing targeted mutagenesis in a eukaryotic cell or organism comprising a step of treating a eukaryotic cell or organism with a composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide an embryo, a genome-modified animal, or genome-modified plant comprising a genome edited by a composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a method of preparing a genome-modified animal comprising a step of introducing the composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein into an embryo of an animal; and a step of transferring the embryo into a oviduct of pseudopregnant foster mother to produce a genome-modified animal.

It is still another object of the present invention to provide a composition for genotyping mutations or variations in an isolated biological sample, comprising a guide RNA specific for the target DNA sequence Cas protein.

It is still another object of the present invention to provide a method of using a RNA-guided endonuclease (RGEN) to genotype mutations induced by engineered nucleases in cells or naturally-occurring mutations or variations, wherein the RGEN comprises a guide RNA specific for target DNA and Cas protein.

It is still another object of the present invention to provide a kit for genotyping mutations induced by engineered nucleases in cells or naturally-occurring mutations or variations, comprising a RNA-guided endonuclease (RGEN), wherein the RGEN comprises a guide RNA specific for target DNA and Cas protein.

It is an object of the present invention to provide a composition for cleaving target DNA in eukaryotic cells or organisms comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is another object of the present invention to provide a composition for inducing targeted mutagenesis in eukaryotic cells or organisms, comprising a guide RNA specific for

4

target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a kit for cleaving a target DNA in eukaryotic cells or organisms comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a kit for inducing targeted mutagenesis in eukaryotic cells or organisms comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a method for preparing a eukaryotic cell or organism comprising Cas protein and a guide RNA comprising a step of co-transfecting or serial-transfecting the eukaryotic cell or organism with a Cas protein-encoding nucleic acid or Cas protein, and a guide RNA or DNA that encodes the guide RNA.

It is still another object of the present invention to provide a eukaryotic cell or organism comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a method for cleaving a target DNA in eukaryotic cells or organisms comprising a step of transfecting the eukaryotic cells or organisms comprising a target DNA with a composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a method for inducing targeted mutagenesis in a eukaryotic cell or organism comprising a step of treating a eukaryotic cell or organism with a composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide an embryo, a genome-modified animal, or genome-modified plant comprising a genome edited by a composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a method of preparing a genome-modified animal comprising a step of introducing the composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein into an embryo of an animal; and a step of transferring the embryo into a oviduct of pseudopregnant foster mother to produce a genome-modified animal.

It is still another object of the present invention to provide a composition for genotyping mutations or variations in an isolated biological sample, comprising a guide RNA specific for the target DNA sequence Cas protein.

It is still another object of the present invention to provide a composition for genotyping nucleic acid sequences in pathogenic microorganisms in an isolated biological sample, comprising a guide RNA specific for the target DNA sequence and Cas protein.

It is still another object of the present invention to provide a kit for genotyping mutations or variations in an isolated biological sample, comprising the composition, specifically comprising a RNA-guided endonuclease (RGEN), wherein the RGEN comprises a guide RNA specific for target DNA and Cas protein.

It is still another object of the present invention to provide a method of genotyping mutations or variations in an

isolated biological sample, using the composition, specifically comprising a RNA-guided endonuclease (RGEN), wherein the RGEN comprises a guide RNA specific for target DNA and Cas protein.

Advantageous Effects of Invention

The present composition for cleaving a target DNA or inducing a targeted mutagenesis in eukaryotic cells or organisms, comprising a guide RNA specific for the target DNA and Cas protein-encoding nucleic acid or Cas protein, the kit comprising the composition, and the method for inducing targeted mutagenesis provide a new convenient genome editing tools. In addition, because custom RGENs can be designed to target any DNA sequence, almost any single nucleotide polymorphism or small insertion/deletion (indel) can be analyzed via RGEN-mediated RFLP, therefore, the composition and method of the present invention may be used in detection and cleaving naturally-occurring variations and mutations.

BRIEF DESCRIPTION OF DRAWINGS

FIGS. 1A and 1B show Cas9-catalyzed cleavage of plasmid DNA in vitro. FIG. 1A: Schematic representation of target DNA (SEQ ID NO: 112) and chimeric RNA sequences (SEQ ID NO: 113). Triangles indicate cleavage sites. The PAM sequence recognized by Cas9 is shown in bold. The sequences in the guide RNA (SEQ ID NO: 113) derived from crRNA and tracrRNA are shown in box and underlined, respectively. FIG. 1B: In vitro cleavage of plasmid DNA by Cas9. An intact circular plasmid or ApaLI-digested plasmid was incubated with Cas9 and guide RNA.

FIGS. 2A and 2B show Cas9-induced mutagenesis at an episomal target site. FIG. 2A: Schematic overview of cell-based assays using a RFP-GFP reporter. GFP is not expressed from this reporter because the GFP sequence is fused to the RFP sequence out-of-frame. The RFP-GFP fusion protein is expressed only when the target site between the two sequences is cleaved by a site-specific nuclease. FIG. 2B: Flow cytometry of cells transfected with Cas9. The percentage of cells that express the RFP-GFP fusion protein is indicated.

FIGS. 3A and 3B show RGEN-driven mutations at endogenous chromosomal sites. FIG. 3A: CCR5 locus. FIG. 3B: C4BPB locus. (Top) The T7E1 assay was used to detect RGEN-driven mutations. Arrows indicate the expected position of DNA bands cleaved by T7E1. Mutation frequencies (Indels (%)) were calculated by measuring the band intensities. (Bottom) DNA sequences of the wild-type (WT) CCR5 (SEQ ID NO: 114) and C4BPB (SEQ ID NO: 122) and mutant clones. DNA sequences of RGEN-induced mutations at the CCR5 locus: +1 (SEQ ID NO: 115), -13 (SEQ ID NO: 116), -14 (SEQ ID NO: 117), -18 (SEQ ID NO: 118), -19 (SEQ ID NO: 119), -24 (SEQ ID NO: 120), and -30 (SEQ ID NO: 121). DNA sequences of RGEN-induced mutations at the C4BPB locus: +1 (SEQ ID NO: 122), +2 (SEQ ID NO: 123), -30 (SEQ ID NO: 125), and -180 (SEQ ID NO: 126). The region of the target sequence complementary to the guide RNA is shown in box. The PAM sequence is shown in bold. Triangles indicate the cleavage site. Bases corresponding to microhomologies are underlined. The column on the right indicates the number of inserted or deleted bases.

FIGS. 4A, 4B, and 4C show that RGEN-driven off-target mutations are undetectable. FIG. 4A: On-target and potential off-target sequences. The human genome was searched in

silico for potential off-target sites. Four sites were identified, ADCY5 (SEQ ID NO: 128), KCNJ6 (SEQ ID NO: 129), CNTNAP2 (SEQ ID NO: 130), and Chr. 5 N/A (SEQ ID NO: 131), each of which carries 3-base mismatches with the CCR5 on-target (SEQ ID NO: 127). Mismatched bases are underlined. FIG. 4B: The T7E1 assay was used to investigate whether these sites were mutated in cells transfected with the Cas9/RNA complex. No mutations were detected at these sites. N/A (not applicable), an intergenic site. FIG. 4C: Cas9 did not induce off-target-associated chromosomal deletions. The CCR5-specific RGEN and ZFN were expressed in human cells. PCR was used to detect the induction of the 15-kb chromosomal deletions in these cells.

FIGS. 5A, 5B, 5C, and 5D show RGEN-induced Foxn1 gene targeting in mice. FIG. 5A: A schematic diagram depicting target DNA (SEQ ID NO: 132) and a sgRNA specific to exon 2 of the mouse Foxn1 gene (SEQ ID NO: 133). PAM in exon 2 is shown in a box and the sequence in the sgRNA that is complementary to exon 2 is underlined. Triangles indicate cleavage sites. FIG. 5B: Representative T7E1 assays demonstrating gene-targeting efficiencies of Cas9 mRNA plus Foxn1-specific sgRNA that were delivered via intra-cytoplasmic injection into one-cell stage mouse embryos. Numbers indicate independent founder mice generated from the highest dose. Arrows indicate bands cleaved by T7E1. FIG. 5C: DNA sequences of wild-type (WT) Foxn1 (SEQ ID NO: 134) and mutant alleles (SEQ ID NOS. 135-141) observed in three Foxn1 mutant founders identified in FIG. 5B. DNA sequences of mutant alleles in founder #108: -44 (SEQ ID NO: 135), -23 (SEQ ID NO: 136), -17 (SEQ ID NO: 137), and +1 (SEQ ID NO: 138). DNA sequences of mutant alleles in founder #111: +1 (SEQ ID NO: 138) and -11 (SEQ ID NO: 139). DNA sequences of mutant alleles in founder #114: -6 (SEQ ID NO: 140), -17 (SEQ ID NO: 137), and -8 (SEQ ID NO: 141). The number of occurrences is shown in parentheses. FIG. 5D: PCR genotyping of F1 progenies derived from crossing Foxn1 founder #108 and wild-type FVB/NTac. Note the segregation of the mutant alleles found in Foxn1 founder #108 in the progenies.

FIGS. 6A, 6B, and 6C show Foxn1 gene targeting in mouse embryos by intra-cytoplasmic injection of Cas9 mRNA and Foxn1-sgRNA. FIG. 6A: A representative result of a T7E1 assay monitoring the mutation rate after injecting the highest dose. Arrows indicate bands cleaved by T7E1. FIG. 6B: A summary of T7E1 assay results. Mutant fractions among in vitro cultivated embryos obtained after intra-cytoplasmic injection of the indicated RGEN doses are indicated. FIG. 6C: DNA sequences of wild-type (WT) Foxn1 (SEQ ID NO: 143) and Foxn1 mutant alleles (SEQ ID NOS. 144-152) identified from a subset of T7E1-positive mutant embryos. The DNA sequences of the mutant alleles are: Δ11 (SEQ ID NO: 144), Δ11+Δ17 (SEQ ID NO: 145) Δ57 (SEQ ID NO: 146), Δ17 (SEQ ID NO: 147), +1 (SEQ ID NO: 148), Δ12 (SEQ ID NO: 149), Δ72 (SEQ ID NO: 150), Δ25 (SEQ ID NO: 151), Δ24 (SEQ ID NO: 152). The target sequence of the wild-type allele is denoted in box.

FIGS. 7A, 7B, and 7C show Foxn1 gene targeting in mouse embryos using the recombinant Cas9 protein: Foxn1-sgRNA complex. FIG. 7A and FIG. 7B are representative T7E1 assays results and their summaries. Embryos were cultivated in vitro after they underwent pronuclear (FIG. 7A) or intra-cytoplasmic injection (FIG. 7B). Underlined numbers indicate T7E1-positive mutant founder mice. FIG. 7C: DNA sequences of wild-type (WT) Foxn1 (SEQ ID NO: 153) and Foxn1 mutant alleles (SEQ ID NOS. 154-166) identified from the in vitro cultivated embryos that were

obtained by the pronucleus injection of recombinant Cas9 protein: Foxn1-sgRNA complex at the highest dose. The target sequence of the wild-type allele is denoted in box. The DNA sequences of the mutant alleles are: $\Delta 18$ (SEQ ID NO: 154), $\Delta 20$ (SEQ ID NO: 155), $\Delta 19$ (SEQ ID NO: 156), $\Delta 17$ (SEQ ID NO: 157), $\Delta 11$ (SEQ ID NO: 158), $\Delta 3+1$ (SEQ ID NO: 159), $\Delta 2$ (SEQ ID NO: 160), +1, Embryo 1 (SEQ ID NO: 161), +1, Embryo 10 (SEQ ID NO: 162), $\Delta 6$ (SEQ ID NO: 163), $\Delta 5$ (SEQ ID NO: 164), $\Delta 28$ (SEQ ID NO: 165), and $\Delta 126$ (SEQ ID NO: 166).

FIGS. 8A, 8B, and 8C show Germ-line transmission of the mutant alleles found in Foxn1 mutant founder #12. FIG. 8A: wild type *i*PCR analysis. FIG. 8B: Foxn1 mutant founder #12 *i*PCR analysis. FIG. 8C: PCR genotyping of wild-type FVB/NTac, the founder mouse, and their F1 progenies.

FIGS. 9A and 9B show Genotypes of embryos generated by crossing Prkdc mutant founders. Prkdc mutant founders $\delta 25$ and $\eta 15$ were crossed and E13.5 embryos were isolated. FIG. 9A: *i*PCR analysis of wild-type, founder $\delta 25$, and founder $\eta 15$. Note that, due to the technical limitations of *i*PCR analysis, these results showed small differences from the precise sequences of the mutant alleles; e.g., from the sequence analysis, $\Delta 269/\Delta 61/WT$ and $\Delta 5+1/+7/+12/WT$ were identified in founders $\delta 25$ and $\eta 15$, respectively. FIG. 9B: Genotypes of the generated embryos.

FIGS. 10A, 10B, 10C, 10D, and 10E show Cas9 protein/sgRNA complex induced targeted mutation at CCR5 gene (FIGS. 10A-10C) and ABCC11 gene (FIGS. 10D-10E). FIG. 10A: Results of a T7E1 assay monitoring the mutation rate at CCR5 locus after introducing Cas9 protein and sgRNA or Cas9 protein and crRNA+tracrRNA into K562 cells. FIG. 10B: Results of a T7E1 assay using 1/5 scaled down doses of Cas9 protein and sgRNA. FIG. 10C: Wild-type (WT) CCR5 sequence (SEQ ID NO: 114) and Cas protein induced mutant sequences (SEQ ID NOs. 167-171 and 115) identified in CCR5 locus. The DNA sequences of the mutant sequences are: -4 (SEQ ID NO: 167), -4 (SEQ ID NO: 168), -7 (SEQ ID NO: 169), -1 (SEQ ID NO: 170), +1 (SEQ ID NO: 115), and -17, +1 (SEQ ID NO: 171). FIG. 10D: Results of a T7E1 assay monitoring the mutation rate at ABCC11 locus after introducing Cas9 protein and sgRNA into K562 cells. FIG. 10E: Wild-type (WT) ABCC11 sequence (SEQ ID NO: 172) and Cas9 protein induced mutant sequences (SEQ ID NOs. 173-176) identified in ABCC11 locus. The DNA sequences of the mutant sequences are: -6 (SEQ ID NO: 173), -3 (SEQ ID NO: 174), -29 (SEQ ID NO: 175), -20 (SEQ ID NO: 176), and -256 (TTCTC).

FIG. 11 shows recombinant Cas9 protein-induced mutations in Arabidopsis protoplasts.

FIG. 12 shows wild type BR11 sequence (SEQ ID NO: 177) and recombinant Cas9 protein-induced mutant sequences (SEQ ID NOs. 178-181) in the *Arabidopsis* BR11 gene. The DNA sequences of the mutant sequences are: -7 (SEQ ID NO: 178), -224 (SEQ ID NO: 179), -223 (SEQ ID NO: 180), and -223, +62 (SEQ ID NO: 181).

FIG. 13 shows T7E1 assay showing endogenous CCR5 gene disruption in 293 cells by treatment of Cas9-mal-9R4L and sgRNA/C9R4LC complex.

FIGS. 14A and 14B show mutation frequencies at on-target and off-target sites of RGENs reported in Fu et al. (2013). T7E1 assays analyzing genomic DNA from K562 cells (R) transfected serially with 20 μ g of Cas9-encoding plasmid and with 60 μ g and 120 μ g of in vitro transcribed GX19 crRNA and tracrRNA, respectively (1×10^6 cells), or (D) co-transfected with 1 μ g of Cas9-encoding plasmid and

1 μ g of GX19 SgRNA expression plasmid (2×10^5 cells). FIG. 14A: VEGFA site 1 on target sequence (SEQ ID NO: 182) and off target sequences, OT1-3 (SEQ ID NO: 183) and OT1-11 (SEQ ID NO: 184). VEGFA site 2 on target sequence (SEQ ID NO: 185) and off target sequences OT2-1 (SEQ ID NO: 186), OT2-9 (SEQ ID NO: 187) and OT2-24 (SEQ ID NO: 188). FIG. 14B: VEGFA site 3 on target sequence (SEQ ID NO: 189) and off target sequence OT3-18 (SEQ ID NO: 190) and EMX1 on target sequence (SEQ ID NO: 191) and off target sequence OT4-1 (SEQ ID NO: 192).

FIGS. 15A and 15B show comparison of guide RNA structure. Mutation frequencies of the RGENs reported in Fu et al. (2013) were measured at on-target and off-target sites using the T7E1 assay. K562 cells were co-transfected with the Cas9-encoding plasmid and the plasmid encoding GX19 sgRNA or GGX20 sgRNA. Off-target sites (OT1-3 etc.) are labeled as in Fu et al. (2013). FIG. 15A: VEGFA site 1 on target sequence (SEQ ID NO: 182) and off target sequences OT1-3 (SEQ ID NO: 183 and OT1-11 (SEQ ID NO: 184). VEGFA site 2 on target sequence (SEQ ID NO: 185) and off target sequences OT2-1 (SEQ ID NO: 186), OT2-9 (SEQ ID NO: 187), and OT2-24 (SEQ ID NO: 188). FIG. 15B: VEGFA site 3 on target sequence (SEQ ID NO: 189) and off target sequence OT3-18 (SEQ ID NO: 190) and EMX1 on target sequence (SEQ ID NO: 191) and off target sequence OT4-1 (SEQ ID NO: 192).

FIGS. 16A, 16B, 16C, and 16D show that in vitro DNA cleavage by Cas9 nickases. FIG. 16A: Schematic overview of the Cas9 nuclease and the paired Cas9 nickase. The PAM sequences and cleavage sites are shown in box. FIG. 16B: Target sites in the human AAVS1 locus. The position of each target site is shown in triangle. FIG. 16C: Schematic overview of DNA cleavage reactions. FAM dyes (shown in box) were linked to both 5' ends of the DNA substrate. FIG. 16D: DSBs and SSBs analyzed using fluorescent capillary electrophoresis. Fluorescently-labeled DNA substrates were incubated with Cas9 nucleases or nickases before electrophoresis.

FIGS. 17A and 17B show comparison of Cas9 nuclease and nickase behavior. FIG. 17A: On-target mutation frequencies associated with Cas9 nucleases (WT), nickases (D10A), and paired nickases at the following target sequences of the AAVS1 locus: S1 (SEQ ID NO: 193), S2 (SEQ ID NO: 194), S3 (SEQ ID NO: 195), S4 (SEQ ID NO: 196), S5 (SEQ ID NO: 197), S6 (SEQ ID NO: 198), AS1 (SEQ ID NO: 199), AS2 (SEQ ID NO: 200), and AS3 (SEQ ID NO: 201). Paired nickases that would produce 5' overhangs or 3' overhangs are indicated. FIG. 17B: Analysis of off-target effects of Cas9 nucleases and paired nickases. A total of seven potential off-target sites (SEQ ID NOs. 202-208) for three sgRNAs were analyzed. The mutation frequency for the S2 on-target sequence (SEQ ID NO: 194) was compared to the off-target sequences, S2 Off-1 (SEQ ID NO: 202) and S2 Off-2 (SEQ ID NO: 203). The mutation frequency for the S3 on-target sequence (SEQ ID NO: 195) was compared to the off-target sequences, S3 Off-1 (SEQ ID NO: 204) and S3 Off-2 (SEQ ID NO: 205). The mutation frequency for the AS2 on-target sequence (SEQ ID NO: 198) was compared to the off-target sequences, AS2Off-1 (SEQ ID NO: 206), AS2 Off-6 (SEQ ID NO: 207), and AS2 Off-9 (SEQ ID NO: 208).

FIGS. 18A, 18B, 18C, and 18D show paired Cas9 nickases tested at other endogenous human loci. The sgRNA target sites at the human CCR5 locus (FIG. 18A; SEQ ID NO: 209) and the BRCA2 locus (FIG. 18C; SEQ ID NO: 210). PAM sequences are indicated in a box. Genome editing activities at CCR5 (FIG. 18B) and BRCA2 (FIG.

18D) target sites were detected by the T7E1 assay. The repair of two nicks that would produce 5' overhangs led to the formation of indels much more frequently than did those producing 3' overhangs.

FIGS. 19A and 19B show that paired Cas9 nickases mediate homologous recombination. FIG. 19A: Strategy to detect homologous recombination. Donor DNA included an XbaI restriction enzyme site between two homology arms, whereas the endogenous target site lacked this site. A PCR assay was used to detect sequences that had undergone homologous recombination. To prevent amplification of contaminating donor DNA, primers specific to genomic DNA were used. FIG. 19B: Efficiency of homologous recombination. Only amplicons of a region in which homologous recombination had occurred could be digested with XbaI; the intensities of the cleavage bands were used to measure the efficiency of this method.

FIGS. 20A, 20B, 20C, and 20D show DNA splicing induced by paired Cas9 nickases. FIG. 20A: The target sites of paired nickases in the human AAVS1 locus. The distances between the AS2 site and each of the other sites are shown. Arrows indicate PCR primers. FIG. 20B: Genomic deletions detected using PCR. Asterisks indicate deletion-specific PCR products. FIG. 20C: DNA sequences of wild-type (WT) (SEQ ID NO: 211 and 332) and the following deletion-specific PCR products (SEQ ID Nos. 212-218) obtained using AS2 sgRNAs or deletion-specific PCR products (SEQ ID NOs. 219-224) using L1 sgRNAs. Target site PAM sequences are shown in box and sgRNA-matching sequences are shown in capital letters. Intact sgRNA-matching sequences are underlined. FIG. 20D: A schematic model of paired Cas9 nickase-mediated chromosomal deletions. Newly-synthesized DNA strands are shown in box.

FIGS. 21A, 21B, and 21C show that paired Cas9 nickases do not induce translocations. FIG. 21A: Schematic overview of chromosomal translocations between the on-target and off-target sites. FIG. 21B: PCR amplification to detect chromosomal translocations. FIG. 21C: Translocations induced by Cas9 nucleases but not by the nickase pair.

FIGS. 22A and 22B show a conceptual diagram of the T7E1 and RFLP assays. FIG. 22A: Comparison of assay cleavage reactions in four possible scenarios after engineered nuclease treatment in a diploid cell: (A) wild type, (B) a monoallelic mutation, (C) different biallelic mutations (hetero), and (D) identical biallelic mutations (homo). Black lines represent PCR products derived from each allele; dashed and dotted boxes indicate insertion/deletion mutations generated by NHEJ. FIG. 22B: Expected results of T7E1 and RGEN digestion resolved by electrophoresis.

FIG. 23 shows in vitro cleavage assay of a linearized plasmid containing the C4BPB target site bearing indels. DNA sequences of individual plasmid substrates (upper panel): WT (SEQ ID NO: 104), I1 (SEQ ID NO: 225), I2 (SEQ ID NO: 226), I3 (SEQ ID NO: 227), D1 (SEQ ID NO: 228), D2 (SEQ ID NO: 229), and D3 (SEQ ID NO: 230). The PAM sequence is underlined. Inserted bases are shown in box. Arrows (bottom panel) indicate expected positions of DNA bands cleaved by the wild-type-specific RGEN after electrophoresis.

FIGS. 24A and 24B show genotyping of mutations induced by engineered nucleases in cells via RGEN-mediated RFLP. FIG. 24A: Genotype of C4BPB wild type (SEQ ID NO: 231) and the following mutant K562 cell clones: +3 (SEQ ID NO: 232), -12 (SEQ ID NO: 233), -9 (SEQ ID NO: 234), -8 (SEQ ID NO: 235), -36 (SEQ ID NO: 236), +1 (SEQ ID NO: 237), +1 (SEQ ID NO: 238), +67 (SEQ ID NO: 239), -7, +1 (SEQ ID NO: 240), -94 (SEQ ID NO:

241). FIG. 24B: Comparison of the mismatch-sensitive T7E1 assay with RGEN-mediated RFLP analysis. Black arrows indicate the cleavage product by treatment of T7E1 enzyme or RGENS.

FIGS. 25A, 25B, and 25C show genotyping of RGEN-induced mutations via the RGEN-RFLP technique. FIG. 25A: Analysis of C4BPB-disrupted clones using RGEN-RFLP and T7E1 assays. Arrows indicate expected positions of DNA bands cleaved by RGEN or T7E1. FIG. 25B: Quantitative comparison of RGEN-RFLP analysis with T7E1 assays. Genomic DNA samples from wild-type and C4BPB-disrupted K562 cells were mixed in various ratios and subjected to PCR amplification. FIG. 25C: Genotyping of RGEN-induced mutations in the HLA-B gene in HeLa cells with RFLP and T7E1 analyses.

FIGS. 26A and 26B show genotyping of mutations induced by engineered nucleases in organisms via RGEN-mediated RFLP. FIG. 26A: Genotype of Pibf1 wild-type (WT) (SEQ ID NO: 242) and the following mutant founder mice: #1 (SEQ ID NO: 243 and SEQ ID NO: 244), #3 (SEQ ID NO: 245 and SEQ ID NO: 246), #4 (SEQ ID NO: 247 and SEQ ID NO: 242), #5 (SEQ ID NO: 246 and SEQ ID NO: 242), #6 (SEQ ID NO: 248 and SEQ ID NO: 249), #8 (SEQ ID NO: 250 and SEQ ID NO: 251), and #11 (SEQ ID NO: 252 and SEQ ID NO: 250). FIG. 26B: Comparison of the mismatch-sensitive T7E1 assay with RGEN-mediated RFLP analysis. Black arrows indicate the cleavage product by treatment of T7E1 enzyme or RGENS.

FIG. 27 shows RGEN-mediated genotyping of ZFN-induced mutations at a wild-type CCR5 sequence (SEQ ID NO: 253). The ZFN target site is shown in box. Black arrows indicate DNA bands cleaved by T7E1.

FIG. 28 shows polymorphic sites in a region of the human HLA-B gene (SEQ ID NO: 254). The sequence, which surrounds the RGEN target site, is that of a PCR amplicon from HeLa cells. Polymorphic positions are shown in box. The RGEN target site and the PAM sequence are shown in dashed and bolded box, respectively. Primer sequences are underlined.

FIGS. 29A and 29B show genotyping of oncogenic mutations via RGEN-RFLP analysis. FIG. 29A: A recurrent mutation (c.133-135 deletion of TCT; SEQ ID NO: 256) in the human CTNNB1 gene in HCT116 cells was detected by RGENS. The wild-type CTNNB1 sequence is represented by SEQ ID NO: 255. HeLa cells were used as a negative control. FIG. 29B: Genotyping of the KRAS substitution mutation (c.34 G>A) in the A549 cancer cell line with RGENs that contain mismatched guide RNA that are WT-specific (SEQ ID NO: 257) or mutant-specific (SEQ ID NO: 258). Mismatched nucleotides are shown in box. HeLa cells were used as a negative control. Arrows indicate DNA bands cleaved by RGENs. DNA sequences confirmed by Sanger sequencing are shown: wild-type (SEQ ID NO: 259) and c. 34G>A (SEQ ID NO: 260).

FIGS. 30A, 30B, 30C, and 30D show genotyping of the CCR5 delta32 allele in HEK293T cells via RGEN-RFLP analysis. FIG. 30A: RGEN-RFLP assays of cell lines. DNA sequences of the wild-type CCR5 locus (SEQ ID NO: 262) and delta 32 mutation (SEQ ID NO: 261) are shown. K562, SKBR3, and HeLa cells were used as wild-type controls. Arrows indicate DNA bands cleaved by RGENs. FIG. 30B: DNA sequence of wild-type (SEQ ID NO: 263) and delta32 CCR5 alleles (SEQ ID NO: 264). Both on-target and off-target sites of RGENs used in RFLP analysis are underlined. A single-nucleotide mismatch between the two sites is shown in box. The PAM sequence is underlined. FIG. 30C: In vitro cleavage of plasmids harboring WT or del32 CCR5

alleles using the wild-type-specific RGEN. FIG. 30D Confirming the presence of an off-target site of the CCR5-delta32-specific RGEN at the CCR5 locus. In vitro cleavage assays of plasmids harboring either on-target (SEQ ID NO: 265) or off-target sequences (SEQ ID NO: 266) using various amounts of the del32-specific RGEN.

FIGS. 31A and 31B show genotyping of a KRAS point mutation (c.34 G>A). FIG. 31A: RGEN-RFLP analysis of the KRAS mutation (c.34 G>A) in cancer cell lines. PCR products from HeLa cells (used as a wild-type control) or A549 cells, which are homozygous for the point mutation, were digested with RGENs with perfectly matched crRNA specific to the wild-type sequence (SEQ ID NO: 259) or the mutant sequence (SEQ ID NO: 260). KRAS genotypes in these cells were confirmed by Sanger sequencing. FIG. 31B: Plasmids harboring either the wild-type (SEQ ID NO: 259) or mutant KRAS sequences (SEQ ID NO: 260) were digested using RGENs with perfectly matched crRNAs or attenuated, one-base mismatched crRNAs: m7 (SEQ ID NO: 267), m6 (SEQ ID NO: 257), m5 (SEQ ID NO: 268), m4 (SEQ ID NO: 269), m8 (SEQ ID NO: 260), m7, 8 (SEQ ID NO: 270), m6, 8 (SEQ ID NO: 258), m5, 8 (SEQ ID NO: 271), and m4, 8 (SEQ ID NO: 272). Attenuated crRNAs that were chosen for genotyping are labeled in box above the gels.

FIGS. 32A and 32B show genotyping of a PIK3CA point mutation (c.3140 A>G). FIG. 32A: RGEN-RFLP analysis of the PIK3CA mutation (c.3140 A>G) in cancer cell lines. PCR products from HeLa cells (used as a wild-type control) or HCT116 cells that are heterozygous for the point mutation were digested with RGENs with perfectly matched crRNA specific to the wild-type sequence (SEQ ID NO: 273) or the mutant sequence (SEQ ID NO: 274). PIK3CA genotypes in these cells were confirmed by Sanger sequencing. FIG. 32B: Plasmids harboring either the wild-type PIK3CA sequence (SEQ ID NO: 273) or mutant PIK3CA sequence (SEQ ID NO: 274) were digested using RGENs with perfectly matched crRNAs or attenuated, one-base mismatched crRNAs: m5 (SEQ ID NO: 275), m6 (SEQ ID NO: 276), m7 (SEQ ID NO: 277), m10 (SEQ ID NO: 278), m13 (SEQ ID NO: 279), m16 (SEQ ID NO: 280), m19 (SEQ ID NO: 281), m4 (SEQ ID NO: 274), m4, 5 (SEQ ID NO: 282), m4, 6 (SEQ ID NO: 283), m4, 7 (SEQ ID NO: 284), m4, 10 (SEQ ID NO: 285), m4, 13 (SEQ ID NO: 286), m4, 16 (SEQ ID NO: 287), and m4, 19 (SEQ ID NO: 288). Attenuated crRNAs that were chosen for genotyping are labeled in box above the gels.

FIGS. 33A, 33B, 33C, and 33D show genotyping of recurrent point mutations in cancer cell lines. FIG. 33A: RGEN-RFLP assays to distinguish between a wild-type IDH gene sequence (SEQ ID NO: 289) and a recurrent oncogenic point mutation sequence in the IDH gene (c.394c>T; SEQ ID NO: 290). RGENs with attenuated, one-base mismatched crRNAs, SEQ ID NO: 291 (WT-Specific RNA) and SEQ ID NO: 292 (Mutant-Specific RNA), distinguished the wild type and mutant IDH sequences. FIG. 33B: RGEN-RFLP assays to distinguish between a wild-type PIK3CA gene sequence (SEQ ID NO: 271) and a recurrent oncogenic point mutation sequence in the PIK3CA gene (c.3140A>G; SEQ ID NO: 273). RGENs with attenuated, one-base mismatched crRNAs, SEQ ID NO: 275 (WT-Specific RNA) and SEQ ID NO: 284 (Mutant-Specific RNA), distinguished the wild type and mutant PIK3CA sequences. FIG. 33C: RGEN-RFLP assays to distinguish between a wild-type NRAS gene sequence (SEQ ID NO: 293) and a recurrent oncogenic point mutation sequence in the NRAS gene (c.181C>A; SEQ ID NO: 294). RGENs with perfectly matched crRNAs, SEQ ID

NO: 293 (WT-Specific RNA) and SEQ ID NO: 294 (Mutant-Specific RNA), distinguished the wild type and mutant NRAS sequences. FIG. 33D: RGEN-RFLP assays to distinguish between a wild-type BRAF gene sequence (SEQ ID NO: 295) and a recurrent oncogenic point mutation sequence in the BRAF gene (c.1799T>A; SEQ ID NO: 296). RGENs with perfectly matched crRNAs, SEQ ID NO: 295 (WT-Specific RNA) and SEQ ID NO: 296 (Mutant-Specific RNA), distinguished the wild type and mutant BRAF sequences. Genotypes of each cell line confirmed by Sanger sequencing are shown. Mismatched nucleotides are shown in box. Black arrows indicate DNA bands cleaved by RGENs.

BEST MODE FOR CARRYING OUT THE INVENTION

In accordance with one aspect of the invention, the present invention provides a composition for cleaving target DNA in eukaryotic cells or organisms comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein. In addition, the present invention provides a use of the composition for cleaving target DNA in eukaryotic cells or organisms comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

In the present invention, the composition is also referred to as a RNA-guided endonuclease (RGEN) composition.

ZFNs and TALENs enable targeted mutagenesis in mammalian cells, model organisms, plants, and livestock, but the mutation frequencies obtained with individual nucleases are widely different from each other. Furthermore, some ZFNs and TALENs fail to show any genome editing activities. DNA methylation may limit the binding of these engineered nucleases to target sites. In addition, it is technically challenging and time-consuming to make customized nucleases.

The present inventors have developed a new RNA-guided endonuclease composition based on Cas protein to overcome the disadvantages of ZFNs and TALENs.

Prior to the present invention, an endonuclease activity of Cas proteins has been known. However, it has not been known whether the endonuclease activity of Cas protein would function in an eukaryotic cell because of the complexity of the eukaryotic genome. Further, until now, a composition comprising Cas protein or Cas protein-encoding nucleic acid and a guide RNA specific for the target DNA to cleave a target DNA in eukaryotic cells or organisms has not been developed.

Compared to ZFNs and TALENs, the present RGEN composition based on Cas protein can be more readily customized because only the synthetic guide RNA component is replaced to make a new genome-editing nuclease. No sub-cloning steps are involved to make customized RNA guided endonucleases. Furthermore, the relatively small size of the Cas gene (for example, 4.2 kbp for Cas9) as compared to a pair of TALEN genes (~6 kbp) provides an advantage for this RNA-guided endonuclease composition in some applications such as virus-mediated gene delivery. Further, this RNA-guided endonuclease does not have off-target effects and thus does not induce unwanted mutations, deletion, inversions, and duplications. These features make the present RNA-guided endonuclease composition a scalable, versatile, and convenient tool for genome engineering in eukaryotic cells and organisms. In addition, RGEN can be designed to target any DNA sequence, almost any single nucleotide polymorphism or small insertion/deletion (indel)

can be analyzed via RGEN-mediated RFLP. The specificity of RGENs is determined by the RNA component that hybridizes with a target DNA sequence of up to 20 base pairs (bp) in length and by the Cas9 protein that recognizes the protospacer-adjacent motif (PAM). RGENs are readily reprogrammed by replacing the RNA component. Therefore, RGENs provide a platform to use simple and robust RFLP analysis for various sequence variations.

The target DNA may be an endogenous DNA, or artificial DNA, preferably, endogenous DNA.

As used herein, the term “Cas protein” refers to an essential protein component in the CRISPR/Cas system, forms an active endonuclease or nickase when complexed with two RNAs termed CRISPR RNA (crRNA) and transactivating crRNA (tracrRNA).

The information on the gene and protein of Cas are available from GenBank of National Center for Biotechnology Information (NCBI), without limitation.

The CRISPR-associated (cas) genes encoding Cas proteins are often associated with CRISPR repeat-spacer arrays. More than forty different Cas protein families have been described. Of these protein families, Cas1 appears to be ubiquitous among different CRISPR/Cas systems. There are three types of CRISPR-Cas system. Among them, Type II CRISPR/Cas system involving Cas9 protein and crRNA and tracrRNA is representative and is well known. Particular combinations of cas genes and repeat structures have been used to define 8 CRISPR subtypes (Ecoli, Ypest, Nmeni, Dvulg, Tneap, Hmari, Apem, and Mtube).

The Cas protein may be linked to a protein transduction domain. The protein transduction domain may be poly-arginine or a TAT protein derived from HIV, but it is not limited thereto.

The present composition may comprise Cas component in the form of a protein or in the form of a nucleic acid encoding Cas protein.

In the present invention, Cas protein may be any Cas protein provided that it has an endonuclease or nickase activity when complexed with a guide RNA.

Preferably, Cas protein is Cas9 protein or variants thereof.

The variant of the Cas9 protein may be a mutant form of Cas9 in which the catalytic aspartate residue is changed to any other amino acid. Preferably, the other amino acid may be an alanine, but it is not limited thereto.

Further, Cas protein may be the one isolated from an organism such as *Streptococcus* sp., preferably *Streptococcus pyogenes* or a recombinant protein, but it is not limited thereto.

The Cas protein derived from *Streptococcus pyogenes* may recognize NGG trinucleotide. The Cas protein may comprise an amino acid sequence of SEQ ID NO: 109, but it is not limited thereto.

The term “recombinant” when used with reference, e.g., to a cell, nucleic acid, protein, or vector, indicates that the cell, nucleic acid, protein or vector, has been modified by the introduction of a heterologous nucleic acid or protein or the alteration of a native nucleic acid or protein, or that the cell is derived from a cell so modified. Thus, for example, a recombinant Cas protein may be generated by reconstituting Cas protein-encoding sequence using the human codon table.

As for the present invention, Cas protein-encoding nucleic acid may be a form of vector, such as plasmid comprising Cas-encoding sequence under a promoter such as CMV or CAG. When Cas protein is Cas9, Cas9 encoding sequence may be derived from *Streptococcus* sp., and preferably derived from *Streptococcus pyogenes*. For example,

Cas9 encoding nucleic acid may comprise the nucleotide sequence of SEQ ID NO: 1. Moreover, Cas9 encoding nucleic acid may comprise the nucleotide sequence having homology of at least 50% to the sequence of SEQ ID NO: 1, preferably at least 60, 70, 80, 90, 95, 97, 98, or 99% to the SEQ ID NO:1, but it is not limited thereto. Cas9 encoding nucleic acid may comprise the nucleotide sequence of SEQ ID NOs. 108, 110, 106, or 107.

As used herein, the term “guide RNA” refers to a RNA which is specific for the target DNA and can form a complex with Cas protein and bring Cas protein to the target DNA.

In the present invention, the guide RNA may consist of two RNA, i.e., CRISPR RNA (crRNA) and transactivating crRNA (tracrRNA) or be a single-chain RNA (sgRNA) produced by fusion of an essential portion of crRNA and tracrRNA.

The guide RNA may be a dualRNA comprising a crRNA and a tracrRNA.

If the guide RNA comprises the essential portion of crRNA and tracrRNA and a portion complementary to a target, any guide RNA may be used in the present invention.

The crRNA may hybridize with a target DNA.

The RGEN may consist of Cas protein, and dualRNA (invariable tracrRNA and target-specific crRNA), or Cas protein and sgRNA (fusion of an essential portion of invariable tracrRNA and target-specific crRNA), and may be readily reprogrammed by replacing crRNA.

The guide RNA further comprises one or more additional nucleotides at the 5' end of the single-chain guide RNA or the crRNA of the dualRNA.

Preferably, the guide RNA further comprises 2-additional guanine nucleotides at the 5' end of the single-chain guide RNA or the crRNA of the dualRNA.

The guide RNA may be transferred into a cell or an organism in the form of RNA or DNA that encodes the guide RNA. The guide RNA may be in the form of an isolated RNA, RNA incorporated into a viral vector, or is encoded in a vector. Preferably, the vector may be a viral vector, plasmid vector, or agrobacterium vector, but it is not limited thereto.

A DNA that encodes the guide RNA may be a vector comprising a sequence coding for the guide RNA. For example, the guide RNA may be transferred into a cell or organism by transfecting the cell or organism with the isolated guide RNA or plasmid DNA comprising a sequence coding for the guide RNA and a promoter.

Alternatively, the guide RNA may be transferred into a cell or organism using virus-mediated gene delivery.

When the guide RNA is transfected in the form of an isolated RNA into a cell or organism, the guide RNA may be prepared by in vitro transcription using any in vitro transcription system known in the art. The guide RNA is preferably transferred to a cell in the form of isolated RNA rather than in the form of plasmid comprising encoding sequence for a guide RNA. As used herein, the term “isolated RNA” may be interchangeable to “naked RNA”. This is cost- and time-saving because it does not require a step of cloning. However, the use of plasmid DNA or virus-mediated gene delivery for transfection of the guide RNA is not excluded.

The present RGEN composition comprising Cas protein or Cas protein-encoding nucleic acid and a guide RNA can specifically cleave a target DNA due to a specificity of the guide RNA for a target and an endonuclease or nickase activity of Cas protein.

As used herein, the term “cleavage” refers to the breakage of the covalent backbone of a nucleotide molecule.

In the present invention, a guide RNA may be prepared to be specific for any target which is to be cleaved. Therefore, the present RGEN composition can cleave any target DNA by manipulating or genotyping the target-specific portion of the guide RNA.

The guide RNA and the Cas protein may function as a pair. As used herein, the term "paired Cas nickase" may refer to the guide RNA and the Cas protein functioning as a pair. The pair comprises two guide RNAs. The guide RNA and Cas protein may function as a pair, and induce two nicks on different DNA strand. The two nicks may be separated by at least 100 bps, but are not limited thereto.

In the Example, the present inventors confirmed that paired Cas nickase allow targeted mutagenesis and large deletions of up to 1-kbp chromosomal segments in human cells. Importantly, paired nickases did not induce indels at off-target sites at which their corresponding nucleases induce mutations. Furthermore, unlike nucleases, paired nickases did not promote unwanted translocations associated with off-target DNA cleavages. In principle, paired nickases double the specificity of Cas9-mediated mutagenesis and will broaden the utility of RNA-guided enzymes in applications that require precise genome editing such as gene and cell therapy.

In the present invention, the composition may be used in the genotyping of a genome in the eukaryotic cells or organisms *in vitro*.

In one specific embodiment, the guide RNA may comprise the nucleotide sequence of Seq ID. No. 1, wherein the portion of nucleotide position 3~22 is a target-specific portion and thus, the sequence of this portion may be changed depending on a target.

As used herein, a eukaryotic cell or organism may be yeast, fungus, protozoa, plant, higher plant, and insect, or amphibian cells, or mammalian cells such as CHO, HeLa, HEK293, and COS-1, for example, cultured cells (*in vitro*), graft cells and primary cell culture (*in vitro* and *ex vivo*), and *in vivo* cells, and also mammalian cells including human, which are commonly used in the art, without limitation.

In one specific embodiment, it was found that Cas9 protein/single-chain guide RNA could generate site-specific DNA double-strand breaks *in vitro* and in mammalian cells, whose spontaneous repair induced targeted genome mutations at high frequencies.

Moreover, it was found that gene-knockout mice could be induced by the injection of Cas9 protein/guide RNA complexes or Cas9 mRNA/guide RNA into one-cell stage embryo and germ-line transmittable mutations could be generated by Cas9/guide RNA system.

Using Cas protein rather than a nucleic acid encoding Cas protein to induce a targeted mutagenesis is advantageous because exogenous DNA is not introduced into an organism. Thus, the composition comprising Cas protein and a guide RNA may be used to develop therapeutics or value-added crops, livestock, poultry, fish, pets, etc.

In accordance with another aspect of the invention, the present invention provides a composition for inducing targeted mutagenesis in eukaryotic cells or organisms, comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein. In addition, the present invention provides a use of the composition for inducing targeted mutagenesis in eukaryotic cells or organisms, comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

A guide RNA, Cas protein-encoding nucleic acid or Cas protein are as described in the above.

In accordance with another aspect of the invention, the present invention provides a kit for cleaving a target DNA or inducing targeted mutagenesis in eukaryotic cells or organisms comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

A guide RNA, Cas protein-encoding nucleic acid or Cas protein are as described in the above.

The kit may comprise a guide RNA and Cas protein-encoding nucleic acid or Cas protein as separate components or as one composition.

The present kit may comprise some additional components necessary for transferring the guide RNA and Cas component to a cell or an organism. For example, the kit may comprise an injection buffer such as DEPC-treated injection buffer, and materials necessary for analysis of mutation of a target DNA, but are not limited thereto.

In accordance with another aspect, the present invention provides a method for preparing a eukaryotic cell or organism comprising Cas protein and a guide RNA comprising a step of co-transfecting or serial-transfecting the eukaryotic cell or organism with a Cas protein-encoding nucleic acid or Cas protein, and a guide RNA or DNA that encodes the guide RNA.

A guide RNA, Cas protein-encoding nucleic acid or Cas protein are as described in the above.

In the present invention, a Cas protein-encoding nucleic acid or Cas protein and a guide RNA or DNA that encodes the guide RNA may be transferred into a cell by various methods known in the art, such as microinjection, electroporation, DEAE-dextran treatment, lipofection, nanoparticle-mediated transfection, protein transduction domain mediated transduction, virus-mediated gene delivery, and PEG-mediated transfection in protoplast, and so on, but are not limited thereto. Also, a Cas protein encoding nucleic acid or Cas protein and a guide RNA may be transferred into an organism by various method known in the art to administer a gene or a protein such as injection. A Cas protein-encoding nucleic acid or Cas protein may be transferred into a cell in the form of complex with a guide RNA, or separately. Cas protein fused to a protein transduction domain such as Tat can also be delivered efficiently into cells.

Preferably, the eukaryotic cell or organism is co-transfected or serial-transfected with a Cas9 protein and a guide RNA.

The serial-transfection may be performed by transfection with Cas protein-encoding nucleic acid first, followed by second transfection with naked guide RNA. Preferably, the second transfection is after 3, 6, 12, 18, 24 hours, but it is not limited thereto.

In accordance with another aspect, the present invention provides a eukaryotic cell or organism comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

The eukaryotic cells or organisms may be prepared by transferring the composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein into the cell or organism.

The eukaryotic cell may be yeast, fungus, protozoa, higher plant, and insect, or amphibian cells, or mammalian cells such as CHO, HeLa, HEK293, and COS-1, for example, cultured cells (*in vitro*), graft cells and primary cell culture (*in vitro* and *ex vivo*), and *in vivo* cells, and also mammalian cells including human, which are commonly

used in the art, without limitation. Further the organism may be yeast, fungus, protozoa, plant, higher plant, insect, amphibian, or mammal.

In accordance with another aspect of the invention, the present invention provides a method for cleaving a target DNA or inducing targeted mutagenesis in eukaryotic cells or organisms, comprising a step of treating a cell or organism comprising a target DNA with a composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

The step of treating a cell or organism with the composition may be performed by transferring the present composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein into the cell or organism.

As described in the above, such transfer may be performed by microinjection, transfection, electroporation, and so on.

In accordance with another aspect of the invention, the present invention provides an embryo comprising a genome edited by the present RGEN composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

Any embryo can be used in the present invention, and for the present invention, the embryo may be an embryo of a mouse. The embryo may be produced by injecting PMSG (Pregnant Mare Serum Gonadotropin) and hCG (human Chorionic Gonadotropin) into a female mouse of 4 to 7 weeks and the super-ovulated female mouse may be mated to males, and the fertilized embryos may be collected from oviducts.

The present RGEN composition introduced into an embryo can cleave a target DNA complementary to the guide RNA by the action of Cas protein and cause a mutation in the target DNA. Thus, the embryo into which the present RGEN composition has been introduced has an edited genome.

In one specific embodiment, it was found that the present RGEN composition could cause a mutation in a mouse embryo and the mutation could be transmitted to offspring.

A method for introducing the RGEN composition into the embryo may be any method known in the art, such as microinjection, stem cell insertion, retrovirus insertion, and so on. Preferably, a microinjection technique can be used.

In accordance with another aspect, the present invention provides a genome-modified animal obtained by transferring the embryo comprising a genome edited by the present RGEN composition into the oviducts of an animal.

In the present invention, the term "genome-modified animal" refers to an animal of which genome has been modified in the stage of embryo by the present RGEN composition and the type of the animal is not limited.

The genome-modified animal has mutations caused by a targeted mutagenesis based on the present RGEN composition. The mutations may be any one of deletion, insertion, translocation, inversion. The site of mutation depends on the sequence of guide RNA of the RGEN composition.

The genome-modified animal having a mutation of a gene may be used to determine the function of the gene.

In accordance with another aspect of the invention, the present invention provides a method of preparing a genome-modified animal comprising a step of introducing the present RGEN composition comprising a guide RNA specific for the target DNA or DNA that encodes the guide RNA and Cas protein-encoding nucleic acid or Cas protein into an embryo

of an animal; and a step of transferring the embryo into a oviduct of pseudopregnant foster mother to produce a genome-modified animal.

The step of introducing the present RGEN composition may be accomplished by any method known in the art such as microinjection, stem cell insertion, retroviral insertion, and so on.

In accordance with another aspect of the invention, the present invention provides a plant regenerated from the genome-modified protoplasts prepared by the method for eukaryotic cells comprising the RGEN composition.

In accordance with another aspect of the invention, the present invention provides a composition for genotyping mutations or variations in an isolated biological sample, comprising a guide RNA specific for the target DNA sequence Cas protein. In addition, the present invention provides a composition for genotyping nucleic acid sequences in pathogenic microorganisms in an isolated biological sample, comprising a guide RNA specific for the target DNA sequence and Cas protein.

A guide RNA, Cas protein-encoding nucleic acid or Cas protein are as described in the above.

As used herein the term "genotyping" refers to the "Restriction fragment length polymorphism (RFLP) assay".

RFLP may be used in 1) the detection of indel in cells or organisms induced by the engineered nucleases, 2) the genotyping naturally-occurring mutations or variations in cells or organisms, or 3) the genotyping the DNA of infected pathogenic microorganisms including virus or bacteria, etc.

The mutations or variation may be induced by engineered nucleases in cells.

The engineered nuclease may be a Zinc Finger Nuclease (ZFNs), Transcription Activator-Like Effector Nucleases (TALENs), or RGENs, but it is not limited thereto.

As used herein the term "biological sample" includes samples for analysis, such as tissues, cells, whole blood, serum, plasma, saliva, sputum, cerebrospinal fluid or urine, but is not limited thereto.

The mutations or variation may be a naturally-occurring mutations or variations.

The mutations or variations are induced by the pathogenic microorganisms. Namely, the mutations or variations occur due to the infection of pathogenic microorganisms, when the pathogenic microorganisms are detected, the biological sample is identified as infected.

The pathogenic microorganisms may be virus or bacteria, but are not limited thereto.

Engineered nuclease-induced mutations are detected by various methods, which include mismatch-sensitive Surveyor or T7 endonuclease I (T7E1) assays, RFLP analysis, fluorescent PCR, DNA melting analysis, and Sanger and deep sequencing. The T7E1 and Surveyor assays are widely used but often underestimate mutation frequencies because the assays detect heteroduplexes (formed by the hybridization of mutant and wild-type sequences or two different mutant sequences); they fail to detect homoduplexes formed by the hybridization of two identical mutant sequences. Thus, these assays cannot distinguish homozygous biallelic mutant clones from wild-type cells nor heterozygous biallelic mutants from heterozygous monoallelic mutants (FIG. 22). In addition, sequence polymorphisms near the nuclease target site can produce confounding results because the enzymes can cleave heteroduplexes formed by hybridization of these different wild-type alleles. RFLP analysis is free of these limitations and therefore is a method of choice. Indeed, RFLP analysis was one of the first methods used to detect

engineered nuclease-mediated mutations. Unfortunately, however, it is limited by the availability of appropriate restriction sites.

In accordance with another aspect of the invention, the present invention provides a kit for genotyping mutations or variations in an isolated biological sample, comprising the composition for genotyping mutations or variations in an isolated biological sample. In addition, the present invention provides a kit for genotyping nucleic acid sequences in pathogenic microorganisms in an isolated biological sample, comprising a guide RNA specific for the target DNA sequence and Cas protein.

A guide RNA, Cas protein-encoding nucleic acid or Cas protein are as described in the above.

In accordance with another aspect of the invention, the present invention provides a method of genotyping mutations or variations in an isolated biological sample, using the composition for genotyping mutations or variations in an isolated biological sample. In addition, the present invention provides a method of genotyping nucleic acid sequences in pathogenic microorganisms in an isolated biological sample, comprising a guide RNA specific for the target DNA sequence and Cas protein.

A guide RNA, Cas protein-encoding nucleic acid or Cas protein are as described in the above.

Mode for the Invention

Hereinafter, the present invention will be described in more detail with reference to Examples. However, these Examples are for illustrative purposes only, and the invention is not intended to be limited by these Examples.

Example 1: Genome Editing Assay

1-1. DNA Cleavage Activity of Cas9 Protein

Firstly, the DNA cleavage activity of Cas9 derived from *Streptococcus pyogenes* in the presence or absence of a chimeric guide RNA in vitro was tested.

To this end, recombinant Cas9 protein that was expressed in and purified from *E. coli* was used to cleave a predigested or circular plasmid DNA that contained the 23-base pair (bp) human CCR5 target sequence. A Cas9 target sequence consists of a 20-bp DNA sequence complementary to crRNA or a chimeric guide RNA and the trinucleotide (5'-NGG-3') protospacer adjacent motif (PAM) recognized by Cas9 itself (FIG. 1A).

Specifically, the Cas9-coding sequence (4,104 bp), derived from *Streptococcus pyogenes* strain MI GAS (NC_002737.1), was reconstituted using the human codon usage table and synthesized using oligonucleotides. First, 1-kb DNA segments were assembled using overlapping ~35-mer oligonucleotides and Phusion™ polymerase (New England Biolabs) and cloned into T-vector (SolGent). A full-length Cas9 sequence was assembled using four 1-kbp DNA segments by overlap PCR. The Cas9-encoding DNA segment was subcloned into p3s, which was derived from pcDNA3.1 (Invitrogen). In this vector, a peptide tag (NH₂-GGSGPPKKRKYVPYDVPDYA-COOH, SEQ ID NO: 2) containing the HA epitope and a nuclear localization signal (NLS) was added to the C-terminus of Cas9. Expression and nuclear localization of the Cas9 protein in HEK 293T cells were confirmed by western blotting using anti-HA antibody (Santa Cruz).

Then, the Cas9 cassette was subcloned into pET28-b(+) and transformed into BL21(DE3). The expression of Cas9 was induced using 0.5 mM IPTG for 4 h at 25° C. The Cas9

protein containing the His6-tag at the C terminus was purified using Ni-NTA agarose resin (Qiagen) and dialyzed against 20 mM HEPES (pH 7.5), 150 mM KCl, 1 mM DTT, and 10% glycerol (1). Purified Cas9 (50 nM) was incubated with super-coiled or pre-digested plasmid DNA (300 ng) and chimeric RNA (50 nM) in a reaction volume of 20 µl in NEB buffer 3 for 1 h at 37° C. Digested DNA was analyzed by electrophoresis using 0.8% agarose gels.

Cas9 cleaved the plasmid DNA efficiently at the expected position only in the presence of the synthetic RNA and did not cleave a control plasmid that lacked the target sequence (FIG. 1B).

1-2. DNA Cleavage by Cas9/Guide RNA Complex in Human Cells

A RFP-GFP reporter was used to investigate whether the Cas9/guide RNA complex can cleave the target sequence incorporated between the RFP and GFP sequences in mammalian cells.

In this reporter, the GFP sequence is fused to the RFP sequence out-of-frame (2). The active GFP is expressed only when the target sequence is cleaved by site-specific nucleases, which causes frameshifting small insertions or deletions (indels) around the target sequence via error-prone non-homologous end-joining (NHEJ) repair of the double-strand break (DSB) (FIG. 2).

The RFP-GFP reporter plasmids used in this study were constructed as described previously (2). Oligonucleotides corresponding to target sites (Table 1) were synthesized (Macrogen) and annealed. The annealed oligonucleotides were ligated into a reporter vector digested with EcoRI and BamHI.

HEK 293T cells were co-transfected with Cas9-encoding plasmid (0.8 µg) and the RFP-GFP reporter plasmid (0.2 µg) in a 24-well plate using Lipofectamine™ 2000 (Invitrogen).

Meanwhile, the in vitro transcribed chimeric RNA had been prepared as follows. RNA was in vitro transcribed through run-off reactions using the MEGAscript™ T7 kit (Ambion) according to the manufacturer's manual. Templates for RNA in vitro transcription were generated by annealing two complementary single strand DNAs or by PCR amplification (Table 1). Transcribed RNA was resolved on a 8% denaturing urea-PAGE gel. The gel slice containing RNA was cut out and transferred to probe elution buffer. RNA was recovered in nuclease-free water followed by phenol: chloroform extraction, chloroform extraction, and ethanol precipitation. Purified RNAs were quantified by spectrometry.

At 12 h post transfection, chimeric RNA (1 µg) prepared by in vitro transcription was transfected using Lipofectamine 2000.

At 3d post-transfection, transfected cells were subjected to flow cytometry and cells expressing both RFP and GFP were counted.

It was found that GFP-expressing cells were obtained only when the cells were transfected first with the Cas9 plasmid and then with the guide RNA 12 h later (FIG. 2), demonstrating that RGENs could recognize and cleave the target DNA sequence in cultured human cells. Thus GFP-expressing cells were obtained by serial-transfection of the Cas9 plasmid and the guide RNA rather than co-transfection.

TABLE 1

| Gene | sequence (5' to 3') | SEQ ID NO. |
|---|--|------------|
| Oligonucleotides used for the construction of the reporter plasmid | | |
| CCR5 | F AATTCATGACATCAATTATTATACATCGGAGGAG | 3 |
| | R GATCCTCCTCCGATGTATAATAATTGATGTCATG | 4 |
| Primers used in the T7E1 assay | | |
| CCR5 | F1 CTCCATGGTGCTATAGAGCA | 5 |
| | F2 GAGCCAAGCTCTCCATCTAGT | 6 |
| | R GCCCTGTCAAGAGTTGACAC | 7 |
| C4BPB | F1 TATTTGGCTGGTTGAAAGGG | 8 |
| | R1 AAAGTCATGAAATAAACACACCCA | 9 |
| | F2 CTGCATTGATATGGTAGTACCATG | 10 |
| | R2 GCTGTTTCATGCAATGGAATG | 11 |
| Primers used for the amplification of off-target sites | | |
| ADCY5 | F1 GCTCCCACCTTAGTGCTCTG | 12 |
| | R1 GGTGGCAGGAACCTGTATGT | 13 |
| | F2 GTCATTGGCCAGAGATGTGGA | 14 |
| | R2 GTCCCATGACAGGCGTGAT | 15 |
| | F GCCTGGCCAAGTTTCAGTTA | 16 |
| KCNJ6 | R1 TGGAGCCATTGGTTTGCATC | 17 |
| | R2 CCAGAACTAAGCCGTTTCTGAC | 18 |
| | F1 ATCACCACCAACAGTTTCC | 19 |
| CNTNAP2 | F2 TGCAGTGCAGACTCTTTCCA | 20 |
| | R AAGGACACAGGGCAACTGAA | 21 |
| | F1 TGTGGAACGAGTGGTGACAG | 22 |
| N/A Chr. 5 | R1 GCTGGATTAGGAGGCAGGATTC | 23 |
| | F2 GTGCTGAGAACGTTTCATAGAG | 24 |
| | R2 GGACCAAACACATTCTTCTCAC | 25 |
| | Primers used for the detection of chromosomal deletions | |
| Deletion | F CCACATCTCGTTCTCGGTTT | 26 |
| | R TCACAAGCCCACAGATATT | 27 |

1-3. Targeted Disruption of Endogenous Genes in Mammalian Cells by RGEN

To test whether RGENs could be used for targeted disruption of endogenous genes in mammalian cells, genomic DNA isolated from transfected cells using T7 endonuclease I (T7E1), a mismatch-sensitive endonuclease that specifically recognizes and cleaves heteroduplexes formed by the hybridization of wild-type and mutant DNA sequences was analyzed (3).

To introduce DSBs in mammalian cells using RGENs, 2×10^6 K562 cells were transfected with 20 μ g of Cas9-encoding plasmid using the 4D-Nucleofector™, SF Cell Line 4D-Nucleofector® X Kit, Program FF-120 (Lonza) according to the manufacturer's protocol. For this experiment, K562 (ATCC, CCL-243) cells were grown in RPMI-1640 with 10% FBS and the penicillin/streptomycin mix (100 U/ml and 100 μ g/ml, respectively).

After 24 h, 10-40 μ g of in vitro transcribed chimeric RNA was nucleofected into 1×10^6 K562 cells. The in vitro transcribed chimeric RNA had been prepared as described in the Example 1-2.

Cells were collected two days after RNA transfection and genomic DNA was isolated. The region including the target site was PCR-amplified using the primers described in Table 1. The amplicons were subjected to the T7E1 assay as described previously (3). For sequencing analysis, PCR products corresponding to genomic modifications were purified and cloned into the T-Blunt vector using the T-Blunt PCR Cloning Kit (SolGent). Cloned products were sequenced using the M13 primer.

It was found that mutations were induced only when the cells were transfected serially with Cas9-encoding plasmid

and then with guide RNA (FIG. 3). Mutation frequencies (Indels (%) in FIG. 3A) estimated from the relative DNA band intensities were RNA-dosage dependent, ranging from 1.3% to 5.1%. DNA sequencing analysis of the PCR amplicons corroborated the induction of RGEN-mediated mutations at the endogenous sites. Indels and microhomologies, characteristic of error-prone NHEJ, were observed at the target site. The mutation frequency measured by direct sequencing was 7.3% (=7 mutant clones/96 clones), on par with those obtained with zinc finger nucleases (ZFNs) or transcription-activator-like effector nucleases (TALENs).

Serial-transfection of Cas9 plasmid and guide RNA was required to induce mutations in cells. But when plasmids that encode guide RNA, serial transfection was unnecessary and cells were co-transfected with Cas9 plasmid and guide RNA-encoding plasmid.

In the meantime, both ZFNs and TALENs have been successfully developed to disrupt the human CCR5 gene (3-6), which encodes a G-protein-coupled chemokine receptor, an essential co-receptor of HIV infection. A CCR5-specific ZEN is now under clinical investigation in the US for the treatment of AIDS (7). These ZFNs and TALENs, however, have off-target effects, inducing both local mutations at sites whose sequences are homologous to the on-target sequence (6, 8-10) and genome rearrangements that arise from the repair of two concurrent DSBs induced at on-target and off-target sites (11-12). The most striking off-target sites associated with these CCR5-specific engineered nucleases reside in the CCR2 locus, a close homolog of CCR5, located 15-kbp upstream of CCR5. To avoid off-target mutations in the CCR2 gene and unwanted deletions, inversions, and duplications of the 15-kbp chromo-

somal segment between the CCR5 on-target and CCR2 off-target sites, the present inventors intentionally chose the target site of our CCR5-specific RGEN to recognize a region within the CCR5 sequence that has no apparent homology with the CCR2 sequence.

The present inventors investigated whether the CCR5-specific RGEN had off-target effects. To this end, we searched for potential off-target sites in the human genome by identifying sites that are most homologous to the intended 23-bp target sequence. As expected, no such sites were found in the CCR2 gene. Instead, four sites, each of which carries 3-base mismatches with the on-target site, were found (FIG. 4A). The T7E1 assays showed that mutations were not detected at these sites (assay sensitivity, ~0.5%), demonstrating exquisite specificities of RGENS (FIG. 4B). Furthermore, PCR was used to detect the induction of chromosomal deletions in cells separately transfected with plasmids encoding the ZEN and RGEN specific to CCR5. Whereas the ZEN induced deletions, the RGEN did not (FIG. 4C).

Next, RGENs was reprogrammed by replacing the CCR5-specific guide RNA with a newly-synthesized RNA designed to target the human C4BPB gene, which encodes the beta chain of C4b-binding protein, a transcription factor. This RGEN induced mutations at the chromosomal target site in K562 cells at high frequencies (FIG. 3B). Mutation frequencies measured by the T7E1 assay and by direct sequencing were 14% and 8.3% (=4 mutant clones/48 clones), respectively. Out of four mutant sequences, two clones contained a single-base or two-base insertion precisely at the cleavage site, a pattern that was also observed at the CCR5 target site. These results indicate that RGENs cleave chromosomal target DNA at expected positions in cells.

Example 2: Proteinaceous RGEN-Mediated Genome Editing

RGENs can be delivered into cells in many different forms. RGENs consist of Cas9 protein, crRNA, and tracrRNA. The two RNAs can be fused to form a single-chain guide RNA (sgRNA). A plasmid that encodes Cas9 under a promoter such as CMV or CAG can be transfected into cells. crRNA, tracrRNA, or sgRNA can also be

regulation procedure before market approval in most developed countries. Furthermore, plasmid DNA can persist in cells for several days post-transfection, aggravating off-target effects of RGENS.

Here, we used recombinant Cas9 protein complexed with in vitro transcribed guide RNA to induce targeted disruption of endogenous genes in human cells. Recombinant Cas9 protein fused with the hexa-histidine tag was expressed in and purified from *E. coli* using standard Ni ion affinity chromatography and gel filtration. Purified recombinant Cas9 protein was concentrated in storage buffer (20 mM HEPES pH 7.5, 150 mM KCl, 1 mM DTT, and 10% glycerol). Cas9 protein/sgRNA complex was introduced directly into K562 cells by nucleofection: 1×10^6 K562 cells were transfected with 22.5-225 (1.4-14 μ M) of Cas9 protein mixed with 100 μ g (29 μ M) of in vitro transcribed sgRNA (or crRNA 40 μ g and tracrRNA 80 μ g) in 100 μ l solution using the 4D-Nucleofector, SF Cell Line 4D-Nucleofector X Kit, Program FF-120 (Lonza) according to the manufacturer's protocol. After nucleofection, cells were placed in growth media in 6-well plates and incubated for 48 hr. When 2×10^5 K562 cells were transfected with $\frac{1}{5}$ scale-downed protocol, 4.5-45 μ g of Cas9 protein mixed with 6-60 μ g of in vitro transcribed sgRNA (or crRNA 8 μ g and tracrRNA 16 μ g) were used and nucleofected in 20 μ l solution. Nucleofected cells were then placed in growth media in 48-well plates. After 48 hr, cells were collected and genomic DNA was isolated. The genomic DNA region spanning the target site was PCR-amplified and subjected to the T7E1 assay.

As shown in FIG. 10, Cas9 protein/sgRNA complex induced targeted mutation at the CCR5 locus at frequencies that ranged from 4.8 to 38% in a sgRNA or Cas9 protein dose-dependent manner, on par with the frequency obtained with Cas9 plasmid transfection (45%). Cas9 protein/crRNA/tracrRNA complex was able to induce mutations at a frequency of 9.4%. Cas9 protein alone failed to induce mutations. When 2×10^5 cells were transfected with $\frac{1}{5}$ scale-downed doses of Cas9 protein and sgRNA, mutation frequencies at the CCR5 locus ranged from 2.7 to 57% in a dose-dependent manner, greater than that obtained with co-transfection of Cas9 plasmid and sgRNA plasmid (32%).

We also tested Cas9 protein/sgRNA complex that targets the ABCC11 gene and found that this complex induced indels at a frequency of 35%, demonstrating general utility of this method.

TABLE 2

| Sequences of guide RNA | | | | |
|------------------------|----------|--------------------------------------|--------|-----------|
| Target RNA type | | RNA sequence (5' to 3') | Length | SEQ ID NO |
| CCR5 | sgRNA | GGUGACAUCAUUUUUUUAUCAUGUUUUUAGAGCUAG | 104 bp | 28 |
| | | AAAUAGCAAGUUAAAAUAAGGCUAGUCCGUUAUCA | | |
| | | ACUUGAAAAAGUGGCCACCGAGUCGGUGCUUUUUUU | | |
| | crRNA | GGUGACAUCAUUUUUUUAUCAUGUUUUUAGAGCUAU | 44 bp | 29 |
| | | GCUGUUUUU | | |
| | tracrRNA | GGAACCAUUCAAAACAGCAUAGCAAGUUAAAAUAA | 86 bp | 30 |
| | | GGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCG | | |
| | | AGUCGGUGCUUUUUUU | | |

expressed in cells using plasmids that encode these RNAs. Use of plasmids, however, often results in integration of the whole or part of the plasmids in the host genome. The bacterial sequences incorporated in plasmid DNA can cause unwanted immune response in vivo. Cells transfected with plasmid for cell therapy or animals and plants derived from DNA-transfected cells must go through a costly and lengthy

Example 3: RNA-Guided Genome Editing in Mice

To examine the gene-targeting potential of RGENs in pronuclear (PN)-stage mouse embryos, the forkhead box N1 (Foxn1) gene, which is important for thymus development and keratinocyte differentiation (Nehls et al., 1996), and the protein kinase, DNA activated, catalytic polypeptide (Prkd)

gene, which encodes an enzyme critical for DNA DSB repair and recombination (Taccioli et al., 1998) were used.

To evaluate the genome-editing activity of the Foxn1-RGEN, we injected Cas9 mRNA (10-ng/ μ l solution) with various doses of the sgRNA (FIG. 5a) into the cytoplasm of PN-stage mouse embryos, and conducted T7 endonuclease I (T7E1) assays (Kim et al. 2009) using genomic DNAs obtained from in vitro cultivated embryos (FIG. 6a).

Alternatively, we directly injected the RGEN in the form of recombinant Cas9 protein (0.3 to 30 ng/ μ l) complexed with the two-fold molar excess of Foxn1-specific sgRNA (0.14 to 14 ng/ μ l) into the cytoplasm or pronucleus of one-cell mouse embryos, and analyzed mutations in the Foxn1 gene using in vitro cultivated embryos (FIG. 7).

Specifically, Cas9 mRNA and sgRNAs were synthesized in vitro from linear DNA templates using the mMACHINE mMACHINE T7 Ultra kit (Ambion) and MEGAscript T7 kit (Ambion), respectively, according to the manufacturers' instructions, and were diluted with appropriate amounts of diethyl pyrocarbonate (DEPC, Sigma)-treated injection buffer (0.25 mM EDTA, 10 mM Tris, pH 7.4). Templates for sgRNA synthesis were generated using oligonucleotides listed in Table 3. Recombinant Cas9 protein was obtained from ToolGen, Inc.

mothers, respectively. Female FVB/NTac mice (7-8 weeks old) were super-ovulated by intra-peritoneal injections of 5 IU pregnant mare serum gonadotropin (PMSG, Sigma) and 5 IU human chorionic gonadotropin (hCG, Sigma) at 48-hour intervals. The super-ovulated female mice were mated to FVB/NTac stud males, and fertilized embryos were collected from oviducts.

Cas9 mRNA and sgRNAs in M2 medium (Sigma) were injected into the cytoplasm of fertilized eggs with well-recognized pronuclei using a Piezo-driven micromanipulator (Prime Tech).

In the case of injection of recombinant Cas9 protein, the recombinant Cas9 protein: Foxn1-sgRNA complex was diluted with DEPC-treated injection buffer (0.25 mM EDTA, 10 mM Tris, pH 7.4) and injected into male pronuclei using a TransferMan NK2 micromanipulator and a FemtoJet® microinjector (Eppendorf).

The manipulated embryos were transferred into the oviducts of pseudopregnant foster mothers to produce live animals, or were cultivated in vitro for further analyses.

To screen F0 mice and in vitro cultivated mouse embryos with RGEN-induced mutations, T7E1 assays were performed as previously described using genomic DNA samples from tail biopsies and lysates of whole embryos (Cho et al., 2013).

TABLE 3

| RNA Name | Direction | Sequence (5' to 3') | SEQ ID NO |
|-------------------|-----------|--|-----------|
| Foxn1 #1 sgRNA | F | <u>GAAATTAATACGACTCACTATAGGCAGTCTGACG</u> TCACACTFCCGTTTTAGAGCTAGAAATAGCAAGT TAAAATAAGGCTAGTCCG | 31 |
| Foxn1 #2 sgRNA | F | <u>GAAATTAATACGACTCACTATAGGACTTCCAGGC</u> TCCACCCGACGTTTTAGAGCTAGAAATAGCAAGT TAAAATAAGGCTAGTCCG | 32 |
| Foxn1 #3 sgRNA | F | <u>GAAATTAATACGACTCACTATAGGCCAGGCTCCA</u> CCCGACTGGAGTTTTAGAGCTAGAAATAGCAAGT TAAAATAAGGCTAGTCCG | 33 |
| Foxn1 #4 sgRNA | F | <u>GAAATTAATACGACTCACTATAGGACTGGAGGGC</u> GAAACCCCAAGGTTTTAGAGCTAGAAATAGCAAGT TAAAATAAGGCTAGTCCG | 34 |
| Foxn1 #5 sgRNA | F | <u>GAAATTAATACGACTCACTATAGGACCCCAAGGG</u> GACCTCATGCGTTTTAGAGCTAGAAATAGCAAGT TAAAATAAGGCTAGTCCG | 35 |
| Prkdc #1 sgRNA | F | <u>GAAATTAATACGACTCACTATAGGTTAGTTTTTT</u> CCAGAGACTTGTTTTAGAGCTAGAAATAGCAAGT TAAAATAAGGCTAGTCCG | 36 |
| Prkdc #2 sgRNA | F | <u>GAAATTAATACGACTCACTATAGGTTGGTTTGCT</u> TGTGTTTATCGTTTTAGAGCTAGAAATAGCAAGT TAAAATAAGGCTAGTCCG | 37 |
| Prkdc #3 sgRNA | F | <u>GAAATTAATACGACTCACTATAGGCACAAGCAAA</u> CCAAAGTCTCGTTTTAGAGCTAGAAATAGCAAGT TAAAATAAGGCTAGTCCG | 38 |
| Prkdc #4 sgRNA | F | <u>GAAATTAATACGACTCACTATAGGCCTCAATGCT</u> AAGCGACTTCGTTTTAGAGCTAGAAATAGCAAGT TAAAATAAGGCTAGTCCG | 39 |

All animal experiments were performed in accordance with the Korean Food and Drug Administration (KFDA) guidelines. Protocols were reviewed and approved by the Institutional Animal Care and Use Committees (IACUC) of the Laboratory Animal Research Center at Yonsei University (Permit Number: 2013-0099). All mice were maintained in the specific pathogen-free facility of the Yonsei Laboratory Animal Research Center. FVB/NTac (Taconic) and ICR mouse strains were used as embryo donors and foster

Briefly, the genomic region encompassing the RGEN target site was PCR-amplified, melted, and re-annealed to form heteroduplex DNA, which was treated with T7 endonuclease 1 (New England Biolabs), and then analyzed by agarose gel electrophoresis. Potential off-target sites were identified by searching with bowtie 0.12.9 and were also similarly monitored by T7E1 assays. The primer pairs used in these assays were listed in Tables 4 and 5.

TABLE 4

| Primers used in the T7E1 assay | | | |
|--------------------------------|-----------|----------------------------------|-----------|
| Gene | Direction | Sequence(5' to 3') | SEQ ID NO |
| Foxn1 | F1 | GTCTGTCTATCATCTCTCCCTTCTCTCC | 40 |
| | F2 | TCCCTAATCCGATGGCTAGCTCCAG | 41 |
| | R1 | ACGAGCAGCTGAAGTTAGCATGC | 42 |
| | R2 | CTACTCAATGCTCTTAGAGCTACCAGGCTTGC | 43 |
| Prkdc | F | GACTGTTGTGGGGAGGGCCG | 44 |
| | F2 | GGGAGGGCCGAAAGTCTTATTTTG | 45 |
| | R1 | CCTGAGACTGAAGTTGGCAGAAGTGAG | 46 |
| | R2 | CTTAGGGCTTCTTCTACAATCACG | 47 |

TABLE 5

| Primers used for amplification of off-target sites | | | | | |
|--|----------|-----------|----------------------------------|------------------------|----|
| Gene | Notation | Direction | Sequence(5' to 3') | SEQ ID NO | |
| Foxn1 | off 1 | F | CTCGTGTGTAGCCCTGAC | 48 | |
| | | R | AGACTGGCCCTGGAACCTCACAG | 49 | |
| | off 2 | F | CACTAAAGCCTGTCAGGAAGCCG | 50 | |
| | | R | CTGTGGAGAGCACACAGCAGC | 51 | |
| | off 3 | F | GCTGCGACCTGAGACCATG | 52 | |
| | | R | CTTCAATGGCTTCTGCTTAGGCTAC | 53 | |
| | off 4 | F | GGTTCAGATGAGCCATCCTTTC | 54 | |
| | | R | CCTGATCTGCAGGCTTAACCCCTG | 55 | |
| | Prkdc | off 1 | F | CTCACCTGCACATCACATGTGG | 56 |
| | | | R | GGCATCCACCCTATGGGGTC | 57 |
| off 2 | | F | GCCTTGACCTAGAGCTTAAAGAGCC | 58 | |
| | | R | GGTCTTGTAGCAGGAAGGACACTG | 59 | |
| off 3 | | F | AAAACCTCTGCTTGATGGGATATGTGGG | 60 | |
| | | R | CTCTCACTGGTTATCTGTGCTCCTTC | 61 | |
| off 4 | | F | GGATCAATAGGTGGTGGGGGATG | 62 | |
| | | R | GTGAATGACACAATGTGACAGCTTCAG | 63 | |
| off 5 | | F | CACAAGACAGACCTCTCAACATTCAGTC | 64 | |
| | | R | GTGCATGCATATAATCCATTCTGATTGCTCTC | 65 | |
| off 6 | | F1 | GGGAGGCAGAGGCAGGT | 66 | |
| | | F2 | GGATCTCTGTGAGTTTGAGGCCA | 67 | |
| | | R1 | GCTCCAGAACTCACTCTTAGGCTC | 68 | |

Mutant founders identified by the T7E1 assay were further analyzed by fPCR. Appropriate regions of genomic DNA were sequenced as described previously (Sung et al., 2013). For routine PCR genotyping of F1 progenies, the following primer pairs were used for both wild-type and mutant alleles:

(SEQ ID NO: 69)
5'-CTACTCCCTCCGAGTCTGA-3'
and

(SEQ ID NO: 70)
5'-CCAGGCCCTAGGTTCCAGGTA-3'
for the Foxn1 gene,

(SEQ ID NO: 71)
5'-CCCCAGCATTGCAGATTTCC-3'
and

-continued

(SEQ ID NO: 72)
5'-AGGGCTTCTTCTCTACAATCACG-3'

for Prkdc gene.

In the case of injection of Cas9 mRNA, mutant fractions (the number of mutant embryos/the number of total embryos) were dose-dependent, ranging from 33% (1 ng/ μ l sgRNA) to 91% (100 ng/ μ l) (FIG. 6b). Sequence analysis confirmed mutations in the Foxn1 gene; most mutations were small deletions (FIG. 6c), reminiscent of those induced by ZFNs and TALENS (Kim et al., 2013).

In the case of injection of Cas9 protein, these injection doses and methods minimally affected the survival and development of mouse embryos in vitro: over 70% of

50 RGEN-injected embryos hatched out normally in both experiments. Again, mutant fractions obtained with Cas9 protein injection were dose-dependent, and reached up to 88% at the highest dose via pronucleus injection and to 71% via intra-cytoplasmic injection (FIGS. 7a and 7b). Similar to the mutation patterns induced by Cas9 mRNA plus sgRNA (FIG. 6c), those induced by the Cas9 protein-sgRNA complex were mostly small deletions (FIG. 7c). These results clearly demonstrate that RGENs have high gene-targeting activity in mouse embryos.

60 Encouraged by the high mutant frequencies and low cytotoxicity induced by RGENs, we produced live animals by transferring the mouse embryos into the oviducts of pseudo-pregnant foster mothers.

65 Notably, the birth rates were very high, ranging from 58% to 738, and were not affected by the increasing doses of Foxn1-sgRNA (Table 6).

TABLE 6

| RGEN-mediated gene-targeting in FVB/NTac mice | | | | | | |
|---|---------------------------|------------------|-------------------------|--------------------|---------------------|----------------|
| Target Gene | Cas9 mRNA + sgRNA (ng/μl) | Injected embryos | Transferred embryos (%) | Total newborns (%) | Live newborns * (%) | Founders † (%) |
| Foxn1 | 10 + 1 | 76 | 62 (82) | 45 (73) | 31 (50) | 12 (39) |
| | 10 + 10 | 104 | 90 (87) | 52 (58) | 58 (64) | 33 (57) |
| | 10 + 100 | 100 | 90 (90) | 62 (69) | 58 (64) | 54 (93) |
| | Total | 280 | 242 (86) | 159 (66) | 147 (61) | 99 (67) |
| Prkdc | 50 + 50 | 73 | 58 (79) | 35 (60) | 33 (57) | 11 (33) |
| | 50 + 100 | 79 | 59 (75) | 22 (37) | 21 (36) | 7 (33) |
| | 50 + 250 | 94 | 73 (78) | 37 (51) | 37 (51) | 21 (57) |
| | Total | 246 | 190 (77) | 94 (49) | 91 (48) | 39 (43) |

Out of 147 newborns, we obtained 99 mutant founder mice. Consistent with the results observed in cultivated embryos (FIG. 6b), mutant fractions were proportional to the doses of Foxn1-sgRNA, and reached up to 93% (100 ng/μl Foxn1-sgRNA) (Tables 6 and 7, FIG. 5b).

TABLE 7

| DNA sequences of Foxn1 mutant alleles identified from a subset of T7E1-positive mutant founders | | | |
|---|-----------|---|-------------------------------|
| Sequence | Δ | # | Founder mice |
| ACTTCCAGGCTCCACCCGACTGGAGGGCGAACCCTCAAGGGGACCTCATGCAGG (SEQ ID NO: 134) | del + ins | | |
| ACTTCCAGGC-----AACCCTCAAGGGGACCTCATGCAGG (SEQ ID NO: 297) | Δ19 | 1 | 20 |
| ACTTCCAGGC-----GAACCCTCAAGGGGACCTCATGCAGG (SEQ ID NO: 298) | Δ18 | 1 | 115 |
| ACTTCCAGGCTCC-----Δ60 (SEQ ID NO: 299) | Δ60 | 1 | 19 |
| ACTTCCAGGCTCC-----Δ44 (SEQ ID NO: 300) | Δ44 | 1 | 108 |
| ACTTCCAGGCTCC-----CAAGGGGACCTCATGCAGG (SEQ ID NO: 301) | Δ21 | 1 | 64 |
| ACTTCCAGGCTCC-----TTAGGAGGCGAACCCTCAAGGGGACCTCA (SEQ ID NO: 302) | Δ12 + 6 | 1 | 126 |
| ACTTCCAGGCTCCACC-----TCATGCAGG (SEQ ID NO: 303) | Δ28 | 1 | 5 |
| ACTTCCAGGCTCCACCC-----CCAAGGGACCTCATG (SEQ ID NO: 304) | Δ21 + 4 | 1 | 61 |
| ACTTCCAGGCTCCACCC-----AAGGGGACCTCATGCAGG (SEQ ID NO: 305) | Δ18 | 2 | 95, 29 |
| ACTTCCAGGCTCCACCC-----CAAGGGGACCTCATGCAGG (SEQ ID NO: 306) | Δ17 | 7 | 12, 14, 27, 66, 108, 114, 126 |
| ACTTCCAGGCTCCACCC-----ACCCAAGGGGACCTCATGCAG (SEQ ID NO: 307) | Δ15 + 1 | 1 | 32 |
| ACTTCCAGGCTCCACCC-----CACCCAAGGGGACCTCATGCA (SEQ ID NO: 308) | Δ15 + 2 | 1 | 124 |
| ACTTCCAGGCTCCACCC-----ACCCAAGGGGACCTCATGCAGG (SEQ ID NO: 309) | Δ13 | 1 | 32 |
| ACTTCCAGGCTCCACCC-----GGCGAACCCTCAAGGGGACCTCATGCAGG (SEQ ID NO: 310) | Δ8 | 1 | 110 |
| ACTTCCAGGCTCCACCT-----GGGGACCTCATGCAGG (SEQ ID NO: 311) | Δ20 + 1 | 1 | 29 |

TABLE 7-continued

| DNA sequences of Foxn1 mutant alleles identified from a subset of T7E1-positive mutant founders | | | |
|---|---------|----|--|
| ACTTCCAGGCTCCACCCG-----AACCCCAAGGGGACCTCATGCAGG (SEQ ID NO: 312) | Δ11 | 1 | 111 |
| ACTTCCAGGCTCCACCCGA-----ACCTCATGCAGG (SEQ ID NO: 313) | Δ22 | 1 | 79 |
| ACTTCCAGGCTCCACCCGA-----GGGGACCTCATGCAGG (SEQ ID NO: 314) | Δ18 | 2 | 13, 127 |
| ACTTCCAGGCTCCACCCCA-----AGGGGACCTCATGCAGG (SEQ ID NO: 315) | Δ17 | 1 | 24 |
| ACTTCCAGGCTCCACCCGA-----ACCCCAAGGGGACCTCATGCAGG (SEQ ID NO: 316) | Δ11 | 5 | 14, 53, 58, 69, 124 |
| ACTTCCAGGCTCCACCCGA-----GACCCCAAGGGGACCTCATGCAGG (SEQ ID NO: 317) | Δ10 | 1 | 14 |
| ACTTCCAGGCTCCACCCGA----GGGCGAACCCCAAGGGGACCTCATGCAGG (SEQ ID NO: 318) | Δ5 | 3 | 53, 79, 115 |
| ACTTCCAGGCTCCACCCGAC-----CTCATGCAGG (SEQ ID NO: 319) | Δ23 | 1 | 108 |
| ACTTCCAGGCTCCACCCGAC-----CCCAAGGGGACCTCATGCAGG (SEQ ID NO: 320) | Δ11 | 1 | 3 |
| ACTTCCAGGCTCCACCCGAC-----GAAGGGCCCCAAGGGGACCTCA (SEQ ID NO: 321) | Δ11 + 6 | 1 | 66 |
| ACTTCCAGGCTCCACCCGAC-----GAACCCCAAGGGGACCTCATGCAGG (SEQ ID NO: 322) | Δ8 | 2 | 3, 66 |
| ACTTCCAGGCTCCACCCGAC----GGGCGAACCCCAAGGGGACCTCATGCAGG (SEQ ID NO: 323) | Δ5 | 1 | 27 |
| ACTTCCAGGCTCCACCCGAC--GTGCTTGAGGGCGAACCCCAAGGGGACCTCA (SEQ ID NO: 324) | Δ2 + 6 | 2 | 5 |
| ACTTCCAGGCTCCACCCGACT-----CACTATCTTCTGGGCTCCTCCATGTC (SEQ ID NO: 325) | Δ6 + 25 | 2 | 21, 114 |
| ACTTCCAGGCTCCACCCGACT----TGGCGAACCCCAAGGGGACCTCATGCAG (SEQ ID NO: 326) | Δ4 + 1 | 1 | 53 |
| ACTTCCAGGCTCCACCCGACT--TGCAGGGCGAACCCCAAGGGGACCTCATGC (SEQ ID NO: 327) | Δ2 + 3 | 1 | 126 |
| ACTTCCAGGCTCCACCCGACTTGGAGGGCGAACCCCAAGGGGACCTCATGCAG (SEQ ID NO: 328) | +1 | 15 | 3, 5, 12, 19, 29, 55, 56, 61, 66, 68, 81, 108, 111, 124, 127 |
| ACTTCCAGGCTCCACCCGACTTTGGAGGGCGAACCCCAAGGGGACCTCATGCA (SEQ ID NO: 329) | +2 | 2 | 79, 120 |
| ACTTCCAGGCTCCACCCGACTGTTGGAGGGCGAACCCCAAGGGGACCTCATGC (SEQ ID NO: 330) | +3 | 1 | 55 |
| ACTTCCAGGCTCCACCCGACTGGAG (+455) GCGAACCCCAAGGGGACCTCC (SEQ ID NO: 331) | +455 | 1 | 13 |

To generate Prkdc-targeted mice, we applied a 5-fold higher concentration of Cas9 mRNA (50 ng/μl) with increasing doses of Prkdc-sgRNA (50, 100, and 250 ng/μl). Again, the birth rates were very high, ranging from 51% to 60%, enough to produce a sufficient number of newborns for the analysis (Table 6). The mutant fraction was 57% (21 mutant founders among 37 newborns) at the maximum dose of

⁶⁰ Prkdc-sgRNA. These birth rates obtained with RGENs were approximately 2- to 10-fold higher than those with TALENs reported in our previous study (Sung et al., 2013). These results demonstrate that RGENs are potent gene-targeting reagents with minimal toxicity.

⁶⁵ To test the germ-line transmission of the mutant alleles, we crossed the Foxn1 mutant founder #108, a mosaic with

four different alleles (FIG. 5c, and Table 8) with wild-type mice, and monitored the genotypes of F1 offspring.

TABLE 8

| Genotypes of Foxn1 mutant mice | | | |
|--------------------------------|---------------|---------------------------|---|
| Founder NO. | sgRNA (ng/ml) | Genotyping Summary | Detected alleles |
| 58* | 1 | not determined | $\Delta 11$ |
| 19 | 100 | bi-allelic | $\underline{\Delta 60/+1}$ |
| 20 | 100 | bi-allelic | $\underline{\Delta 67/\Delta 19}$ |
| 13 | 100 | bi-allelic | $\underline{\Delta 18/+455}$ |
| 32 | 10 | bi-allelic (heterozygote) | $\underline{\Delta 13/\Delta 15+1}$ |
| 115 | 10 | bi-allelic (heterozygote) | $\underline{\Delta 18/\Delta 5}$ |
| 111 | 10 | bi-allelic (heterozygote) | $\underline{\Delta 11/+1}$ |
| 110 | 10 | bi-allelic (homozygote) | $\underline{\Delta 8/\Delta 8}$ |
| 120 | 10 | bi-allelic (homozygote) | $\underline{+2/+2}$ |
| 81 | 100 | heterozygote | $\underline{+1/WT}$ |
| 69 | 100 | homozygote | $\underline{\Delta 11/\Delta 11}$ |
| 55 | 1 | mosaic | $\underline{\Delta 18/\Delta 1/+1/+3}$ |
| 56 | 1 | mosaic | $\underline{\Delta 127/\Delta 41/\Delta 2/+1}$ |
| 127 | 1 | mosaic | $\underline{\Delta 18/+1/WT}$ |
| 53 | 1 | mosaic | $\underline{\Delta 11/\Delta 5/\Delta 4+1/WT}$ |
| 27 | 10 | mosaic | $\underline{\Delta 17/\Delta 5/WT}$ |
| 29 | 10 | mosaic | $\underline{\Delta 18/\Delta 20+1/+1}$ |
| 95 | 10 | mosaic | $\underline{\Delta 18/\Delta 14/\Delta 8/\Delta 4}$ |
| 108 | 10 | mosaic | $\underline{+1/\Delta 17/\Delta 23/\Delta 44}$ |
| 114 | 10 | mosaic | $\underline{\Delta 17/\Delta 8/\Delta 6+25}$ |
| 124 | 10 | mosaic | $\underline{\Delta 11/\Delta 15+2/+1}$ |
| 126 | 10 | mosaic | $\underline{\Delta 17/\Delta 2+3/\Delta 12+6}$ |
| 12 | 100 | mosaic | $\underline{\Delta 30/\Delta 28/\Delta 17/+1}$ |
| 5 | 100 | mosaic | $\underline{\Delta 28/\Delta 11/\Delta 2+6/+1}$ |
| 14 | 100 | mosaic | $\underline{\Delta 17/\Delta 11/\Delta 10}$ |
| 21 | 100 | mosaic | $\underline{\Delta 127/\Delta 41/\Delta 2/\Delta 6+25}$ |
| 24 | 100 | mosaic | $\underline{\Delta 17/+1/WT}$ |
| 64 | 100 | mosaic | $\underline{\Delta 31/\Delta 21/+1/WT}$ |
| 68 | 100 | mosaic | $\underline{\Delta 17/\Delta 11/+1/WT}$ |
| 79 | 100 | mosaic | $\underline{\Delta 22/\Delta 5/+2/WT}$ |
| 61 | 100 | mosaic | $\underline{\Delta 21+4/\Delta 6/+1/+9}$ |

TABLE 8-continued

| Genotypes of Foxn1 mutant mice | | | |
|--------------------------------|---------------|--------------------|--|
| Founder NO. | sgRNA (ng/ml) | Genotyping Summary | Detected alleles |
| 66** | 100 | mosaic | $\underline{\Delta 17/\Delta 8/\Delta 11+6/+1/WT}$ |
| 3 | 100 | mosaic | $\underline{\Delta 11/\Delta 8/+1}$ |

Underlined alleles were sequenced.
 Alleles in red, detected by sequencing, but not by fPCR.
 *only one clone sequenced.
 **Not determined by fPCR.

As expected, all the progenies were heterozygous mutants possessing the wild-type allele and one of the mutant alleles (FIG. 5d). We also confirmed the germ-line transmission in independent founder mice of Foxn1 (FIG. 8) and Prkdc (FIG. 9). To the best of our knowledge, these results provide the first evidence that RGEN-induced mutant alleles are stably transmitted to F1 progenies in animals.

Example 4: RNA-Guided Genome Editing in Plants

4-1. Production of Cas9 Protein

The Cas9 coding sequence (4104 bps), derived from *Streptococcus pyogenes* strain M1 GAS (NC_002737.1), was cloned to pET28-b(+) plasmid. A nuclear targeting sequence (NLS) was included at the protein N terminus to ensure the localization of the protein to the nucleus. pET28-b(+) plasmid containing Cas9 ORF was transformed into BL21(DE3). Cas9 was then induced using 0.2 mM IPTG for 16 hrs at 18° C. and purified using Ni-NTA agarose beads (Qiagen) following the manufacturer's instructions. Purified Cas9 protein was concentrated using Ultracel—100K (Millipore).

4-2. Production of Guide RNA

The genomic sequence of the Arabidopsis gene encoding the BRI1 was screened for the presence of a NGG motif, the so called protospacer adjacent motif (PAM), in an exon which is required for Cas9 targeting. To disrupt the BRI1 gene in Arabidopsis, we identified two RGEN target sites in an exon that contain the NGG motif. sgRNAs were produced in vitro using template DNA. Each template DNA was generated by extension with two partially overlapped oligonucleotides (Macrogen, Table X1) and Phusion polymerase (Thermo Scientific) using the following conditions—98° C. 30 sec {98° C. 10 sec, 54° C. 20 sec, 72° C. 2 min}×20, 72° C. 5 min.

TABLE 9

| Oligonucleotides for the production of the template DNA for in vitro transcription | | |
|--|---|-----------|
| Oligonucleotides | Sequence (5'-3') | SEQ ID NO |
| BRI1 target 1 (Forward) | GAAATTAATACGACTCACTATAGGTTTGAAAGATGG AAGCGCGGGTTTAGAGCTAGAAATAGCAAGTTAAA ATAAGGCTAGTCCG | 73 |
| BRI1 target 2 (Forward) | GAAATTAATACGACTCACTATAGGTGAACTAACT GGTCCACAGTTTTAGAGCTAGAAATAGCAAGTTAAA ATAAGGCTAGTCCG | 74 |
| Universal (Reverse) | AAAAAAGCACCGACTCGGTGCCACTTTTTCAAGTTG ATAACGGACTAGCCTTATTTAACTTGC | 75 |

The extended DNA was purified and used as a template for the in vitro production of the guide RNA's using the MEGAshortscript T7 kit (Life Technologies). Guide RNA were then purified by Phenol/Chloroform extraction and ethanol precipitation. To prepare Cas9/sgRNA complexes, 10 μ l of purified Cas9 protein (12 μ g/ μ l) and 4 μ l each of two sgRNAs (11 μ g/ μ l) were mixed in 20 μ l NEB3 buffer (New England Biolabs) and incubated for 10 min at 37° C.

4-3. Transfection of Cas9/sgRNA Complex to Protoplast

The leaves of 4-week-old *Arabidopsis* seedlings grown aseptically in petri dishes were digested in enzyme solution (1% cellulose R10, 0.5% macerozyme R10, 450 mM mannitol, 20 mM MES pH 5.7 and CPW salt) for 8–16 hrs at 25° C. with 40 rpm shaking in the dark. Enzyme/protoplast solutions were filtered and centrifuged at 100 \times g for 3–5 min. Protoplasts were re-suspended in CPW solution after counting cells under the microscope (\times 100) using a hemacytometer. Finally, protoplasts were re-suspended at 1×10^6 /ml in MMG solution (4 mM HEPES pH 5.7, 400 mM mannitol and 15 mM MgCl₂). To transfect the protoplasts with Cas9/sgRNA complex, 200 μ l (200,000 protoplasts) of the protoplast suspension were gently mixed with 3.3 or 10 μ l of Cas9/sgRNA complex [Cas9 protein (6 μ g/ μ l) and two sgRNAs (2.2 μ g/ μ l each)] and 200 μ l of 40% polyethylene glycol transfection buffer (40% PEG4000, 200 mM mannitol and 100 mM CaCl₂) in 2 ml tubes. After 5–20 min incubation at room temperature, transfection was stopped by adding wash buffer with W5 solution (2 mM MES pH 5.7, 154 mM NaCl, 125 mM CaCl₂ and 5 mM KCl). Protoplasts were then collected by centrifugation for 5 min at 100 \times g, washed with 1 ml of W5 solution, centrifuged for another 5 min at 100 \times g. The density of protoplasts was adjusted to 1×10^5 /ml and they were cultured in modified KM 8p liquid medium with 400 mM glucose.

4-4. Detection of Mutations in *Arabidopsis* Protoplasts and Plants

After 24 hr or 72 hr post-transfection, protoplasts were collected and genomic DNA was isolated. The genomic DNA region spanning the two target sites was PCR-amplified and subjected to the T7E1 assay. As shown in FIG. 11, indels were induced by RGENs at high frequencies that ranged from 50% to 70%. Surprisingly, mutations were induced at 24 hr post-transfection. Apparently, Cas9 protein functions immediately after transfection. PCR products were purified and cloned into T-Blunt PCR Cloning Kit (Solgent). Plasmids were purified and subjected to Sanger sequencing with M13F primer. One mutant sequence had a 7-bp deletion at one site (FIG. 12). The other three mutant sequences had deletions of ~220-bp DNA segments between the two RGEN site.

Example 5: Cas9 Protein Transduction Using a Cell-Penetrating Peptide or Protein Transduction Domain

5-1. Construction of his-Cas9-Encoding Plasmid

Cas9 with a cysteine at the C-terminal was prepared by PCR amplification using the previously described Cas9 plasmid {Cho, 2013 #166} as the template and cloned into pET28-(a) vector (Novagen, Merck Millipore, Germany) containing His-tag at the N-terminus.

5-2. Cell Culture

293T (Human embryonic kidney cell line), and HeLa (human ovarian cancer cell line) were grown in DMEM (GIBCO-BRL Rockville) supplemented with 10% FBS and 1% penicillin and streptomycin.

5-3. Expression and Purification of Cas9 Protein

To express the Cas9 protein, *E. coli* BL21 cells were transformed with the pET28-(a) vector encoding Cas9 and plated onto Luria-Bertani (LB) agar medium containing 50 μ g/mL kanamycin (Amresco, Solon, OH). Next day, a single colony was picked and cultured in LB broth containing 50 μ g/mL kanamycin at 37° C. overnight. Following day, this starter culture at 0.1 OD600 was inoculated into Luria broth containing 50 μ g/mL kanamycin and incubated for 2 hrs at 37° C. until OD600 reached to 0.6-0.8. To induce Cas9 protein expression, the cells were cultured at 30° C. overnight after addition of isopropyl- β -D-thiogalactopyranoside (IPTG) (Promega, Madison, WI) to the final concentration of 0.5 mM.

The cells were collected by centrifugation at 4000 rpm for 15-20 mins, resuspended in a lysis buffer (20 mM Tris-Cl pH8.0, 300 mM NaCl, 20 mM imidazole, 1 \times protease inhibitor cocktail, 1 mg/ml lysozyme), and lysed by sonication (40% duty, 10 sec pulse, 30 sec rest, for 10 mins on ice). The soluble fraction was separated as the supernatant after centrifugation at 15,000 rpm for 20 mins at 4° C. Cas9 protein was purified at 4° C. using a column containing Ni-NTA agarose resin (QIAGEN) and AKTA prime instrument (AKTA prime, GE Healthcare, UK). During this chromatography step, soluble protein fractions were loaded onto Ni-NTA agarose resin column (GE Healthcare, UK) at the flow rate of 1 mL/min. The column was washed with a washing buffer (20 mM Tris-Cl pH8.0, 300 mM NaCl, 20 mM imidazole, 1 \times protease inhibitor cocktail) and the bound protein was eluted at the flow rate of 0.5 ml/min with an elution buffer (20 mM Tris-Cl pH8.0, 300 mM NaCl, 250 mM imidazole, 1 \times protease inhibitor cocktail). The pooled eluted fraction was concentrated and dialyzed against storage buffer (50 mM Tris-HCl, pH8.0, 200 mM KCl, 0.1 mM EDTA, 1 mM DTT, 0.5 mM PMSF, 20% Glycerol). Protein concentration was quantitated by Bradford assay (Biorad, Hercules, CA) and purity was analyzed by SDS-PAGE using bovine serum albumin as the control.

5-4. Conjugation of Cas9 to 9R4L

1 mg Cas9 protein diluted in PBS at the concentration of 1 mg/mL and 50 μ g of maleimide-9R4L peptide in 25 μ l DW (Pepton, Korea) were gently mixed using a rotor at room temperature for 2 hrs and at 4° C. overnight. To remove unconjugated maleimide-9R4L, the samples were dialyzed using 50kDa molecular weight cutoff membrane against of DPBS (pH 7.4) at 4° C. for 24 hrs. Cas9-9R4L protein was collected from the dialysis membrane and the protein amount was determined using Bradford assay.

5-5. Preparation of sgRNA-9R4L

sgRNA (1 μ g) was gently added to various amounts of C9R4LC peptide (ranging from 1 to 40 weight ratio) in 100 μ l of DPBS (pH 7.4). This mixture was incubated at room temperature for 30 mins and diluted to 10 folds using RNase-free deionized water. The hydrodynamic diameter and z-potential of the formed nanoparticles were measured using dynamic light scattering (Zetasizer-nano analyzer ZS; Malvern instruments, Worcestershire, UK).

5-6. Cas9 Protein and sgRNA Treatments

Cas9-9R4L and sgRNA-C9R4LC were treated to the cells as follows: 1 μ g of sgRNA and 15 μ g of C9R4LC peptide were added to 250 mL of OPTIMEM medium and incubated at room temperature for 30 mins. At 24 hrs after seeding, cells were washed with OPTIMEM medium and treated with sgRNA-C9R4LC complex for 4 hrs at 37° C. Cells were washed again with OPTIMEM medium and treated with Cas9-9R4L for 2 hrs at 37° C. After treatment, culture media was replaced with serum-containing complete medium and incubated at 37° C. for 24 hrs before the next treatment.

Same procedure was followed for multiple treatments of Cas9 and sgRNA for three consecutive days.

5-7. Cas9-9R4L and sgRNA-9R4L Can Edit Endogenous Genes in Cultured Mammalian Cells Without the Use of Additional Delivery Tools

To determine whether Cas9-9R4L and sgRNA-9R4L can edit endogenous genes in cultured mammalian cells without the use of additional delivery tools, we treated 293 cells with Cas9-9R4L and sgRNA-9R4L targeting the CCR5 gene and analyzed the genomic DNA. T7E1 assay showed that 9% of CCR5 gene was disrupted in cells treated with both Cas9-9R4L and sgRNA-9R4L and that the CCR5 gene disruption was not observed in control cells including those untreated, treated with either Cas9-9R or sgRNA-9R4L, or treated with both unmodified Cas-9 and sgRNA (FIG. 13), suggesting that the treatment with Cas9-9R4L protein and sgRNA conjugated with 9R4L, but not unmodified Cas9 and sgRNA, can lead to efficient genome editing in mammalian cells.

Example 6: Control of Off-Target Mutation According to Guide RNA Structure

Recently, three groups reported that RGENs had off-target effects in human cells. To our surprise, RGENs induced mutations efficiently at off-target sites that differ by 3 to 5 nucleotides from on-target sites. We noticed, however, that there were several differences between our RGENs and those used by others. First, we used dualRNA, which is crRNA plus tracrRNA, rather than single-guide RNA (sgRNA) that is composed of essential portions of crRNA and tracrRNA. Second, we transfected K562 cells (but not HeLa cells) with synthetic crRNA rather than plasmids encoding crRNA. HeLa cells were transfected with crRNA-encoding plasmids. Other groups used sgRNA-encoding plasmids. Third, our guide RNA had two additional guanine nucleotides at the 5' end, which are required for efficient transcription by T7 polymerase in vitro. No such additional nucleotides were included in the sgRNA used by others. Thus, the RNA sequence of our guide RNA can be shown as 5'-GGX₂₀, whereas 5'-GX₁₉, in which X₂₀ or GX₁₉ corresponds to the 20-bp target sequence, represents the sequence used by others. The first guanine nucleotide is required for transcription by RNA polymerase in cells. To test whether off-target RGEN effects can be attributed to these differences, we chose four RGENs that induced off-target mutations in human cells at high frequencies (13). First, we compared our method of using in vitro transcribed dualRNA with the method of transfecting sgRNA-encoding plasmids in K562 cells and measured mutation frequencies at the on-target and off-target sites via the T7E1 assay. Three RGENs showed comparable mutation frequencies at on-target and off-target sites regardless of the composition of guide RNA. Interestingly, one RGEN (VEFGA site 1) did not induce indels at one validated off-target site, which differs by three nucleotides from the on-target site (termed OT1-11, FIG. 14), when synthetic dualRNA was used. But the synthetic dualRNA did not discriminate the other validated off-target site (OT1-3), which differs by two nucleotides from the on-target site.

Next, we tested whether the addition of two guanine nucleotides at the 5' end of sgRNA could make RGENs more specific by comparing 5'-GGX₂₀ (or 5'-GGGX₁₉) sgRNA with 5'-GX₁₉ sgRNA. Four GX₁₉ sgRNAs complexed with Cas9 induced indels equally efficiently at on-target and off-target sites, tolerating up to four nucleotide mismatches. In sharp contrast, GGX₂₀ sgRNAs discriminated off-target

sites effectively. In fact, the T7E1 assay barely detected RGEN-induced indels at six out of the seven validated off-target sites when we used the four GGX₂₀ sgRNAs (FIG. 15). We noticed, however, that two GGX₂₀ sgRNAs (VEFGA sites 1 and 3) were less active at on-target sites than were the corresponding GX₁₉ sgRNAs. These results show that the extra nucleotides at the 5' end can affect mutation frequencies at on-target and off-target sites, perhaps by altering guide RNA stability, concentration, or secondary structure.

These results suggest that three factors—the use of synthetic guide RNA rather than guide RNA-encoding plasmids, dualRNA rather than sgRNA, and GGX₂₀ sgRNA rather than GX₁₉ sgRNA—have cumulative effects on the discrimination of off-target sites.

Example 7: Paired Cas9 Nickases

In principle, single-strand breaks (SSBs) cannot be repaired by error-prone NHEJ but still trigger high fidelity homology-directed repair (HDR) or base excision repair. But nickase-induced targeted mutagenesis via HDR is much less efficient than is nuclease-induced mutagenesis. We reasoned that paired Cas9 nickases would produce composite DSBs, which trigger DNA repair via NHEJ or HDR, leading to efficient mutagenesis (FIG. 16A). Furthermore, paired nickases would double the specificity of Cas9-based genome editing.

We first tested several Cas9 nucleases and nickases designed to target sites in the AAVS1 locus (FIG. 16B) in vitro via fluorescent capillary electrophoresis. Unlike Cas9 nucleases that cleaved both strands of DNA substrates, Cas9 nickases composed of guide RNA and a mutant form of Cas9 in which a catalytic aspartate residue is changed to an alanine (D10A Cas9) cleaved only one strand, producing site-specific nicks (FIG. 16C, D). Interestingly, however, some nickases (AS1, AS2, AS3, and S6 in FIG. 17A) induced indels at target sites in human cells, suggesting that nicks can be converted to DSBs, albeit inefficiently, in vivo. Paired Cas9 nickases producing two adjacent nicks on opposite DNA strands yielded indels at frequencies that ranged from 14% to 91%, comparable to the effects of paired nucleases (FIG. 17A). The repair of two nicks that would produce 5' overhangs led to the formation of indels much more frequently than those producing 3' overhangs at three genomic loci (FIG. 17A and FIG. 18). In addition, paired nickases enabled targeted genome editing via homology-directed repair more efficiently than did single nickases (FIG. 19).

We next measured mutation frequencies of paired nickases and nucleases at off-target sites using deep sequencing. Cas9 nucleases complexed with three sgRNAs induced off-target mutations at six sites that differ by one or two nucleotides from their corresponding on-target sites with frequencies that ranged from 0.5% to 10% (FIG. 17B). In contrast, paired Cas9 nickases did not produce indels above the detection limit of 0.1% at any of the six off-target sites. The S2 Off-1 site that differs by a single nucleotide at the first position in the PAM (i.e., N in NGG) from its on-target site can be considered as another on-target site. As expected, the Cas9 nuclease complexed with the S2 sgRNA was equally efficient at this site and the on-target site. In sharp contrast, D10A Cas9 complexed with the S2 and AS2 sgRNAs discriminated this site from the on-target site by a factor of 270 fold. This paired nickase also discriminated the

AS2 off-target sites (Off-1 and Off-9 in FIG. 17B) from the on-target site by factors of 160 fold and 990 fold, respectively.

Example 8: Chromosomal DNA Splicing Induced by Paired Cas9 Nickases

Two concurrent DSBs produced by engineered nucleases such as ZENs and TALENs can promote large deletions of the intervening chromosomal segments has been reported. We tested whether two SSBs induced by paired Cas9 nickases can also produce deletions in human cells. We used PCR to detect deletion events and found that seven paired nickases induced deletions of up to 1.1-kbp chromosomal segments as efficiently as paired Cas9 nucleases did (FIG. 20A, B). DNA sequences of the PCR products confirmed the deletion events (FIG. 20C). Interestingly, the sgRNA-matching sequence remained intact in two out of seven deletion-specific PCR amplicons (underlined in FIG. 20C). In contrast, Cas9 nuclease pairs did not produce sequences that contained intact target sites. This finding suggests that two distant nicks were not converted to two separate DSBs to promote deletions of the intervening chromosomal segment. In addition, it is unlikely that two nicks separated by more than a 100 bp can produce a composite DSB with large overhangs under physiological conditions because the melting temperature is very high.

We propose that two distant nicks are repaired by strand displacement in a head-to-head direction, resulting in the formation of a DSB in the middle, whose repair via NHEJ causes small deletions (FIG. 20D). Because the two target sites remain intact during this process, nickases can induce SSBs again, triggering the cycle repeatedly until the target sites are deleted. This mechanism explains why two offset nicks producing 5' overhangs but not those producing 3' overhangs induced indels efficiently at three loci.

We then investigated whether Cas9 nucleases and nickases can induce unwanted chromosomal translocations that result from NHEJ repair of on-target and off-target DNA cleavages (FIG. 21A). We were able to detect translocations induced by Cas9 nucleases using PCR (FIG. 21B, C). No such PCR products were amplified using genomic DNA isolated from cells transfected with the plasmids encoding the AS2+S3 Cas9 nickase pair. This result is in line with the fact that both AS2 and S3 nickases, unlike their corresponding nucleases, did not produce indels at off-target sites (FIG. 17B).

These results suggest that paired Cas9 nickases allow targeted mutagenesis and large deletions of up to 1-kbp chromosomal segments in human cells. Importantly, paired nickases did not induce indels at off-target sites at which their corresponding nucleases induce mutations. Furthermore, unlike nucleases, paired nickases did not promote unwanted translocations associated with off-target DNA cleavages. In principle, paired nickases double the specificity of Cas9-mediated mutagenesis and will broaden the utility of RNA-guided enzymes in applications that require precise genome editing such as gene and cell therapy. One caveat to this approach is that two highly active sgRNAs are needed to make an efficient nickase pair, limiting targetable sites. As shown in this and other studies, not all sgRNAs are equally active. When single clones rather than populations of cells are used for further studies or applications, the choice of guide RNAs that represent unique sequences in the genome and the use of optimized guide RNAs would suffice to avoid off-target mutations associated with Cas9 nucleases.

We propose that both Cas9 nucleases and paired nickases are powerful options that will facilitate precision genome editing in cells and organisms.

Example 9: Genotyping with CRISPR/Cas-Derived RNA-Guided Endonucleases

Next, we reasoned that RGENs can be used in Restriction fragment length polymorphism (RFLP) analysis, replacing conventional restriction enzymes. Engineered nucleases including RGENs induce indels at target sites, when the DSBs caused by the nucleases are repaired by the error-prone non-homologous end-joining (NHEJ) system. RGENs that are designed to recognize the target sequences cannot cleave mutant sequences with indels but will cleave wild-type target sequences efficiently.

9-1. RGEN Components

crRNA and tracrRNA were prepared by in vitro transcription using MEGAshortscript T7 kit (Ambion) according to the manufacturer's instruction. Transcribed RNAs were resolved on a 8% denaturing urea-PAGE gel. The gel slice containing RNA was cut out and transferred to elution buffer. RNA was recovered in nuclease-free water followed by phenol: chloroform extraction, chloroform extraction, and ethanol precipitation. Purified RNA was quantified by spectrometry. Templates for crRNA were prepared by annealing an oligonucleotide whose sequence is shown as 5'-GAAATTAATACGACTCACTATAGGX₂₀GTTTTAGAGCTATGCTGTTTTG-3' (SEQ ID NO: 76), in which X₂₀ is the target sequence, and its complementary oligonucleotide. The template for tracrRNA was synthesized by extension of forward and reverse oligonucleotides (5'-GAAATTAATACGACTCACTATAGGAACCAATCAAAACAGCATAGCAAGTTAAAATAAGGCTA GTCCG-3' (SEQ ID NO: 77) and 5'-AAAAAAGCACCGACTCGGTGC-CACCTTTTCAAGTTGATAACGGACTAGCCTTATTT-TAACT TGCTATG-3' (SEQ ID NO: 78)) using Phusion polymerase (New England Biolabs).

9-2. Recombinant Cas9 Protein Purification

The Cas9 DNA construct used in our previous Example, which encodes Cas9 fused to the His6-tag at the C terminus, was inserted in the pET-28a expression vector. The recombinant Cas9 protein was expressed in *E. coli* strain BL21 (DE3) cultured in LB medium at 25° C. for 4 hours after induction with 1 mM IPTG. Cells were harvested and resuspended in buffer containing 20 mM Tris pH 8.0, 500 mM NaCl, 5 mM imidazole, and 1 mM PMSF. Cells were frozen in liquid nitrogen, thawed at 4° C., and sonicated. After centrifugation, the Cas9 protein in the lysate was bound to Ni-NTA agarose resin (Qiagen), washed with buffer containing 20 mM Tris pH 8.0, 500 mM NaCl, and 20 mM imidazole, and eluted with buffer containing 20 mM Tris pH 8.0, 500 mM NaCl, and 250 mM imidazole. Purified Cas9 protein was dialyzed against 20 mM HEPES (pH 7.5), 150 mM KCl, 1 mM DTT, and 10% glycerol and analyzed by SDS-PAGE.

9-3. T7 Endonuclease I Assay

The T7E1 assay was performed as following. In brief, PCR products amplified using genomic DNA were denatured at 95° C., reannealed at 16° C., and incubated with 5 units of T7 Endonuclease I (New England BioLabs) for 20 min at 37° C. The reaction products were resolved using 2 to 2.5% agarose gel electrophoresis.

9-4. RGEN-RFLP Assay

PCR products (100-150 ng) were incubated for 60 min at 37° C. with optimized concentrations (Table 10) of Cas9 protein, tracrRNA, crRNA in 10 µl NEB buffer 3 (1x). After

41

the cleavage reaction, RNase A (4 μ g) was added, and the reaction mixture was incubated for 30 min at 37° C. to remove RNA. Reactions were stopped with 6 \times stop solution buffer containing 30% glycerol, 1.2% SDS, and 100 mM EDTA. Products were resolved with 1-2.5% agarose gel electrophoresis and visualized with EtBr staining.

TABLE 10

| Concentration of RGEN components in RFLP assays | | | |
|---|--------------------|---------------------|------------------------|
| Target Name | Cas9 (ng/ μ l) | crRNA (ng/ μ l) | tracrRNA (ng/ μ l) |
| C4BPB | 100 | 25 | 60 |
| PIBF-NGG-RGEN | 100 | 25 | 60 |
| HLA-B | 1.2 | 0.3 | 0.7 |
| CCR5-ZFN | 100 | 25 | 60 |
| CTNNB1 Wild type specific | 30 | 10 | 20 |
| CTNNB1 mutant specific | 30 | 10 | 20 |
| CCR5 WT-specific | 100 | 25 | 60 |
| CCR5 Δ 32-specific | 10 | 2.5 | 6 |
| KRAS WT specific(wt) | 30 | 10 | 20 |
| KRAS mutant specific(m8) | 30 | 10 | 20 |

42

TABLE 10-continued

| Concentration of RGEN components in RFLP assays | | | |
|---|--------------------|---------------------|------------------------|
| Target Name | Cas9 (ng/ μ l) | crRNA (ng/ μ l) | tracrRNA (ng/ μ l) |
| KRAS WT specific (m6) | 30 | 10 | 20 |
| KRAS mutant specific (m6, 8) | 30 | 10 | 20 |
| PIK3CA WT specific (wt) | 100 | 25 | 60 |
| PIK3CA mutant specific(m4) | 30 | 10 | 20 |
| PIK3CA WT specific (m7) | 100 | 25 | 60 |
| PIK3CA mutant specific(m4, 7) | 30 | 10 | 20 |
| BRAF WT-specific | 30 | 10 | 20 |
| BRAF mutant-specific | 100 | 25 | 60 |
| NRAS WT-specific | 100 | 25 | 60 |
| NRAS mutant-specific | 30 | 10 | 20 |
| IDH WT-specific | 30 | 10 | 20 |
| IDH mutant-specific | 30 | 10 | 20 |
| PIBF-NAG-RGEN | 30 | 10 | 60 |

TABLE 11

| Primers | | | |
|----------------------|-----------|----------------------------------|-----------|
| Gene(site) | Direction | Sequence(5' to 3') | SEQ ID NO |
| CCR5 (RGEN) | F1 | CTCCATGGTGCTATAGAGCA | 79 |
| | F2 | GAGCCAAGCTCTCCATCTAGT | 80 |
| | R | GCCCTGTCAAGAGTTGACAC | 81 |
| CCR5 (ZFN) | F | GCACAGGGTGAACAAGATGGA | 82 |
| | R | GCCAGGTACCTATCGATTGTGAGG | 83 |
| CCR5 (Δ 132) | F | GAGCCAAGCTCTCCATCTAGT | 84 |
| | R | ACTCTGACTG GGTACCAGC | 85 |
| C4BPB | F1 | TATTTGGCTGGTTGAAAGGG | 86 |
| | R1 | AAAGTCATGAAATAAACACACCCA | 87 |
| | F2 | CTGCATTGATATGGTAGTACCATG | 88 |
| | R2 | GCTGTTCAATTGCAATGGAATG | 89 |
| CTNNB1 | F | ATGGAGTTGGACATGGCCATGG | 90 |
| | R | ACTCACTATCCACAGTTCAGCATTTACC | 91 |
| KRAS | F | TGGAGATAGCTGTCAGCAACTTT | 92 |
| | R | CAACAA AGCAAAGGTAAGTTGGTAATAG | 93 |
| PIK3CA | F | GGTTTCAGGAGATGTGTACAAGGC | 94 |
| | R | GATTGTGCAATTCCTATGCAATCGGTC | 95 |
| NRAS | F | CACTGGGTACTTAATCTGTAGCCTC | 96 |
| | R | GGTTCCAAGTCATTCCCAGTAGC | 97 |
| IDH1 | F | CATCACTGCAGTTGTAGGTTATAACTATCC | 98 |
| | R | TTGAAAACCAACAGATCTGGTTGAACC | 99 |
| BRAF | F | GGAGTGCCAAGAGAATATCTGG | 100 |
| | R | CTGAAACTGGTTTCAAAATATTCGTTTAAAGG | 101 |
| PIBF | F | GCTCTGTATGCCCTGTAGTAGG | 102 |
| | R | TTTGCATCTGACCTTACCTTTG | 103 |

9-5. Plasmid Cleavage Assay

Restriction enzyme-treated linearized plasmid (100 ng) was incubated for 60 min at 37° C. with Cas9 protein (0.1 µg), tracrRNA (60 ng), and crRNA (25 ng) in 10 µl NEB 3 buffer (1×). Reactions were stopped with 6× stop solution containing 30% glycerol, 1.2% SDS, and 100 mM EDTA. Products were resolved with 1% agarose gel electrophoresis and visualized with EtBr staining.

9-6. Strategy of RFLP

New RGENs with desired DNA specificities can be readily created by replacing crRNA; no de novo purification of custom proteins is required once recombinant Cas9 protein is available. Engineered nucleases, including RGENs, induce small insertions or deletions (indels) at target sites when the DSBs caused by the nucleases are repaired by error-prone non-homologous end-joining (NHEJ). RGENs that are designed to recognize the target sequences cleave wild-type sequences efficiently but cannot cleave mutant sequences with indels (FIG. 22).

We first tested whether RGENs can differentially cleave plasmids that contain wild-type or modified C4BPB target sequences that harbor 1- to 3-base indels at the cleavage site. None of the six plasmids with these indels were cleaved by a C4BPB-specific RGEN5 composed of target-specific crRNA, tracrRNA, and recombinant Cas9 protein (FIG. 23). In contrast, the plasmid with the intact target sequence was cleaved efficiently by this RGEN.

9-7. Detection of Mutations Induced by the Same RGENs Using RGEN-Mediated RFLP

Next, to test the feasibility of RGEN-mediated RFLP for detection of mutations induced by the same RGENs, we utilized gene-modified K562 human cancer cell clones established using an RGEN targeting C4BPB gene (Table 12).

TABLE 12

| Target sequence of RGENs used in this study | | |
|---|--------------------------------|-----------|
| Gene | Target sequence | SEQ ID NO |
| human C4BPB | <u>AATGACCACTACATCCTCAAGGG</u> | 104 |
| mouse Pibf1 | <u>AGATGATGTCTCATCATCAGAGG</u> | 105 |

C4BPB mutant clones used in this study have various mutations ranging from 94 bp deletion to 67 bp insertion (FIG. 24A). Importantly, all mutations occurred in mutant clones resulted in the loss of RGEN target site. Among 6 C4BPB clones analyzed, 4 clones have both wildtype and mutant alleles (+/-) and 2 clones have only mutant alleles (-/-).

The PCR products spanning the RGEN target site amplified from wildtype K562 genomic DNA were digested completely by the RGEN composed of target-specific crRNA, tracrRNA, and recombinant Cas9 protein expressed in and purified from *E. coli* (FIG. 24B/Lane 1). When the C4BPB mutant clones were subjected to RFLP analysis using the RGEN, PCR amplicons of +/- clones that contained both wildtype and mutant alleles were partially digested, and those of -/- clones that did not contain the wildtype allele were not digested at all, yielding no cleavage products corresponding to the wildtype sequence (FIG. 24B). Even a single-base insertion at the target site blocked the digestion (#12 and #28 clones) of amplified mutant alleles by the C4BPB RGEN, showing the high specificity of RGEN-mediated RFLP. We subjected the PCR amplicons to the mismatch-sensitive T7E1 assay in parallel (FIG. 24B).

Notably, the T7E1 assay was not able to distinguish -/- clones from +/- clones. To make it matters worse, the T7E1 assay cannot distinguish homozygous mutant clones that contain the same mutant sequence from wildtype clones, because annealing of the same mutant sequence will form a homoduplex. Thus, RGEN-mediated RFLP has a critical advantage over the conventional mismatch-sensitive nuclease assay in the analysis of mutant clones induced by engineered nucleases including ZENS, TALENS and RGENS.

9-8. Quantitative Assay for RGEN-RFLP Analysis

We also investigated whether RGEN-RFLP analysis is a quantitative method. Genomic DNA samples isolated from the C4BPB null clone and the wild-type cells were mixed at various ratios and used for PCR amplifications. The PCR products were subjected to RGEN genotyping and the T7E1 assay in parallel (FIG. 25b). As expected, DNA cleavage by the RGEN was proportional to the wild type to mutant ratio. In contrast, results of the T7E1 assay correlated poorly with mutation frequencies inferred from the ratios and were inaccurate, especially at high mutant %, a situation in which complementary mutant sequences can hybridize with each other to form homoduplexes.

9-9. Analysis of Mutant Mouse Founders Using a RGEN-Mediated RFLP Genotyping

We also applied RGEN-mediated RFLP genotyping (RGEN genotyping in short) to the analysis of mutant mouse founders that had been established by injection of TALENs into mouse one-cell embryos (FIG. 26A). We designed and used an RGEN that recognized the TALEN target site in the Pibf1 gene (Table 10). Genomic DNA was isolated from a wildtype mouse and mutant mice and subjected to RGEN genotyping after PCR amplification. RGEN genotyping successfully detected various mutations, which ranged from one to 27-bp deletions (FIG. 26B). Unlike the T7E1 assay, RGEN genotyping enabled differential detection of +/- and -/- founder.

9-10. Detection of Mutations Induced in Human Cells by a CCR5-Specific ZFN Using RGENS

In addition, we used RGENs to detect mutations induced in human cells by a CCR5-specific ZFN, representing yet another class of engineered nucleases (FIG. 27). These results show that RGENs can detect mutations induced by nucleases other than RGENs themselves. In fact, we expect that RGENs can be designed to detect mutations induced by most, if not all, engineered nucleases. The only limitation in the design of an RGEN genotyping assay is the requirement for the GG or AG (CC or CT on the complementary strand) dinucleotide in the PAM sequence recognized by the Cas9 protein, which occurs once per 4 bp on average. Indels induced anywhere within the seed region of several bases in crRNA and the PAM nucleotides are expected to disrupt RGEN-catalyzed DNA cleavage. Indeed, we identified at least one RGEN site in most (98%) of the ZEN and TALEN sites.

9-11. Detection of Polymorphisms or Variations Using RGEN

Next, we designed and tested a new RGEN that targets a highly polymorphic locus, HLA-B, that encodes Human Leukocyte Antigen B (a.k.a. MHC class I protein) (FIG. 28). HeLa cells were transfected with RGEN plasmids, and the genomic DNA was subjected to T7E1 and RGEN-RFLP analyses in parallel. T7E1 produced false positive bands that resulted from sequence polymorphisms near the target site (FIG. 25c). As expected, however, the same RGEN used for gene disruption cleaved PCR products from wild-type cells completely but those from RGEN-transfected cells partially,

indicating the presence of RGEN-induced indels at the target site. This result shows that RGEN-RFLP analysis has a clear advantage over the T7E1 assay, especially when it is not known whether target genes have polymorphisms or variations in cells of interest.

9-12. Detection of Recurrent Mutations Found in Cancer and Naturally-Occurring Polymorphisms Through RGEN-RFLP Analysis

RGEN-RFLP analysis has applications beyond genotyping of engineered nuclease-induced mutations. We sought to use RGEN genotyping to detect recurrent mutations found in cancer and naturally-occurring polymorphisms. We chose the human colorectal cancer cell line, HCT116, which carries a gain-of-function 3-bp deletion in the oncogenic CTNNB1 gene encoding beta-catenin. PCR products amplified from HCT116 genomic DNA were cleaved partially by both wild-type-specific and mutant-specific RGENs, in line with the heterozygous genotype in HCT116 cells (FIG. 29a). In sharp contrast, PCR products amplified from DNA from HeLa cells harboring only wild-type alleles were digested completely by the wild-type-specific RGEN and were not cleaved at all by the mutation-specific RGEN.

We also noted that HEK293 cells harbor the 32-bp deletion (del132) in the CCR5 gene, which encodes an essential co-receptor of HIV infection: Homozygous del32 CCR5 carriers are immune to HIV infection. We designed one RGEN specific to the del32 allele and the other to the wild-type allele. As expected, the wild-type-specific RGEN cleaved the PCR products obtained from K562, SKBR3, or HeLa cells (used as wild-type controls) completely but those from HEK293 cells partially (FIG. 30a), confirming the presence of the uncleavable del32 allele in HEK293 cells. Unexpectedly, however, the del32-specific RGEN cleaved the PCR products from wild-type cells as efficiently as those from HEK293 cells. Interestingly, this RGEN had an off-target site with a single-base mismatch immediately downstream of the on-target site (FIG. 30). These results suggest that RGENs can be used to detect naturally-occurring indels but cannot distinguish sequences with single nucleotide polymorphisms or point mutations due to their off-target effects.

To genotype oncogenic single-nucleotide variations using RGENs, we attenuated RGEN activity by employing a single-base mismatched guide RNA instead of a perfectly-matched RNA. RGENs that contained the perfectly-matched guide RNA specific to the wild-type sequence or mutant sequence cleaved both sequences (FIGS. 31a and 32a). In contrast, RGENs that contained a single-base mismatched guide RNA distinguished the two sequences, enabling genotyping of three recurrent oncogenic point mutations in the KRAS, PIK3CA, and IDH1 genes in human cancer cell lines (FIG. 29b and FIGS. 33a, b). In addition, we were able to detect point mutations in the BRAF and NRAS genes using RGENs that recognize the NAG PAM sequence (FIGS. 33c,

d). We believe that we can use RGEN-RFLP to genotype almost any, if not all, mutations or polymorphisms in the human and other genomes.

The above data proposes RGENs as providing a platform to use simple and robust RFLP analysis for various sequence variations. With high flexibility in reprogramming target sequence, RGENs can be used to detect various genetic variations (single nucleotide variations, small insertion/deletions, structural variations) such as disease-related recurring mutations, genotypes related to drug-response by a patient and also mutations induced by engineered nucleases in cells. Here, we used RGEN genotyping to detect mutations induced by engineered nucleases in cells and animals. In principle, one could also use RGENs that will specifically detect and cleave naturally-occurring variations and mutations.

Based on the above description, it should be understood by those skilled in the art that various alternatives to the embodiments of the invention described herein may be employed in practicing the invention without departing from the technical idea or essential features of the invention as defined in the following claims. In this regard, the above-described examples are for illustrative purposes only, and the invention is not intended to be limited by these examples. The scope of the present invention should be understood to include all of the modifications or modified form derived from the meaning and scope of the following claims or its equivalent concepts.

REFERENCES

1. M. Jinek et al., *Science* 337, 816 (Aug. 17, 2012).
2. H. Kim, E. Um, S. R. Cho, C. Jung, J. S. Kim, *Nat Methods* 8, 941 (November 2011).
3. H. J. Kim, H. J. Lee, H. Kim, S. W. Cho, J. S. Kim, *Genome Res* 19, 1279 (July 2009).
4. E. E. Perez et al., *Nat Biotechnol* 26, 808 (July 2008).
5. J. C. Miller et al., *Nat Biotechnol* 29, 143 (February 2011).
6. C. Mussolino et al., *Nucleic Acids Res* 39, 9283 (November 2011).
7. J. Cohen, *Science* 332, 784 (May 13, 2011).
8. V. Pattanayak, C. L. Ramirez, J. K. Joung, D. R. Liu, *Nat Methods* 8, 765 (September 2011).
9. R. Gabriel et al., *Nat Biotechnol* 29, 816 (September 2011).
10. E. Kim et al., *Genome Res*, (Apr. 20, 2012).
11. H. J. Lee, J. Kweon, E. Kim, S. Kim, J. S. Kim, *Genome Res* 22, 539 (March 2012).
12. H. J. Lee, E. Kim, J. S. Kim, *Genome Res* 20, 81 (January 2010).
13. Fu Y, Foden J A, Khayter C, Maeder M L, Reyon D, Joung J K, Sander J D. High-frequency off-target mutagenesis induced by CRISPR-Cas nucleases in human cells. *Nat Biotech advance online publication* (2013)

SEQUENCE LISTING

```

Sequence total quantity: 332
SEQ ID NO: 1          moltype = DNA length = 4107
FEATURE              Location/Qualifiers
misc_feature          1..4107
                       note = Description of Artificial Sequence: Synthetic
                       polynucleotide
misc_feature          1..4107
                       note = Cas9-coding sequence
source                1..4107

```

-continued

```

mol_type = other DNA
organism = synthetic construct

SEQUENCE: 1
atgacaaga agtacagcat cggcctggac atcgggtacca acagcgtggg ctgggcccgtg 60
atcaccgacg agtacaaggt gccagcaag aagttcaagg tgetgggcaa caccgaccgc 120
cacagcatca agaagaacct gatcggcgcc ctgctgttcg acagcggcga gaccgcccag 180
gccaccgcc tgaagcgacc cgcccgcgc cgctacccc gcccaagaa ccgatctgc 240
tacctgcagg agatcttcag caacgagatg gccaaagtg acgacagctt cttccaccgc 300
ctggaggaga gcttcctggg ggaggaggac aagaagcac agcgcaccc catctctcgc 360
aacatcgtgg acgaggtggc ctaccacgag aagtacccca ccattacca cctgcgcaag 420
aagctggggg acagcaccga caaggccgac ctgcccctga tctacctggc cctggcccac 480
atgatcaagt tcccgcgcca cttcctgatc gagggcgacc tgaaccccga caacagcgac 540
gtggacaagc tgttcattca gctgggtgcag acctacaacc agctgttcga ggagaacccc 600
atcaacgcca gccgctggga cgccaagggc atcctgagcg cccgcctgag caagagcccgc 660
cgccctggaga acctgatcgc ccagctgccc ggcgagaaga agaaccggct gttcggcaac 720
ctgatcgccc tgagcctggg cctgaccccc aacttcaaga gcaacttcca cctggccgag 780
gacccaagc tgcagctgag caaggacacc tacgacgacg acctgggaaa cctgctggcc 840
cagatcggcg acctgacgac cgacctgttc ctggccgcca agaacctgag cgacgccatc 900
ctgctgagcg acctcctgcg gctgaacacc gagatcacca agggcccctt gagcggccagc 960
atgatcaagc gctacgacga gcaccaccag gacctgaccc tgetgaaagg cctggctgcg 1020
cagcagctgc ccgagaagta caaggagatc ttcttcgacc agagcaagaa cggctacgcc 1080
ggctacatcg accggcgccg gaccaggag gagttctaca agtctatcaa gccatctcg 1140
gagaagatgg accgcaaccga ggagctgctg gtgaaagtg acccgagga cctgctgcgc 1200
aagcagcgca ccttcgacaa cggcagcatc ccccaccaga tccacctggg cgagctgcac 1260
gccatcctgc gcccaccgga ggactctac ccttctctga aggacaaccg cgagaagatc 1320
gagaagatcc tgaccttcgc catcccctac tacgtgggccc cctggcccgc cggcaacagc 1380
cgcttcgccc ggatgacccc caagagcgag gagaccatca cccctggaa cttcggaggag 1440
gtgggtggaca agggcgccag gcccagagc ttcctgagc gcatgaccaa cttcgacaag 1500
aacctgcccc acgagaaggt gctgcccacg cacagcctgc tgtacgagta cttcaccgtg 1560
tacaacgagc tgaccaaggt gaagtacgtg accgagggca tgcgcaagcc cgcttctctg 1620
agcggcgagc agaagaaggc catcgtggac ctgctgttca agaccaaccg caaggtgacc 1680
gtgaaagcagc tgaaggaagga ctacttcaag aagatcgagt gcttcgacag cgtggagatc 1740
agcggcgtgg aggaccgctt caacgcccagc ctgggcaact accacgacct gctgaagatc 1800
atcaaggaca ccgacttctt ggacaacgag gagaacgagg acatcctgga ggacatctg 1860
ctgaccctga cctgttcga ggaccgagc atgatcgagg agcgcctgaa gacctacgcc 1920
cacctgttcg acgacaaggt gatgaagcag ctgaagcgcc gccgctacac cggctggggc 1980
cgccctgagc gcaagcttat caacgcccgc cgcaacaagc agagcggcaa gacctctg 2040
gacttctga agagcgacgg cttcgccaac cgcaacttca tgcagctgat ccacgacgac 2100
agcctgacct tcaaggagga catccagaag gccacgggtga gcggccaggg cgacagcctg 2160
cacgagcaca tcgccaacct ggccggcagc cccgcccata agaagggcat cctgcagacc 2220
gtgaaaggtg tggacgagct ggtgaaagtg atgggcccgc acaagcccga gaacatcgtg 2280
atcgagatgg cccgagagaa ccagaccacc cagaagggcc agaagaacag ccgagcagcg 2340
atgaaagcga tcgaggaggc catcaaggag ctgggcaacc agatcctgaa ggagcaaccc 2400
gtgggaaaca cccagctgca gaacgagaag ctgtacctgt actacctgca gaacggcccgc 2460
gacatgtacg tggaccagga gctggacatc aaccgcccga gcgactacga cgtggaccac 2520
atcgtgcccc agagcttctt gaaaggacgac agcatcgaca acaaggtgct gaaccgagc 2580
gacaagaacc gccgcaagag cgacaacgtg cccagcagag aggtgggtgaa gaagatgaa 2640
aactactggc gccagctgct gaacgcccag ctgatcacc agcgcgaagt cgacaacctg 2700
accaagggcg agcggcgccg cctgagcagc ctggacaagg ccgcttctat caagcggcag 2760
ctgggtggaga cccgccagat caaccaagcagc gtggcccaga tccctggacag ccgcatgaa 2820
accaagtacg acgagaacga caagctgatc cgcgaggtga aggtgatcac cctgaagagc 2880
aagctggtga gccacttccg caaggacttc cagttctaca aggtgcgca gatcaacaac 2940
taccaccagc gccacgacgc ctacctgaac gccgtggtg gcaccgcccct gatcaagaag 3000
taccccaagc tggagagcga gttcgtgtac ggcgactaca aggtgtacga cgtgcgcaag 3060
atgatcgcca agagcagga ggagatcggc aaggcccacc ccaagtaact cttctacagc 3120
aacatcatga acttctcaa gaccgagatc accctggcca acggcgagat ccgcaagcgc 3180
cccctgatcg agaccaacgg cgagaccggc gagatcgtgt gggacaaggg ccgagcattc 3240
gccaccgtgc gcaaggtgct gagcatgccc caggtgaaaca tctgtaagaa gaccgaggtg 3300
cagaccggcg gcttcagcaa ggagagcatc ctgcccgaag gcaacagcga caagctgatc 3360
gcccgcaaga aggactggga cccaagaag tacggcgctc tcgacagccc caccgtggcc 3420
tacagcgtgc tgggtggtgg caaggtggag aagggcaaga gcaagaagct gaagagcgtg 3480
aaggagctgc tgggcatcac catcatggag cgcagcagct tcgagaagaa ccccatcgac 3540
ttcctggagg ccaagggtca caaggaggtg aagaaggacc tgatcatcaa gctgcccagg 3600
tacagcctgt tcgagctgga aaacggcccg aagcgcctgc tggccagcgc cggcagctg 3660
cagaagggca acgagctggc cctgcccagc aagtacgtga acttctctga cctggcccagc 3720
cactacgaga agctgaaagg cagccccgag gacaacgagc agaagcagct gttcgtggag 3780
cagcacaagc actacctgga cgagatcatc gagcagatca gcgagttcag caagcggctg 3840
atcctggccc ggaccaacct ggacaaggtg ctgagcgcct acaacaagca ccgcaacaag 3900
cccatccgag agcaggccga gaacatcatc cacctgttca cctgaccaca cctgggcccgc 3960
cccggccgct tcaagtactt cgacaccacc atcgaccgca agcgcctacac cagcaccagg 4020
gaggtgctgg acgcccacct gatccaccg agcatcaccg gtctgtacga gaccgccatc 4080
gacctgagcc agctggggcg cgactaa 4107

```

```

SEQ ID NO: 2          moltype = AA length = 21
FEATURE              Location/Qualifiers
REGION               1..21
                    note = Description of Artificial Sequence: Synthetic peptide
REGION               1..21
                    note = Peptide tag

```

-continued

source 1..21
 mol_type = protein
 organism = synthetic construct

SEQUENCE: 2
 GSGGPPKKKR KVYPYDVPDY A 21

SEQ ID NO: 3 moltype = DNA length = 34
 FEATURE Location/Qualifiers
 misc_feature 1..34
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..34
 note = F primer for CCR5
 source 1..34
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 3
 aattcatgac atcaattatt atacatcgga ggag 34

SEQ ID NO: 4 moltype = DNA length = 34
 FEATURE Location/Qualifiers
 misc_feature 1..34
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..34
 note = R primer for CCR5
 source 1..34
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 4
 gatcctcctc cgatgtataa taattgatgt catg 34

SEQ ID NO: 5 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..20
 note = F1 primer for CCR5
 source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 5
 ctccatggtg ctatagagca 20

SEQ ID NO: 6 moltype = DNA length = 21
 FEATURE Location/Qualifiers
 misc_feature 1..21
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..21
 note = F2 primer for CCR5
 source 1..21
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 6
 gagccaagct ctccatctag t 21

SEQ ID NO: 7 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..20
 note = R primer for CCR5
 source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 7
 gccctgtcaa gagttgacac 20

SEQ ID NO: 8 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..20
 note = F1 primer for C4BPB
 source 1..20

-continued

```

mol_type = other DNA
organism = synthetic construct
SEQUENCE: 8
tatttgctg gttgaaaggg                20

SEQ ID NO: 9      moltype = DNA length = 24
FEATURE          Location/Qualifiers
misc_feature     1..24
                 note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
misc_feature     1..24
                 note = R1 primer for C4BPB
source          1..24
                 mol_type = other DNA
                 organism = synthetic construct

SEQUENCE: 9
aaagtcatga aataaacaca ccca          24

SEQ ID NO: 10     moltype = DNA length = 24
FEATURE          Location/Qualifiers
misc_feature     1..24
                 note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
misc_feature     1..24
                 note = F2 primer for C4BPB
source          1..24
                 mol_type = other DNA
                 organism = synthetic construct

SEQUENCE: 10
ctgcattgat atggtagtag catg          24

SEQ ID NO: 11     moltype = DNA length = 21
FEATURE          Location/Qualifiers
misc_feature     1..21
                 note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
misc_feature     1..21
                 note = R2 primer for C4BPB
source          1..21
                 mol_type = other DNA
                 organism = synthetic construct

SEQUENCE: 11
gctgttcatt gcaatggaat g             21

SEQ ID NO: 12     moltype = DNA length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
                 note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
misc_feature     1..20
                 note = F1 primer for ADCY5
source          1..20
                 mol_type = other DNA
                 organism = synthetic construct

SEQUENCE: 12
gtccccacct tagtgctctg              20

SEQ ID NO: 13     moltype = DNA length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
                 note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
misc_feature     1..20
                 note = R1 primer for ADCY5
source          1..20
                 mol_type = other DNA
                 organism = synthetic construct

SEQUENCE: 13
ggtggcagga acctgtatgt              20

SEQ ID NO: 14     moltype = DNA length = 21
FEATURE          Location/Qualifiers
misc_feature     1..21
                 note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
misc_feature     1..21
                 note = F2 primer for ADCY5
source          1..21
                 mol_type = other DNA

```

-continued

organism = synthetic construct
 SEQUENCE: 14
 gtcattggcc agagatgtgg a 21

SEQ ID NO: 15 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..20
 note = R2 primer for ADCY5
 source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 15
 gtcccatgac aggcgtgtat 20

SEQ ID NO: 16 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..20
 note = F primer for KCNJ6
 source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 16
 gctggccea gtttcagtta 20

SEQ ID NO: 17 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..20
 note = R1 primer for KCNJ6
 source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 17
 tggagccatt ggtttgcac 20

SEQ ID NO: 18 moltype = DNA length = 22
 FEATURE Location/Qualifiers
 misc_feature 1..22
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..22
 note = R2 primer for KCNJ6
 source 1..22
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 18
 ccagaactaa gccgtttctg ac 22

SEQ ID NO: 19 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..20
 note = F1 primer for CNTNAP2
 source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 19
 atcaccgaca accagtttcc 20

SEQ ID NO: 20 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..20
 note = F2 primer for CNTNAP2
 source 1..20
 mol_type = other DNA
 organism = synthetic construct

-continued

SEQUENCE: 20
 tgcagtgcag actctttoca 20

SEQ ID NO: 21 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..20
 note = R primer for CNTNAP2
 source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 21
 aaggacacag ggcaactgaa 20

SEQ ID NO: 22 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..20
 note = F1 primer for N/A Chr. 5
 source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 22
 tgtggaacga gtggtgacag 20

SEQ ID NO: 23 moltype = DNA length = 22
 FEATURE Location/Qualifiers
 misc_feature 1..22
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..22
 note = R1 primer for N/A Chr. 5
 source 1..22
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 23
 gctggattag gaggcaggat tc 22

SEQ ID NO: 24 moltype = DNA length = 22
 FEATURE Location/Qualifiers
 misc_feature 1..22
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..22
 note = F2 primer for N/A Chr. 5
 source 1..22
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 24
 gtgctgagaa cgcttcatag ag 22

SEQ ID NO: 25 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..23
 note = R2 primer for N/A Chr. 5
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 25
 ggaccaaacc acattcttct cac 23

SEQ ID NO: 26 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..20
 note = F primer for deletion
 source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 26

-continued

```

ccacatctcg ttctcggttt                20

SEQ ID NO: 27      moltype = DNA length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..20
                  note = R primer for deletion
source           1..20
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 27
tcacaagccc acagatattt                20

SEQ ID NO: 28      moltype = RNA length = 105
FEATURE           Location/Qualifiers
misc_feature      1..105
                  note = Description of Artificial Sequence: Synthetic
                  polynucleotide
misc_feature      1..105
                  note = sgRNA for CCR5
source           1..105
                  mol_type = other RNA
                  organism = synthetic construct

SEQUENCE: 28
ggtgacatca attattatac atgttttaga gctagaaata gcaagttaaa ataaggctag 60
tccgttatca acttgaaaaa gtggcaccga gtcggtgctt ttttt                105

SEQ ID NO: 29      moltype = RNA length = 44
FEATURE           Location/Qualifiers
misc_feature      1..44
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..44
                  note = crRNA for CCR5
source           1..44
                  mol_type = other RNA
                  organism = synthetic construct

SEQUENCE: 29
ggtgacatca attattatac atgttttaga gctatgctgt tttg                44

SEQ ID NO: 30      moltype = RNA length = 86
FEATURE           Location/Qualifiers
misc_feature      1..86
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..86
                  note = tracrRNA for CCR5
source           1..86
                  mol_type = other RNA
                  organism = synthetic construct

SEQUENCE: 30
ggaaccattc aaaacagcat agcaagttaa aataaggcta gtccggtatc aacttgaaaa 60
agtggcaccg agtcggtgct tttttt                86

SEQ ID NO: 31      moltype = DNA length = 86
FEATURE           Location/Qualifiers
misc_feature      1..86
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..86
                  note = Foxn1 #1 sgRNA
source           1..86
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 31
gaaattaata cgactcacta taggcagtct gacgtcacac ttccgtttta gagctagaaa 60
tagcaagtta aaataaggct agtccc                86

SEQ ID NO: 32      moltype = DNA length = 86
FEATURE           Location/Qualifiers
misc_feature      1..86
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..86
                  note = Foxn1 #2 sgRNA
source           1..86
                  mol_type = other DNA

```

-continued

```

                organism = synthetic construct
SEQUENCE: 32
gaaattaata cgactcacta taggacttcc aggctccacc cgacgtttta gagctagaaa 60
tagcaagtta aaataaggct agtccg 86

SEQ ID NO: 33      moltype = DNA length = 86
FEATURE           Location/Qualifiers
misc_feature      1..86
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..86
                  note = Foxn1 #3 sgRNA
source           1..86
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 33
gaaattaata cgactcacta taggccaggc tccaccgcac tggagtttta gagctagaaa 60
tagcaagtta aaataaggct agtccg 86

SEQ ID NO: 34      moltype = DNA length = 86
FEATURE           Location/Qualifiers
misc_feature      1..86
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..86
                  note = Foxn1 #4 sgRNA
source           1..86
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 34
gaaattaata cgactcacta taggactgga gggcgaaccc caaggtttta gagctagaaa 60
tagcaagtta aaataaggct agtccg 86

SEQ ID NO: 35      moltype = DNA length = 86
FEATURE           Location/Qualifiers
misc_feature      1..86
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..86
                  note = Foxn1 #5 sgRNA
source           1..86
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 35
gaaattaata cgactcacta taggacccca aggggacctc atgcgtttta gagctagaaa 60
tagcaagtta aaataaggct agtccg 86

SEQ ID NO: 36      moltype = DNA length = 86
FEATURE           Location/Qualifiers
misc_feature      1..86
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..86
                  note = Prkdc #1 sgRNA
source           1..86
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 36
gaaattaata cgactcacta taggttagtt tttccagag acttgtttta gagctagaaa 60
tagcaagtta aaataaggct agtccg 86

SEQ ID NO: 37      moltype = DNA length = 86
FEATURE           Location/Qualifiers
misc_feature      1..86
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..86
                  note = Prkdc #2 sgRNA
source           1..86
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 37
gaaattaata cgactcacta taggttggtt tgcttggtt tategtttta gagctagaaa 60
tagcaagtta aaataaggct agtccg 86

SEQ ID NO: 38      moltype = DNA length = 86
FEATURE           Location/Qualifiers
misc_feature      1..86
                  note = Description of Artificial Sequence: Synthetic

```

-continued

```

misc_feature      oligonucleotide
                  1..86
                  note = Prkdc #3 sgRNA
source            1..86
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 38
gaaattaata cgactcacta taggcacaag caaaccaaag tctcgtttta gagctagaaa 60
tagcaagtta aaataaggct agtccc                                     86

SEQ ID NO: 39      moltype = DNA length = 86
FEATURE           Location/Qualifiers
misc_feature      1..86
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..86
                  note = Prkdc #4 sgRNA
source            1..86
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 39
gaaattaata cgactcacta taggcctcaa tgctaagcga cttcgtttta gagctagaaa 60
tagcaagtta aaataaggct agtccc                                     86

SEQ ID NO: 40      moltype = DNA length = 29
FEATURE           Location/Qualifiers
misc_feature      1..29
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..29
                  note = F1 primer for Foxn1
source            1..29
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 40
gtctgtctat catctcttcc cttctctcc                               29

SEQ ID NO: 41      moltype = DNA length = 25
FEATURE           Location/Qualifiers
misc_feature      1..25
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..25
                  note = F2 primer for Foxn1
source            1..25
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 41
tccctaatacc gatggctagc tccag                                  25

SEQ ID NO: 42      moltype = DNA length = 23
FEATURE           Location/Qualifiers
misc_feature      1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..23
                  note = R1 primer for Foxn1
source            1..23
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 42
acgagcagct gaagttagca tgc                                    23

SEQ ID NO: 43      moltype = DNA length = 32
FEATURE           Location/Qualifiers
misc_feature      1..32
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..32
                  note = R2 primer for Foxn1
source            1..32
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 43
ctactcaatg ctcttagagc taccaggctt gc                          32

SEQ ID NO: 44      moltype = DNA length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20

```

-continued

note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..20
 note = F primer for Prkdc
 source 1..20
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 44
 gactgttg gggaggccg 20
 SEQ ID NO: 45 moltype = DNA length = 24
 FEATURE Location/Qualifiers
 misc_feature 1..24
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..24
 note = F2 primer for Prkdc
 source 1..24
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 45
 gggaggccg aaagtcttat ttg 24
 SEQ ID NO: 46 moltype = DNA length = 28
 FEATURE Location/Qualifiers
 misc_feature 1..28
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..28
 note = R1 primer for Prkdc
 source 1..28
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 46
 cctgaagact gaagttggca gaagtgag 28
 SEQ ID NO: 47 moltype = DNA length = 27
 FEATURE Location/Qualifiers
 misc_feature 1..27
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..27
 note = R2 primer for Prkdc
 source 1..27
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 47
 ctttaggct tcttctctac aatcacg 27
 SEQ ID NO: 48 moltype = DNA length = 38
 FEATURE Location/Qualifiers
 misc_feature 1..38
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..38
 note = F primer for Foxn1
 source 1..38
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 48
 ctcggtgtg agccctgacc tcggtgtga gccctgac 38
 SEQ ID NO: 49 moltype = DNA length = 21
 FEATURE Location/Qualifiers
 misc_feature 1..21
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..21
 note = R primer for Foxn1
 source 1..21
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 49
 agactggcct ggaactcaca g 21
 SEQ ID NO: 50 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic

-continued

```

misc_feature      oligonucleotide
                  1..23
                  note = F primer for Foxn1
source           1..23
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 50
cactaaagcc tgtcaggaag ccg                               23

SEQ ID NO: 51      moltype = DNA length = 21
FEATURE           Location/Qualifiers
misc_feature      1..21
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..21
                  note = R primer for Foxn1
source           1..21
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 51
ctgtggagag cacacagcag c                                21

SEQ ID NO: 52      moltype = DNA length = 19
FEATURE           Location/Qualifiers
misc_feature      1..19
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..19
                  note = F primer for Foxn1
source           1..19
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 52
gctgcgacct gagaccatg                                   19

SEQ ID NO: 53      moltype = DNA length = 26
FEATURE           Location/Qualifiers
misc_feature      1..26
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..26
                  note = R primer for Foxn1
source           1..26
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 53
cttcaatggc ttctgetta ggctac                           26

SEQ ID NO: 54      moltype = DNA length = 23
FEATURE           Location/Qualifiers
misc_feature      1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..23
                  note = F primer for Foxn1
source           1..23
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 54
ggttcagatg aggccatcct ttc                             23

SEQ ID NO: 55      moltype = DNA length = 24
FEATURE           Location/Qualifiers
misc_feature      1..24
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..24
                  note = R primer for Foxn1
source           1..24
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 55
cctgatctgc aggettaacc cttg                             24

SEQ ID NO: 56      moltype = DNA length = 22
FEATURE           Location/Qualifiers
misc_feature      1..22
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide

```

-continued

```

misc_feature      1..22
                  note = F primer for Prkdc
source            1..22
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 56
ctcacctgca catcacatgt gg                               22

SEQ ID NO: 57      moltype = DNA length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..20
                  note = R primer for Prkdc
source            1..20
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 57
ggcatccacc ctatggggtc                                 20

SEQ ID NO: 58      moltype = DNA length = 25
FEATURE           Location/Qualifiers
misc_feature      1..25
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..25
                  note = F primer for Prkdc
source            1..25
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 58
gccttgacct agagcttaaa gagcc                           25

SEQ ID NO: 59      moltype = DNA length = 25
FEATURE           Location/Qualifiers
misc_feature      1..25
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..25
                  note = R primer for Prkdc
source            1..25
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 59
ggtcttgta gcaggaagga cactg                             25

SEQ ID NO: 60      moltype = DNA length = 27
FEATURE           Location/Qualifiers
misc_feature      1..27
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..27
                  note = F primer for Prkdc
source            1..27
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 60
aaaactctgc ttgatgggat atgtggg                          27

SEQ ID NO: 61      moltype = DNA length = 26
FEATURE           Location/Qualifiers
misc_feature      1..26
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..26
                  note = R primer for Prkdc
source            1..26
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 61
ctctcactgg ttatctgtgc tccttc                           26

SEQ ID NO: 62      moltype = DNA length = 23
FEATURE           Location/Qualifiers
misc_feature      1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..23

```

-continued

source note = F primer for Prkdc
 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 62
 ggatcaatag gtggtggggg atg 23

SEQ ID NO: 63 moltype = DNA length = 27
 FEATURE Location/Qualifiers
 misc_feature 1..27
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..27
 note = R primer for Prkdc
 source 1..27
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 63
 gtgaatgaca caatgtgaca gcttcag 27

SEQ ID NO: 64 moltype = DNA length = 28
 FEATURE Location/Qualifiers
 misc_feature 1..28
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..28
 note = F primer for Prkdc
 source 1..28
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 64
 cacaagacag acctctcaac attcagtc 28

SEQ ID NO: 65 moltype = DNA length = 32
 FEATURE Location/Qualifiers
 misc_feature 1..32
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..32
 note = R primer for Prkdc
 source 1..32
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 65
 gtgcatgcat ataatccatt ctgattgctc tc 32

SEQ ID NO: 66 moltype = DNA length = 17
 FEATURE Location/Qualifiers
 misc_feature 1..17
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..17
 note = F1 primer for Prkdc
 source 1..17
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 66
 gggaggcaga ggcaggt 17

SEQ ID NO: 67 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..23
 note = F2 primer for Prkdc
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 67
 ggatctctgt gagtttgagg cca 23

SEQ ID NO: 68 moltype = DNA length = 24
 FEATURE Location/Qualifiers
 misc_feature 1..24
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..24
 note = R1 primer for Prkdc

-continued

source 1..24
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 68
 gctccagaac tcactcttag gctc 24

SEQ ID NO: 69 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..20
 note = Primer for Foxn1
 source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 69
 ctactccctc gcgagtctga 20

SEQ ID NO: 70 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..20
 note = Primer for Foxn1
 source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 70
 ccaggcctag gttccaggta 20

SEQ ID NO: 71 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..20
 note = Primer for Prkdc
 source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 71
 ccccagcatt gcagatttcc 20

SEQ ID NO: 72 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..23
 note = Primer for Prkdc
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 72
 agggcttctt ctctacaatc acg 23

SEQ ID NO: 73 moltype = DNA length = 86
 FEATURE Location/Qualifiers
 misc_feature 1..86
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..86
 note = BRI1 target 1
 source 1..86
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 73
 gaaattaata cgactcacta taggtttgaa agatggaagc gcggtttta gagctagaaa 60
 tagcaagtta aaataaggct agtccg 86

SEQ ID NO: 74 moltype = DNA length = 86
 FEATURE Location/Qualifiers
 misc_feature 1..86
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..86
 note = BRI1 target 2

-continued

source 1..86
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 74
 gaaattaata cgactcacta taggtgaaac taaactggtc cacagtttta gagctagaaa 60
 tagcaagtta aaataaggct agtccg 86

SEQ ID NO: 75 moltype = DNA length = 64
 FEATURE Location/Qualifiers
 misc_feature 1..64
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

misc_feature 1..64
 note = Universal

source 1..64
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 75
 aaaaaagcac cgactcggcg ccactttttc aagttgataa cggactagcc ttattttaac 60
 ttgc 64

SEQ ID NO: 76 moltype = DNA length = 65
 FEATURE Location/Qualifiers
 misc_feature 1..65
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

misc_feature 1..65
 note = Templates for crRNA

misc_difference 25..44
 note = modified_base - a, c, t, g, unknown or other

source 1..65
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 76
 gaaattaata cgactcacta taggnnnnnn nnnnnnnnnn nnnngtttta gagctatgct 60
 gtttt 65

SEQ ID NO: 77 moltype = DNA length = 67
 FEATURE Location/Qualifiers
 misc_feature 1..67
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

misc_feature 1..67
 note = tracrRNA

source 1..67
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 77
 gaaattaata cgactcacta taggaacct tcaaacagc atagcaagtt aaaataaggc 60
 tagtccg 67

SEQ ID NO: 78 moltype = DNA length = 69
 FEATURE Location/Qualifiers
 misc_feature 1..69
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

misc_feature 1..69
 note = tracrRNA

source 1..69
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 78
 aaaaaaagca ccgactcggg gccacttttt caagttgata acggactagc cttattttaa 60
 cttgctatg 69

SEQ ID NO: 79 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

misc_feature 1..20
 note = Primer

source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 79
 ctccatgggtg ctatagagca 20

SEQ ID NO: 80 moltype = DNA length = 21

-continued

| | | |
|----------------------------|--|----|
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..21 | |
| | note = Description of Artificial Sequence: Synthetic | |
| | oligonucleotide | |
| misc_feature | 1..21 | |
| | note = Primer | |
| source | 1..21 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 80 | | |
| gagccaagct ctccatctag t | | 21 |
| SEQ ID NO: 81 | moltype = DNA length = 20 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..20 | |
| | note = Description of Artificial Sequence: Synthetic | |
| | oligonucleotide | |
| misc_feature | 1..20 | |
| | note = Primer | |
| source | 1..20 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 81 | | |
| gccctgtcaa gagttgacac | | 20 |
| SEQ ID NO: 82 | moltype = DNA length = 22 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..22 | |
| | note = Description of Artificial Sequence: Synthetic | |
| | oligonucleotide | |
| misc_feature | 1..22 | |
| | note = Primer | |
| source | 1..22 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 82 | | |
| gcacagggtg gaacaagatg ga | | 22 |
| SEQ ID NO: 83 | moltype = DNA length = 24 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..24 | |
| | note = Description of Artificial Sequence: Synthetic | |
| | oligonucleotide | |
| misc_feature | 1..24 | |
| | note = Primer | |
| source | 1..24 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 83 | | |
| gccaggtacc tatcgattgt cagg | | 24 |
| SEQ ID NO: 84 | moltype = DNA length = 21 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..21 | |
| | note = Description of Artificial Sequence: Synthetic | |
| | oligonucleotide | |
| misc_feature | 1..21 | |
| | note = Primer | |
| source | 1..21 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 84 | | |
| gagccaagct ctccatctag t | | 21 |
| SEQ ID NO: 85 | moltype = DNA length = 20 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..20 | |
| | note = Description of Artificial Sequence: Synthetic | |
| | oligonucleotide | |
| misc_feature | 1..20 | |
| | note = Primer | |
| source | 1..20 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 85 | | |
| actctgactg ggtcaccagc | | 20 |
| SEQ ID NO: 86 | moltype = DNA length = 20 | |
| FEATURE | Location/Qualifiers | |

-continued

```

misc_feature      1..20
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..20
                  note = Primer
source            1..20
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 86
tatttgctg gttgaaaggg                               20

SEQ ID NO: 87    moltype = DNA length = 24
FEATURE         Location/Qualifiers
misc_feature    1..24
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature    1..24
                  note = Primer
source          1..24
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 87
aaagtcatga aataaacaca ccca                          24

SEQ ID NO: 88    moltype = DNA length = 24
FEATURE         Location/Qualifiers
misc_feature    1..24
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature    1..24
                  note = Primer
source          1..24
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 88
ctgcattgat atggtagtag catg                          24

SEQ ID NO: 89    moltype = DNA length = 21
FEATURE         Location/Qualifiers
misc_feature    1..21
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature    1..21
                  note = Primer
source          1..21
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 89
gctgttcatt gcaatggaat g                             21

SEQ ID NO: 90    moltype = DNA length = 22
FEATURE         Location/Qualifiers
misc_feature    1..22
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature    1..22
                  note = Primer
source          1..22
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 90
atggagttgg acatggccat gg                            22

SEQ ID NO: 91    moltype = DNA length = 28
FEATURE         Location/Qualifiers
misc_feature    1..28
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature    1..28
                  note = Primer
source          1..28
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 91
actcactatc cacagttcag catttacc                      28

SEQ ID NO: 92    moltype = DNA length = 23
FEATURE         Location/Qualifiers
misc_feature    1..23

```

-continued

note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..23
 note = Primer
 source 1..23
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 92
 tggagatagc tgtcagcaac ttt 23

SEQ ID NO: 93 moltype = DNA length = 29
 FEATURE Location/Qualifiers
 misc_feature 1..29
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..29
 note = Primer
 source 1..29
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 93
 caacaaagca aaggtaaagt tgtaatatag 29

SEQ ID NO: 94 moltype = DNA length = 25
 FEATURE Location/Qualifiers
 misc_feature 1..25
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..25
 note = Primer
 source 1..25
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 94
 ggtttcagga gatgtgttac aaggc 25

SEQ ID NO: 95 moltype = DNA length = 27
 FEATURE Location/Qualifiers
 misc_feature 1..27
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..27
 note = Primer
 source 1..27
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 95
 gattgtgcaa ttcctatgca atcggtc 27

SEQ ID NO: 96 moltype = DNA length = 25
 FEATURE Location/Qualifiers
 misc_feature 1..25
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..25
 note = Primer
 source 1..25
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 96
 cactgggtac ttaatctgta gcctc 25

SEQ ID NO: 97 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..23
 note = Primer
 source 1..23
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 97
 ggtccaagt cattcccagt agc 23

SEQ ID NO: 98 moltype = DNA length = 30
 FEATURE Location/Qualifiers
 misc_feature 1..30
 note = Description of Artificial Sequence: Synthetic

-continued

```

misc_feature      oligonucleotide
                  1..30
                  note = Primer
source           1..30
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 98
catcactgca gttgtaggtt ataactatcc                               30

SEQ ID NO: 99      moltype = DNA length = 26
FEATURE           Location/Qualifiers
misc_feature      1..26
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..26
                  note = Primer
source           1..26
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 99
ttgaaaacca cagatctggt tgaacc                                   26

SEQ ID NO: 100    moltype = DNA length = 22
FEATURE           Location/Qualifiers
misc_feature      1..22
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..22
                  note = Primer
source           1..22
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 100
ggagtgccaa gagaatatct gg                                     22

SEQ ID NO: 101    moltype = DNA length = 32
FEATURE           Location/Qualifiers
misc_feature      1..32
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..32
                  note = Primer
source           1..32
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 101
ctgaaactgg tttcaaaata ttcgtttaa gg                           32

SEQ ID NO: 102    moltype = DNA length = 22
FEATURE           Location/Qualifiers
misc_feature      1..22
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..22
                  note = Primer
source           1..22
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 102
gctctgtatg ccctgtagta gg                                     22

SEQ ID NO: 103    moltype = DNA length = 22
FEATURE           Location/Qualifiers
misc_feature      1..22
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..22
                  note = Primer
source           1..22
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 103
ttgcatctg accttacctt tg                                     22

SEQ ID NO: 104    moltype = DNA length = 23
FEATURE           Location/Qualifiers
misc_feature      1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide

```

-continued

```

misc_feature      1..23
                  note = Target sequence of RGEN
source            1..23
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 104
aatgaccact acatcctcaa ggg                                     23

SEQ ID NO: 105      moltype = DNA length = 23
FEATURE            Location/Qualifiers
misc_feature       1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature       1..23
                  note = Target sequence of RGEN
source            1..23
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 105
agatgatgtc tcatcatcag agg                                     23

SEQ ID NO: 106      moltype = DNA length = 4170
FEATURE            Location/Qualifiers
misc_feature       1..4170
                  note = Description of Artificial Sequence: Synthetic
                  polynucleotide
misc_feature       1..4170
                  note = Cas9-coding sequence in p3s-Cas9HC (humanized,
                  C-term tagging, human cell experiments)
source            1..4170
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 106
atggacaaga agtacagcat cggcctggac atcgggtacca acagcgtggg ctgggccgtg 60
atcaccgacg agtacaaggt gccacgcaag aagttcaagg tgctgggcaa caccgaccgc 120
cacagcatca agaagaacct gatcggcgcc ctgctgttcg acagcggcga gaccgcccag 180
gccaccgcgc tgaagcgcac cgcgccgcgc cgctacaccc gccccaagaa ccgcatctgc 240
tacctgcagg agatcttcag caacgagatg gccaaaggtg acgacagctt cttccaccgc 300
ctggaggaga gcttcctggt ggaggaggac aagaagcagc agcgcaccc catctctcgc 360
aacatcgtgg acgaggtggc ctaccacgag aagtacccca ccattacca cctgcgcaag 420
aagctggtgg acagcaccga caaggccgac ctgcccctga tctacctggc cctggcccac 480
atgatcaagt tccgcccga cttcctgatc gaggcgacc tgaaccccga caacagcgac 540
gtggacaagc tgttcattca gctggtgcag acctacaacc agctgttoga ggagaacccc 600
atcaacgcca gggcgctgga gcccaaggcc atcctgagcg cccgcctgag caagagccgc 660
cgccctggaga acctgatcgc ccagctgccc ggcgagaaga agaaccgccc gttcggcaac 720
ctgatcgcgc tgaagcctggg cctgcacccc aactcaaga gcaacttoga cctggcccag 780
gacgccaagc tgcagctgag caaggacacc tacgacgacg acctggacaa cctgctggcc 840
cagatcggcg accgatcagc cgacctgttc ctggcccga agaacctgag cgacgccatc 900
ctgctgagcg acatcctgcg cgtgaacacc gagatcacca agccccctc gagcgcagc 960
atgatcaagc gctacgacga gccaccaccg gacctgacc tgetgaagge cctgggtgcg 1020
cagcagctgc ccgagaagta caaggagatc ttcttcgacc agagcaagaa cggctacgcc 1080
ggctacatcg acgcccagga cagccagggg gagttctaca agttcatcaa gccctcctg 1140
gagaagatgg acggcaccga ggagctgctg gtgaagctga accgcgagga cctgctgctg 1200
aagcagcgca ccttcgacaa cggcagcatc ccccaccaga tccacctggg cgagctgca 1260
gccatcctgc tgcgcccaga ggaactctac cccttctcga aggacaaccg cgagaagatc 1320
gagaagatcc tgaacctccg catcccctac tacgtgggcc ccctggcccg cggcaacagc 1380
cgcttcgccc ggtgatcccg caagagcgag gagaccatca cccctgggaa cttcagggag 1440
gtggtggaca agggcgcag cgcccagagc ttcctcagag ccatgaccaa cttcagcaag 1500
aacctgcccc acgagaaggt gctgcccagg cacagcctgc tgtacgagta cttcaccgtg 1560
tacaacgagc tgaccaaggt gaagtacgtg accgaggcca tgcgcaagcc cgccttctct 1620
agcggcgaagc agaagaaggg catcgtggac ctgctgttca agaccaaccg caaggtgacc 1680
gtgaagcagc tgaaggagga ctacttcaag aagatcgagt gcttcgacag cgtggagatc 1740
agcggcgtgg aggaccgctt caacgcccagc ctgggcaact accacagcct gctgaagatc 1800
atcaaggaca aggacttctt ggacaacgag gagaacgagg acatcctgga ggacatcgtg 1860
ctgacctgga ccctgttoga ggaccgagag atgatcgagg agcgcctgaa gacctacgcc 1920
cacctgttcg acgacaaggt gatgaagcag ctgaagcgcg gccctacac cggctggggc 1980
cgcctgagcc gcaagcttat caacggcatc cgcgacaagc agagcggcaa gaccatcctg 2040
gacttcttga agagcgacag cttcgccaac cgcaacttca tgcagctgat ccaagcagac 2100
agcctgacct tcaaggagga catccagaag gccaggtgga gcgcccaggg cgacagcctg 2160
cacgagcaca tgcccaacct ggcggcgagc cccgccatca agaagggcat cctgcagacc 2220
gtgaaggtgg tggacgagct ggtgaaggtg atgggcccgc acaagcccga gaacatcgtg 2280
atcgagatgg ccccgagaa ccagaccacc cagaagggcc agaagaacag ccgagagcgc 2340
atgaagcgca tccaggagga catcaaggag ctgggagccc agatcctgaa ggagcacc 2400
gtggagaaca ccagctcga gaacgagaag ctgtacctgt actacctgca gaacggcgcg 2460
gacatgtacg tggaccagga gctggacatc aaccgcctga gcgactacga cgtggaccac 2520
atcgtgcccc agagcttctt gaaggacgac agcatcgaca acaaggtgct gaccccgagc 2580
gacaagaacc gccgcaagag cgacaacgtg cccagcgagg aggtggtgaa gaagatgaag 2640
aactactggc cccagctgct gaacgccaag ctgatcacc agcgcgaagt cgacaacctg 2700
accaagggcg agcggcgggg cctgagcgag ctggacaagg cgggcttcat caagcggcag 2760

```

-continued

```

ctggtggaga cccgccagat caccaagcac gtggcccaga tccctggacag ccgcatgaac 2820
accaagtacg acgagaacga caagctgata cgcgaggatg aggtgatcac cctgaagagc 2880
aagctggtga gcgactcccg caaggacttc cagttctaca aggtgcgcga gatcaacaac 2940
taccaccaag cccacgagcg ctacctgaac gccgtggtgg gcaccgccct gatcaagaag 3000
taccccaagc tggagagcga gttcgtgtac ggcgactaca aggtgtacga cgtgcccgaag 3060
atgatcgcca agagcgagca ggagatcggc aaggccaccg ccaagtactt cttctacagc 3120
aacatcatga acttcttcaa gaccgagatc accctggcca acggcgagat ccgcaagcgc 3180
cccctgatcg agaccaacgg cgagaccggc gagatcgtgt gggacaaggg ccgcgacttc 3240
gccaccgtgc gcaaggtgct gagcatgcc caggtgaaca tcgtgaagaa gaccgaggtg 3300
cagaccggcg gcttcagcaa ggagagcatc ctgcccaagc gcaacagcga caagctgatc 3360
gcccgcaaga aggttgggga ccccaagaag tacggcggct tcgacagccc caccgtggcc 3420
tacagcgtgc tgggtggtggc caaggtggag aagggcaaga gcaagaagct gaagagcgtg 3480
aaggagctgc tgggcatcac catcatggag cgcagcagct tcgagaagaa ccccatcgac 3540
ttcctggagg ccaagggcta caaggggtg aagaaggacc tgatcatcaa gctgcccgaag 3600
tacagcctgt tcgagctgga gaacggccgc aagcgcctgc tggccagcgc cggcgagctg 3660
cagaagggca acgagctggc ctgcccagc aagtaactga acttctgtga cctggccagc 3720
cactacgaga agctgaaggg cagccccgag gacaacgagc agaagcagct gttcgtggag 3780
cagcacaagc actacctgga cgagatcatc gaggagatca gcgagttcag caagcgcgtg 3840
atcctggcgg agcccaacct ggacaaggtg ctgagcgcct acaacaagca ccgcgacaag 3900
cccatccgcg agcagggcga gaacatcatc cacctgttca cctgaccaa cctggggcgc 3960
cccggcgcct tcaagtaact cgacaccacc atcgaccgca agcgtctacac cagcaccgaag 4020
gaggtgctgg acgccacctg gatccaccag agcatcaccg gtctgtacga gaccgccatc 4080
gacctgagcc agctggggcg gcagcggcgc tccggacctc caaagaaaaa gagaaaagta 4140
taccctacg acgtgcccga ctacgcctaa 4170

```

```

SEQ ID NO: 107          moltype = DNA length = 4194
FEATURE                Location/Qualifiers
misc_feature           1..4194
                        note = Description of Artificial Sequence: Synthetic
                        polynucleotide
misc_feature           1..4194
                        note = Cas9 coding sequence in p3s-Cas9HN (humanized codon,
                        N-term tagging (underlined), human cell experiments)
source                 1..4194
                        mol_type = other DNA
                        organism = synthetic construct

```

```

SEQUENCE: 107
atggtgtacc cctacgagct gcccgactac gccgaattgc ctccaaaaaa gaagagaaaag 60
gtagggatcc gaattcccgg gaaaaaaccc gacaagaagt acagcatcgg cctggacatc 120
ggtaccaaca gcgtgggctg ggcctgtgat accgacgagt acaaggtgat cagcaagaag 180
ttcaaggtgc tgggcaaacac cgaccgccac agcatcaaga agaacctgat cggcgccctg 240
ctgttcgaca gcgcgagagc cgcggaggcc accgcctcga agcgcaccgc ccgcccgcgc 300
tacaccggcc gcaagaacct catctgctac ctgcaggaga tcttcagcaa cgagatggcc 360
aaggtggacg acagcttctt ccaccgctcg gaggagagct tccctggtgga ggaggacaag 420
aagcacgagc gccaccacct cttcggcaac atcgtggacg aggtggccta ccacgagaag 480
taccocacca tctaccacct cgcgaagaag ctggtggaca gcaaccgcaa ggccgacctg 540
cgctgatct acctggccct ggcccacatg atcaagttcc ggggccactt cctgatcgag 600
ggcgacctga accccgacaa cagcgcagct gacaagctgt tcatccagct ggtgcagacc 660
tacaaccagc tgttcgagca gaaccacctc aacgcccagc gcgtggagcg caaggacctc 720
ctgagcggcc gcctgagcaa gagccgcccgc ctggagaacc tgatgcgcca gctgcccggc 780
gagaagaaga accgctctgt cggcaacctg atcgccctga gcctgggccc gaccoccaa 840
ttcaagagca acttcgacct ggcggagacc gccaaagctgc agctgagcaa ggacacctac 900
gacgacgacc tggacaacct gctggcccag atcggcagcc agtacgccc cctgttctctg 960
gccgccaaga accctgagcga cgccatcctg ctgagcgaca tccctgctgt gaacaccgag 1020
atcaccgaag cctccctgag gcaccgcatg atcaagcgtc acgacgagca ccaccaggac 1080
ctgacctcgc tgaaggccct ggtgcccagc cagctgcccg agaagtacaa ggagatcttc 1140
ttcgaccaga gcaagaacgg ctacgcccgc tacatcgacg gcggcgccag ccaggaggag 1200
ttctacaagt tcatcaagcc catcctggag aagatggacg gcaaccgagga gctgctgggtg 1260
aagctgaacc gcgaggacct gctgcccagc cagcgcacct tcgacaacgg cagcatcccc 1320
caccagatcc acctggggcga gctgacgccc atcctgcccg gccaggagga cttctacccc 1380
ttcctgaagg acaaccgcca gaagatcgag aagatcctga ccttccgcat cccctactac 1440
gtgggccccc tggcccggcg caacagcccgc ttcgctctga tgaccgcaa gagcgaggag 1500
accatcacc cctggaactt cgaggagggt gtggacaagg gcgcccagcg ccagagcttc 1560
atcgagcgca tgaccaactt gcacaagaac ctgcccacag agaaggtgct gcccaagcac 1620
agcctgctgt acgagtactt caccgtgtac aacgagctga ccaaggtgaa gtactgtacc 1680
gagggcatgc gcaagcccgc cttcctgagc ggcgagcaga agaagggcat cgtggacctg 1740
ctgttcaaga ccaaccgcaa ggtgaccgtg aagcagctga aggaggacta cttcaagaag 1800
atcgagtctc tcgacagcgt ggagatcagc ggcgtggagg accgcttcaa cgccagcctg 1860
ggcaccatcc acgacctgct gaagatcatc aaggacaagg acttccctgga caaccaggag 1920
aacgaggaca tcctggagga catcgtgctg accctgacct tgttcgagga ccgcgagatg 1980
atcgaggagc gcctgaagac ctacgcccac ctggtcgacg acaaggtgat gaagcagctg 2040
aagcggccgc gctacaccgg ctggggcccgc ctgagccgca agcttatcaa cggcatccgc 2100
gacaagcaga gcggaacagc catcctggac ttcctgaaga gcgacggctt cgccaaccgc 2160
aaactcatgc agtctgatcca gcacgacgc ctgaccttca aggaggacat ccagaaggcc 2220
caggtgagcg gccagggcga cagcctgcac gagcacatcg ccaacctggc cggcagcccc 2280
gccatcaaga agggcatcct gcagaccgtg aagtggtggc acgagctggt gaaggtgatg 2340
ggccgccaca agcccagaa ccatcgtgatc gagatggccc gcgagaacca gaccaccag 2400
aagggccaga agcaacgccc cgagcgcctg aagcgcctcg aggaggcat caaggagctg 2460
ggcagccaga tccctgaagga gaccccgtg gagaacaccg agctgcagaa cgagaagctg 2520

```

-continued

| | | | | | | |
|------------|-------------|------------|-------------|-------------|------------|------|
| tacctgtact | acctgacgaa | cgcccgcgac | atgtactgtg | accaggagct | ggacatcaac | 2580 |
| cgcttgagcg | actacgacgt | ggaccacatc | gtgccccaga | gcttcoctgaa | ggacgacagc | 2640 |
| atcgacaaca | aggtgctgac | ccgcagcgac | aagaaccgcg | gcaagagcga | caacgtgccc | 2700 |
| agcgaggagg | tggtgaagaa | gatgaagaac | tactggcgcc | agctgctgaa | cgccaagctg | 2760 |
| atcaccacgc | gcaagttoga | caacctgacc | aaggccgagc | gcgccggcct | gagcgagctg | 2820 |
| gacaaggccg | gcttcatcaa | gcccagctg | gtggagacc | gccagatcac | caagcacgtg | 2880 |
| gcccagatcc | tggaacgccc | catgaacacc | aagtaacgac | agaacgacaa | gctgatccgc | 2940 |
| gaggtgaagg | tgatccacct | gaagagcaag | ctggtgagcg | acttccgcaa | ggacttccag | 3000 |
| ttctacaagg | tgccgagat | caacaactac | caccacgccc | acgacgccta | cctgaaacgc | 3060 |
| gtggtgggca | ccgcccctgat | caagaagtac | cccaagctgg | agagcaggt | cgtgtacggc | 3120 |
| gactacaagg | tgtacgacgt | gcgcaagatg | atcgccaaga | gcgagcagga | gatcggcaag | 3180 |
| gccaccgcca | agtacttctt | ctacagcaac | atcatgaact | tcttcaagac | cgagatcacc | 3240 |
| ctggccaacg | gagagatccg | caagcgcgcc | ctgatcgaga | ccaacggcga | gaccggcgag | 3300 |
| atcgtgtggg | acaagggccg | cgacttcgcc | accgtgcgca | aggtgctgag | catgccccag | 3360 |
| gtgaacatcg | tgaagaagac | cgaggtgcag | accggcggt | tcagcaagga | gagcatcctg | 3420 |
| cccaagcgca | acagcgacaa | gctgatcgcc | cgcaagaagg | actgggacc | caagaagtac | 3480 |
| ggcggtctcg | acagcccctac | gctggcctac | agcgtgctgg | tggtggccaa | ggtggagaag | 3540 |
| ggcaagagca | agaagctgaa | gagcgtgaa | gagctgctgg | gcatcaccat | catggagcgc | 3600 |
| agcagcttcg | agaagaacct | ctcgacttc | ctggaggcca | agggctacaa | ggaggtgaag | 3660 |
| aagacactga | tcatcaagct | gccccagtac | agcctgttcg | agctggagaa | cgcccgaag | 3720 |
| cgcatgctgg | ccagcgcgcg | cgagctgcag | aagggcaacg | agctggccct | gcccagcaag | 3780 |
| tacgtgaact | tcctgtacct | ggccagccac | tacgagaagc | tgaagggcag | ccccaggac | 3840 |
| aacgagcaga | agcagctggt | cgtggagcag | cacaagcact | acctggacga | gatcatcgag | 3900 |
| cagatcagcg | agttcagcaa | gcgcgtgatc | ctggccgacg | ccaacctgga | caagtgctg | 3960 |
| agcgcttaca | acaagcacc | gcacaagccc | atcccgcgagc | agggcagaaa | catcatccac | 4020 |
| ctgttcaccc | tgaccaacct | ggggcgcgcc | gccccttca | agtacttca | caccaccatc | 4080 |
| gaccgcaagc | gctacaccag | caccaaggag | gtgctggacg | ccacctgat | ccaccagagc | 4140 |
| atcaccggtc | tgtacgagac | ccgcatcgac | ctgagccagc | tggcgcgca | ctaa | 4194 |

SEQ ID NO: 108 moltype = DNA length = 4107
FEATURE
misc_feature 1..4107
 note = Description of Artificial Sequence: Synthetic
 polynucleotide
misc_feature 1..4107
 note = Cas9-coding sequence in Streptococcus pyogenes
source 1..4107
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 108

| | | | | | | |
|-------------|-------------|-------------|------------|-------------|-------------|------|
| atggataaga | aataactcaat | aggcttagat | atcggcacaa | atagcgtcgg | atggcggtg | 60 |
| atcactgatg | aataataaggt | tccgtctaaa | aagttcaagg | ttctgggaaa | tacagaccgc | 120 |
| cacagtatca | aaaaaatct | tatagggct | cttttatttg | acagtgagga | gacagcggaa | 180 |
| gagactcgtc | tcaaacggac | agctcgtaga | aggtatacac | gtcggaaaga | tcgtatttgt | 240 |
| tacttacagg | agatTTTTT | aaatgagatg | gcgaaagtag | atgatagttt | ctttcatcga | 300 |
| cttgaagagt | ctttttggt | ggaagaagac | aagaagcatg | aacgtcatcc | tatttttggg | 360 |
| aatatagtag | atgaagttgc | ttatcatgag | aaatatccaa | ctatctatca | tctgcgaaaa | 420 |
| aaattgtag | attctactga | taaagcggat | ttgcgcttaa | tctatttggc | cttagcgcat | 480 |
| atgattaagt | ttcgtggtca | tttttgatt | gagggagatt | taaatcctga | taatagtgat | 540 |
| gtggacaaac | tatttatcca | gttggtacaa | acctacaatc | aattatttga | agaaaacct | 600 |
| ataacgcaa | gtggagtaga | tgctaaagcg | attctttctg | cacgattgag | taaatcaaga | 660 |
| cgattagaaa | atctcattgc | tcagctcccc | ggtgagaaga | aaaatggctt | atTTGGGAAT | 720 |
| ctcattgctt | tgtcattggg | tttgaccctt | aatTTTAAAT | caaatTTTGA | tttggcagaa | 780 |
| gatgctaagt | tacagctttc | aaaagatact | tacgatgatg | atTTAGATAA | tttattggcg | 840 |
| caaattggag | atcaatatgc | tgatttggtt | ttggcagcta | agaatttatc | agatgctatt | 900 |
| ttactttcag | atatacctaa | agtaataact | gaaataacta | aggtccccct | atcagcttca | 960 |
| atgattaaac | gctacgatga | acatcatcaa | gacttgactc | ttttaaaagc | tttagttcga | 1020 |
| caacaacttc | cgaaaaagta | taaaagaatc | ttttttgatc | aatcaaaaaa | cggatagca | 1080 |
| ggttatattg | atgggggagc | tagccaagaa | gaattttata | aatttatcaa | accaatttta | 1140 |
| gaaaaaatgg | atggtactga | ggaattattg | gtgaaactaa | atcgtgaaga | tttgcgtcgc | 1200 |
| aagcaacgga | cttttgacaa | cggtcttatt | ccccatcaaa | ttcacttggg | tgagctgcat | 1260 |
| gctattttga | gaagacaaga | agacttttat | ccatttttaa | aagacaatcg | tgagaagatt | 1320 |
| gaaaaaatct | tgacttttct | aattccttat | tatgttggtc | catTGGCGCG | tgccaatagt | 1380 |
| cgttttgcat | ggatgactcg | gaagtctgaa | gaaacaatta | ccccatggaa | ttttgagaaa | 1440 |
| gttgcgata | aaggtgcttc | agctcaatca | tttattgaac | gcatgacaaa | ctttgataaa | 1500 |
| aatcttccaa | atgaaaaagt | actaccaaaa | catagtttgc | tttatgagta | ttttaccggt | 1560 |
| tataacgaat | tgacaaaagg | caaatatggt | actgaaggaa | tgcaaaaacc | agcatttctt | 1620 |
| tcaggtgaac | agaagaagc | catgtttgat | tactcttca | aaacaatcg | aaaagtaacc | 1680 |
| gttaagcaat | taaaagaaga | ttatttcaaa | aaaatagaat | gttttgatag | tgttgaatt | 1740 |
| tcaggagtty | agatagatt | taatgcttca | ttaggtacct | accatgatt | gctaaaaatt | 1800 |
| ataaagata | aagatttttt | ggataatgaa | gaaaatgaag | atatcttaga | ggatattggt | 1860 |
| ttacattga | ccttatttga | agatagggag | atgattgagg | aaagacttaa | aacatagct | 1920 |
| cacctcttct | atgataaggt | gatgaaacag | cttaaacctc | gcccgttatac | tggttgggga | 1980 |
| cgtttgcctc | gaaaattgat | taatggtatt | agggataagc | aatctggcaa | aacaattatta | 2040 |
| gatttttga | aatcagatgg | ttttgccaat | cgcaatttta | tgcagctgat | ccatgatgat | 2100 |
| agtttgacat | ttaaagaaga | catTCAAAAA | gcacaagtgt | ctggacaagg | cgatagttta | 2160 |
| catgaaacata | ttgcaaaatt | agctggtagc | cctgctatta | aaaaaggtat | tttacagact | 2220 |
| gtaaaagtty | ttgatgaatt | ggtcaaaagta | atggggcgcc | ataagccaga | aaatatcggt | 2280 |
| attgaaatgg | cacgtgaaaa | tcagacaact | caaaaaggcc | agaaaaattc | gcgagagcgt | 2340 |

-continued

SEQUENCE: 110
atgggcagca gccatcatca tcatcatcat gtgtaccctt acgaogtgcc cgactaogcc 60
gaattgcttc caaaaaagaa gagaaaggta gggatcgaga acctgtactt ccaggggcagc 120
aagaagtaca gcatcgccct ggacatcggg accaacacgcy tgggctgggc cgtgatcacc 180
gacgagtaca aggtgcccag caagaagttc aaggtgctgg gcaaacocga ccgocacagc 240
atcaagaaga acctgatcgg cgcctgctg ttcgacagcy gcgagaccgc cgaggccacc 300
cgcctgaagc gcaccgccc gcgcgctac acccgccca agaaccgcat ctgctactg 360
caggagatct tcagcaacga gatggccaag gtggacgaca gcttctcca ccgctggag 420
gagagcttcc tgggtggagg ggacaagaag cacgagcccc accccatctt cggcaacatc 480
gtggacgagg tggcctacca cgagaagta cccaccatct accacctgcy caagaagctg 540
gtggacagca ccgacaagcc cgacctgcy ctgatctacc tggcctggc ccacatgatc 600
aagttccgcy gccacttct gatcgagggc gacctgaacc ccgacaacag cgacgtggac 660
aagctgttca tccagctggt gcagacctac aaccagctgt tcgaggagaa ccccatcaac 720
gccagcggcy tggacgccc ggccatcctg agcccccgc tgagcaagag ccgcccctg 780
gagaacctga tcgcccagct gcccgcgag aagaagaagc gcctgttcgg caacctgatc 840
gccctgagcc tggcctgac ccccaactc aagagcaact tcgacctggc cgaggagcc 900
aagctgcagc tgagcaagga cacctacgac gacgacctg acaacctgcy ggccagatc 960
ggcgaccagt acgcccagct gttcctggcc gccaaagacc tgagcgagcy catcctgctg 1020
agcgacatcc tgcgcgtgaa cccgagatc accaaggccc cctgagcgc cagcatgatc 1080
aagcgtctac cgagcaacca ccaggacctg acctgctga aggcctggt gcgcccagc 1140
ctgcccagca agtacaagga gatcttcttc gaccagagca agaaccgcta ccgcccgtac 1200
atcgacggcy gcgccagcca ggaggagttc tacaagttca tcaagccat cctggagaag 1260
atggacggca ccgaggagct gctggtgaa gtgaaccgcy aggacctgct gcgcaagcag 1320
cgcacctctc acaaccgcy catccccac cagatccacc tggcgagct gcacgccatc 1380
ctgcccgcctc aggaggaact ctacccttc ctgaaaggaca acccgagaa gatcgagaag 1440
atcctgacct tccgcatccc ctactacgtg ggccccctg cccgcccga cagcccctc 1500
gcctggatga cccgcaagag cgaggagacc atccccctt ggaactcga ggaggtggtg 1560
gacaagggcy ccagcgccca gagcttcat gagcgcgatg ccaactcga caagaacctg 1620
cccaacgaga aggtgctgccc caagcacagc ctgctgtacg agtacttca cgtgtacaac 1680
gagctgacca aggtgaagta cgtgaccgag ggcctgcgca agcccgcctt cctgagcggc 1740
gagcagaaga aggccatcgt ggaacctgct tcaagacca accgcaaggt gacogtgaag 1800
cagctgaagg aggaactct caagaagatc gagtctctc acagcgtgga gatcagcggc 1860
gtggaggacc gcttcaacgc cagcctgggc acctaccagc acctgctgaa gatcatcaag 1920
gacaaggact tcctgggaca cgaggagaac gaggacatcc tggaggacat cgtgctgacc 1980
ctgacctgt tcgaggaccg cgagatgatc gaggagcgc tgaagacct a cgcaccctg 2040
ttcgacgaca agtgatgaa gcagctgaag cgcgcccgt acaccgctg gggcccctg 2100
agccgcaagc ttatcaagcy catccgagc aagcagagcy gcaagaccat cctggactt 2160
ctgaagagcy accgcttcgc caaccgcaac ttcctgcagc tgatccacga cgacagcctg 2220
accttcaagg aggcattcca gaagcccag gtgagcggcc agggcgacag cctgcaagc 2280
cacatcgcca acctggccgc cagcccgcct atcaagaagc gcatcctgca gacogtgaag 2340
gtggtggagc agctggtgaa ggtgatgggc cgcacaagc ccgagaacat cgtgatcgag 2400
atggcccgcg agaaccagc caccagaag ggcagaaga acagccgca gcgcatgaa 2460
cgcctcgagg aggcctcaa ggagctggc agccagatcc tgaaggagca cccogtggag 2520
aacaccagc tgcagaacga gaagctgtac ctgtactacc tgcagaagc ccgagacatg 2580
tacctggacc aggagctgga catcaaccgc ctgagcagct acgacgtgga ccacatcgtg 2640
ccccagact tcctgaagcy cgacagcatc gacaacaagc tgcctgaccgc cagocaaag 2700
aacccggca agagcgaca cgtgcccagc gaggaggtg tgaagaagat gaagaactac 2760
tggcgcagc tgcgaaagc caagctgatc acccagcga agtctgaca cctgaccaag 2820
gccagcgcg cgcgctgag cgagctggac aaggcccgtc tcatcaagcy ccagctggctg 2880
gagaccgccc agatcaacca gcacgtggcc cagatcctg acagccgcat gaacaccaag 2940
tacgacgaga acgacaagct gatccgagc gtgaaggta tcacctgaa gagcaagctg 3000
gtgagcgaact tccgcaagcy ctccaagttc tacaaggtgc gcgagatcaa caactaccac 3060
cagcccagc acgctacct gaacgcccgt gtgggcaacc cctgatcaa gaagtacccc 3120
aagctggaga gcgagttcgt gtacggcagc tacaaggtgt acgacgtgcy caagatgatc 3180
gccaagagcy agcaggagat cggcaagccc accgccaagt acttctcta cagcaacatc 3240
atgaactct tcaagaccga gatcacctc gccaacggcy agatccgca gcgcccctg 3300
atcgagacca accgagagc cggcgagatc gtgtgggaca agggccgca cttcgccacc 3360
gtcgcaagc tgcgagcat cccccaggt aacatcgtga agaagaccga ggtgcagacc 3420
ggcgcttca gcaaggagag catcctgccc aagcgcaaca gcgacaagct gatcgcccgc 3480
aagaaggact gggaccocaa gaagtacgcy ggtctcgaca gcccccagct ggctacagc 3540
gtgctggtgg tggccaagcy aagagaagggc aagagaagag agctgaagag cgtgaagag 3600
ctgctgggca tcaccatcat ggagcgcagc agcttcgaga agaaccctc cgaactcctg 3660
gaggccaagc gctacaagga ggtgaagaag gacctgatca tcaagctgccc caagtacagc 3720
ctgttcagc tggagaacgc ccgcaagcgc atgctggcca gcgcccgcga gctgcagaag 3780
ggcaacgagc tggcctgccc cagcaagta gtgaacttcc tgtacctggc cagccactac 3840
gagaagctga agggcagccc cgaggacaac gagcagaagc agctgttctg ggagcagc 3900
aagcactacc tggacgagat catcgagcag atcagcagct tcagcaagcy cgtgatcctg 3960
gccagccca acctggacaa ggtgctgagc gcctacaaca agcaccgca caagccatc 4020
cgcgagcagc ccgagaacat catccacctg ttcacctgca ccaacctggc cgcgccgc 4080
gccttcaagt acttcgacac caccatcgac cgaagcgtc acaccagcag caaggaggtg 4140
ctggacgcca cctgatcca ccagagcatc accggtctgt acgagaccgc catcgactg 4200
agccagctgg gcggcgacta a 4221

SEQ ID NO: 111 moltype = AA length = 1406
FEATURE Location/Qualifiers
REGION 1..1406
note = Description of Artificial Sequence: Synthetic polypeptide
REGION 1..1406

-continued

```

note = Amino acid sequence of Cas9 (pET-Cas9N3T)
source      1..1406
            mol_type = protein
            organism = synthetic construct

SEQUENCE: 111
MGSSHHHHHH VYPYDVPDYA ELPPKKKRKV GIENLYFQGD KKYSIGLDIG TNSVGWAVIT 60
DEYKVPSPKKF KVLGNTDRHS IKKNLIGALL FDSGETAEAT RLKRTARRRY TRRKNRICYL 120
QEIFSNEMAK VDDSFHHRLE ESFLVEEDKK HERHPIFGNI VDEVAYHEKY PTIYHLRKKL 180
VDSTDKADLR LIYLALAHMI KFRGHFLIEG DLNPDNSDVD KLFIQLVQTY NQLFEENPIN 240
ASGVDAKAIL SARLSKSRRL ENLIAQLPGE KKNGLFGNLI ALSLGLTPNF KSNFDLAEDA 300
KLQLSKDITYD DDLNLLAQI GDQYADLFLA AKNLSDAILL SDILRVNTEI TKAPLSASMI 360
KRYDEHHQDL TLLKALVRQQ LPEKYKEIFF DQSKNGYAGY IDGGASQEEF YKFIKPILEK 420
MDGTEELLVK LNREDLLRQK RTFDNGSIPH QIHLGELHAI LRRQEDFYFP LKDNREKIEK 480
ILTFRIPYVYV GPLARGNSRF AWMTRKSEET ITPWNFEEV DKGASAQSF IERMNFDKNL 540
PNEKVLPKHS LLYEYFTVYN ELTKVKYVTE GMRKPAFLSG EQKKAIVDLL FKTNRKVTVK 600
QLKEDYFKKI ECFDSVEISG VEDRFNASLG TYHLLKIIK DKDFLDNEEN EDILEDIVLT 660
LTLFEDREMI EERLKYAHL FDDKVMKQLK RRRYTGWGR LSRKLINGIRD KQSGKTILDF 720
LKSDGFANRN FMQLIHDDSL TPKEDIQKAQ VSGQGDLSHE HIANLAGSPA IKKGILQTVK 780
VVDELVKVMG RHKPENIVIE MARENQTTQK GQKNSRERMK RIEEGIKELG SQLKEHPVE 840
NTQLQNEKLY LYYLQNGRDM YVDQELDINR LSDYVDVHIV PQSFLKDDSI DNKVLTRSDK 900
NRGKSDNVPS EEVVKMKKNY WRQLLNAKLI TQRKFDNLTK AERGGSELD KAGFIKRQLV 960
ETRQITKHVA QILDSRMNTK YDENDKLIRE VKVITLKS KL VSDFRKDFQF YKVREINNYH 1020
HAHDAYLNAV VGTALIKKYP KLESEFVYGD YKVYDVRKMI AKSEQEI GKA TAKYFFYSNI 1080
MNFFKTEITL ANGEIRKRPL IETNGETGEI VWDKGRDPAT VRKVLSPQV NIVKKTVEVQT 1140
GGFSKESLIP KRNSDKLIAR KKDWDPKKYG GFDSPTVAYS VLVVAKVEKG KSKKLKSVKE 1200
LLGITIMERS SFEKNPIDFL EAKGYKEVKK DLIKLPKYS LFELENGRKR MLASAGELQK 1260
GNELALPSKY VNFLYLASHY EKLGKSPEDN EQKQLFVEQH KHYLDEIIEQ ISEFSKRVIL 1320
ADANLDKVL S AYNKHRDKPI REQAENIIHL FTLTNLGAPA AFKYFDTTID RKRYTSTKEV 1380
LDATLIHQSI TGLYETRIDL SQLGGD 1406

SEQ ID NO: 112      moltype = DNA length = 34
FEATURE            Location/Qualifiers
source             1..34
                  mol_type = unassigned DNA
                  organism = Homo sapiens

SEQUENCE: 112
caatctatga catcaattat tatacatcgg agcc 34

SEQ ID NO: 113      moltype = RNA length = 64
FEATURE            Location/Qualifiers
misc_feature       1..64
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source             1..64
                  mol_type = other RNA
                  organism = synthetic construct

SEQUENCE: 113
ggtgacatca attattatac atgttttaga gctagaataa gcaagttaa ataagcctag 60
tccg 64

SEQ ID NO: 114      moltype = DNA length = 49
FEATURE            Location/Qualifiers
misc_feature       1..49
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source             1..49
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 114
caatctatga catcaattat tatacatcgg agccctgcc aaaaatcaa 49

SEQ ID NO: 115      moltype = DNA length = 50
FEATURE            Location/Qualifiers
misc_feature       1..50
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source             1..50
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 115
caatctatga catcaattat tataacatcg gagccctgcc aaaaatcaa 50

SEQ ID NO: 116      moltype = DNA length = 36
FEATURE            Location/Qualifiers
misc_feature       1..36
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source             1..36
                  mol_type = other DNA
    
```

-continued

organism = synthetic construct
 SEQUENCE: 116
 caatctatga catcaattat tatgccaaaa atcaa 36

SEQ ID NO: 117 moltype = DNA length = 35
 FEATURE Location/Qualifiers
 misc_feature 1..35
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..35
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 117
 caatctatga catcgaggcc ctgccaaaa atcaa 35

SEQ ID NO: 118 moltype = DNA length = 31
 FEATURE Location/Qualifiers
 misc_feature 1..31
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..31
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 118
 caatctatga catgccctgc caaaaaatca a 31

SEQ ID NO: 119 moltype = DNA length = 30
 FEATURE Location/Qualifiers
 misc_feature 1..30
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..30
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 119
 caatctatga catcaattat tataaatcaa 30

SEQ ID NO: 120 moltype = DNA length = 25
 FEATURE Location/Qualifiers
 misc_feature 1..25
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..25
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 120
 caatctatga catccaaaa atcaa 25

SEQ ID NO: 121 moltype = DNA length = 19
 FEATURE Location/Qualifiers
 misc_feature 1..19
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..19
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 121
 caatctatga caaatcaa 19

SEQ ID NO: 122 moltype = DNA length = 46
 FEATURE Location/Qualifiers
 source 1..46
 mol_type = unassigned DNA
 organism = Homo sapiens

SEQUENCE: 122
 tatgtgcaat gaccactaca tcctcaaggg cagcaatcgg agccag 46

SEQ ID NO: 123 moltype = DNA length = 47
 FEATURE Location/Qualifiers
 misc_feature 1..47
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..47
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 123
 tatgtgcaat gaccactaca tccttcaagg gcagcaatcg gagccag 47

SEQ ID NO: 124 moltype = DNA length = 48

-continued

| | | |
|--|--|----|
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..48 | |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..48 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 124 | | |
| tatgtgcaat gaccactaca tcctctcaag ggcagcaatc ggagccag | | 48 |
| SEQ ID NO: 125 | moltype = DNA length = 18 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..18 | |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..18 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 125 | | |
| tatgtgcaat ggagccag | | 18 |
| SEQ ID NO: 126 | moltype = DNA length = 13 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..13 | |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..13 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 126 | | |
| tatgtgcaat gac | | 13 |
| SEQ ID NO: 127 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| source | 1..23 | |
| | mol_type = unassigned DNA | |
| | organism = Homo sapiens | |
| SEQUENCE: 127 | | |
| tgacatcaat tattatacat cgg | | 23 |
| SEQ ID NO: 128 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| source | 1..23 | |
| | mol_type = unassigned DNA | |
| | organism = Homo sapiens | |
| SEQUENCE: 128 | | |
| tgacatcaat tattatagat gga | | 23 |
| SEQ ID NO: 129 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| source | 1..23 | |
| | mol_type = unassigned DNA | |
| | organism = Homo sapiens | |
| SEQUENCE: 129 | | |
| tgacatcaat tattatgcat ggg | | 23 |
| SEQ ID NO: 130 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| source | 1..23 | |
| | mol_type = unassigned DNA | |
| | organism = Homo sapiens | |
| SEQUENCE: 130 | | |
| tgacataaat tattctacat ggg | | 23 |
| SEQ ID NO: 131 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| source | 1..23 | |
| | mol_type = unassigned DNA | |
| | organism = Homo sapiens | |
| SEQUENCE: 131 | | |
| tgaaatcaat tatcatagat cgg | | 23 |
| SEQ ID NO: 132 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| source | 1..23 | |
| | mol_type = unassigned DNA | |
| | organism = Homo sapiens | |
| SEQUENCE: 132 | | |
| ccaggctcca cccgactgga ggg | | 23 |

-continued

SEQ ID NO: 133 moltype = RNA length = 106
FEATURE Location/Qualifiers
misc_feature 1..106
 note = Description of Artificial Sequence: Synthetic
 polynucleotide
source 1..106
 mol_type = other RNA
 organism = synthetic construct

SEQUENCE: 133
ggccaggctc caccggactg gagttttaga gctagaaata gcaagttaaa ataaggctag 60
tccggtatca acttgaaaaa gtggcaccga gtcggtgctt tttttt 106

SEQ ID NO: 134 moltype = DNA length = 53
FEATURE Location/Qualifiers
source 1..53
 mol_type = unassigned DNA
 organism = Homo sapiens

SEQUENCE: 134
acttccaggc tccaccggac tggagggcga accccaaggg gacctcatgc agg 53

SEQ ID NO: 135 moltype = DNA length = 13
FEATURE Location/Qualifiers
misc_feature 1..13
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..13
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 135
acttccaggc tcc 13

SEQ ID NO: 136 moltype = DNA length = 30
FEATURE Location/Qualifiers
misc_feature 1..30
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..30
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 136
acttccaggc tccaccggac ctcatgcagg 30

SEQ ID NO: 137 moltype = DNA length = 36
FEATURE Location/Qualifiers
misc_feature 1..36
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..36
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 137
acttccaggc tccaccgcaa ggggacctca tgcagg 36

SEQ ID NO: 138 moltype = DNA length = 54
FEATURE Location/Qualifiers
misc_feature 1..54
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..54
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 138
acttccaggc tccaccggac ttggagggcg aaccgcaagg ggacctcatg cagg 54

SEQ ID NO: 139 moltype = DNA length = 43
FEATURE Location/Qualifiers
misc_feature 1..43
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..43
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 139
acttccaggc tccaccgaa ccccaagggg acctcatgca ggg 43

SEQ ID NO: 140 moltype = DNA length = 47
FEATURE Location/Qualifiers
misc_feature 1..47

-continued

| | | |
|--|---|----|
| | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..47 mol_type = other DNA organism = synthetic construct | |
| SEQUENCE: 140 | | |
| acttccaggc tccaccggac tcactatctt ctgggtcct ccatgtc | | 47 |
| SEQ ID NO: 141 | moltype = DNA length = 45 Location/Qualifiers | |
| FEATURE | 1..45 | |
| misc_feature | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..45 mol_type = other DNA organism = synthetic construct | |
| SEQUENCE: 141 | | |
| acttccaggc tccaccggac gaacccaag gggacctcat gcagg | | 45 |
| SEQ ID NO: 142 | moltype = AA length = 18 Location/Qualifiers | |
| FEATURE | 1..18 | |
| source | mol_type = protein organism = Homo sapiens | |
| SEQUENCE: 142 | | |
| LPGSTRLEGE PQGDLMQA | | 18 |
| SEQ ID NO: 143 | moltype = DNA length = 57 Location/Qualifiers | |
| FEATURE | 1..57 | |
| source | mol_type = unassigned DNA organism = Homo sapiens | |
| CDS | 2..55 | |
| SEQUENCE: 143 | | |
| acttccaggc tccaccggac tggagggcga accccaaggg gacctcatgc aggctcc | | 57 |
| SEQ ID NO: 144 | moltype = DNA length = 46 Location/Qualifiers | |
| FEATURE | 1..46 | |
| misc_feature | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..46 mol_type = other DNA organism = synthetic construct | |
| SEQUENCE: 144 | | |
| acttccaggc tccaccggaa cccaagggg acctcatgca ggctcc | | 46 |
| SEQ ID NO: 145 | moltype = DNA length = 43 Location/Qualifiers | |
| FEATURE | 1..43 | |
| misc_feature | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..43 mol_type = other DNA organism = synthetic construct | |
| SEQUENCE: 145 | | |
| acttccaggc tccaccggaa cccaagggg acctcatgca ggc | | 43 |
| SEQ ID NO: 146 | moltype = DNA length = 20 Location/Qualifiers | |
| FEATURE | 1..20 | |
| misc_feature | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..20 mol_type = other DNA organism = synthetic construct | |
| SEQUENCE: 146 | | |
| acttccaggc tccaccggac | | 20 |
| SEQ ID NO: 147 | moltype = DNA length = 40 Location/Qualifiers | |
| FEATURE | 1..40 | |
| misc_feature | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..40 mol_type = other DNA organism = synthetic construct | |
| SEQUENCE: 147 | | |
| acttccaggc tccaccggaa gggacctca tgcaggctcc | | 40 |

-continued

SEQ ID NO: 148 moltype = DNA length = 58
FEATURE Location/Qualifiers
misc_feature 1..58
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..58
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 148
acttcaggc tccaccgac ttggaggcg aacccaagg gacctcatg caggctcc 58

SEQ ID NO: 149 moltype = DNA length = 46
FEATURE Location/Qualifiers
misc_feature 1..46
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..46
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 149
acttcaggc tccaggcga cccaagggg acctcatgca ggctcc 46

SEQ ID NO: 150 moltype = DNA length = 32
FEATURE Location/Qualifiers
misc_feature 1..32
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..32
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 150
ggcgaacccc aaggggacct catgcaggct cc 32

SEQ ID NO: 151 moltype = DNA length = 32
FEATURE Location/Qualifiers
misc_feature 1..32
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..32
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 151
acttcaggc aaggggacct catgcaggct cc 32

SEQ ID NO: 152 moltype = DNA length = 33
FEATURE Location/Qualifiers
misc_feature 1..33
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..33
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 152
acttcaggc taaggggacc tcatgcaggc tcc 33

SEQ ID NO: 153 moltype = DNA length = 52
FEATURE Location/Qualifiers
source 1..52
 mol_type = unassigned DNA
 organism = Homo sapiens

SEQUENCE: 153
acttcaggc tccaccgac ttgaggcga accccaagg gacctcatg ag 52

SEQ ID NO: 154 moltype = DNA length = 34
FEATURE Location/Qualifiers
misc_feature 1..34
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..34
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 154
acttcaggc gaacccaag gggacctcat gcag 34

SEQ ID NO: 155 moltype = DNA length = 32
FEATURE Location/Qualifiers
misc_feature 1..32
 note = Description of Artificial Sequence: Synthetic

-continued

source oligonucleotide
 1..32
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 155
 acttccaggc tccacaaggg gacctcatgc ag 32

SEQ ID NO: 156 moltype = DNA length = 34
 FEATURE Location/Qualifiers
 misc_feature 1..34
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..34
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 156
 acttccaggc tccaccaag gggacctcat gcc 34

SEQ ID NO: 157 moltype = DNA length = 35
 FEATURE Location/Qualifiers
 misc_feature 1..35
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..35
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 157
 acttccaggc tccaccccaa ggggacctca tgcag 35

SEQ ID NO: 158 moltype = DNA length = 41
 FEATURE Location/Qualifiers
 misc_feature 1..41
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..41
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 158
 acttccaggc tccacccgaa cccaagggg acctcatgca g 41

SEQ ID NO: 159 moltype = DNA length = 50
 FEATURE Location/Qualifiers
 misc_feature 1..50
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..50
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 159
 acttccaggc tccacccgaa ggagggcgaa cccaagggg acctcatgca 50

SEQ ID NO: 160 moltype = DNA length = 50
 FEATURE Location/Qualifiers
 misc_feature 1..50
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..50
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 160
 acttccaggc tccacccgac tagggcgaac cccaagggga cctcatgcag 50

SEQ ID NO: 161 moltype = DNA length = 52
 FEATURE Location/Qualifiers
 misc_feature 1..52
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..52
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 161
 acttccaggc tccacccgac tgggagggcg aacccaagg gacctcatg ca 52

SEQ ID NO: 162 moltype = DNA length = 52
 FEATURE Location/Qualifiers
 misc_feature 1..52
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..52

-continued

```

mol_type = other DNA
organism = synthetic construct
SEQUENCE: 162
acttccaggc tccacccgac ttggagggcg aacccaagg ggacctcatg ca          52

SEQ ID NO: 163      moltype = DNA length = 46
FEATURE           Location/Qualifiers
misc_feature      1..46
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source           1..46
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 163
acttccaggc tccacccgag gcgaaccca aggggacctc atgcag          46

SEQ ID NO: 164      moltype = DNA length = 47
FEATURE           Location/Qualifiers
misc_feature      1..47
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source           1..47
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 164
acttccaggc tccacccgag gggaacccc aaggggacct catgcag          47

SEQ ID NO: 165      moltype = DNA length = 24
FEATURE           Location/Qualifiers
misc_feature      1..24
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source           1..24
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 165
acttccaggc tccacctcat gcag          24

SEQ ID NO: 166      moltype = DNA length = 29
FEATURE           Location/Qualifiers
misc_feature      1..29
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source           1..29
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 166
agggcgaacc ccaaggggac ctcatgcag          29

SEQ ID NO: 167      moltype = DNA length = 45
FEATURE           Location/Qualifiers
misc_feature      1..45
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source           1..45
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 167
caatctatga catcaattat tatcggagcc ctgccaaaaa atcaa          45

SEQ ID NO: 168      moltype = DNA length = 45
FEATURE           Location/Qualifiers
misc_feature      1..45
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source           1..45
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 168
caatctatga catcaattat catcggagcc ctgccaaaaa atcaa          45

SEQ ID NO: 169      moltype = DNA length = 42
FEATURE           Location/Qualifiers
misc_feature      1..42
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source           1..42
                  mol_type = other DNA
                  organism = synthetic construct

```

-continued

SEQUENCE: 169
caatctatga catcaattat cggagccctg ccaaaaaatc aa 42

SEQ ID NO: 170 moltype = DNA length = 48
FEATURE Location/Qualifiers
misc_feature 1..48
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..48
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 170
caatctatga catcaattat tatcatcgga gcctgcca aaaatcaa 48

SEQ ID NO: 171 moltype = DNA length = 33
FEATURE Location/Qualifiers
misc_feature 1..33
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..33
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 171
caatctatga caagagccct gccaaaaaat caa 33

SEQ ID NO: 172 moltype = DNA length = 52
FEATURE Location/Qualifiers
source 1..52
mol_type = unassigned DNA
organism = Homo sapiens

SEQUENCE: 172
ttctcaaggc agcatcatac ttccccacg gtgggacagc tgcctcct gg 52

SEQ ID NO: 173 moltype = DNA length = 46
FEATURE Location/Qualifiers
misc_feature 1..46
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..46
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 173
ttctcaaggc agcatcatac ttccctggga cagctgcct cctgg 46

SEQ ID NO: 174 moltype = DNA length = 49
FEATURE Location/Qualifiers
misc_feature 1..49
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..49
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 174
ttctcaaggc agcatcatac ttccaagggtg ggacagctgc cctcctgg 49

SEQ ID NO: 175 moltype = DNA length = 25
FEATURE Location/Qualifiers
misc_feature 1..25
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..25
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 175
ttctcaaggc agctgcctc cctgg 25

SEQ ID NO: 176 moltype = DNA length = 32
FEATURE Location/Qualifiers
misc_feature 1..32
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..32
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 176
ttctcaaggc agcatcatac ttccctcct gg 32

SEQ ID NO: 177 moltype = DNA length = 264
FEATURE Location/Qualifiers

-continued

misc_difference 38..227
 note = modified_base - a, c, t, g, unknown or other
 source 1..264
 mol_type = unassigned DNA
 organism = Homo sapiens

SEQUENCE: 177
 acaaagcgat tttgaaagat ggaagcgcgg tggctatnnn nnnnnnnnnn nnnnnnnnnn 60
 nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 120
 nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 180
 nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnggg gtgaaactaa 240
 actggtccac acggcggaag attg 264

SEQ ID NO: 178 moltype = DNA length = 257
 FEATURE Location/Qualifiers
 misc_feature 1..257
 note = Description of Artificial Sequence: Synthetic
 polynucleotide

misc_difference 38..227
 note = modified_base - a, c, t, g, unknown or other
 source 1..257
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 178
 acaaagcgat tttgaaagat ggaagcgcgg tggctatnnn nnnnnnnnnn nnnnnnnnnn 60
 nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 120
 nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 180
 nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnggg gtgaaactaa 240
 aacacggcgg aagattg 257

SEQ ID NO: 179 moltype = DNA length = 43
 FEATURE Location/Qualifiers
 misc_feature 1..43
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..43
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 179
 acaaagcgat tttgaaagat ggaagcgaca cggcggaaga ttg 43

SEQ ID NO: 180 moltype = DNA length = 44
 FEATURE Location/Qualifiers
 misc_feature 1..44
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..44
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 180
 acaaagcgat tttgaaagat ggaagcgcac acggcggaag attg 44

SEQ ID NO: 181 moltype = DNA length = 106
 FEATURE Location/Qualifiers
 misc_feature 1..106
 note = Description of Artificial Sequence: Synthetic
 polynucleotide

source 1..106
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 181
 acaaagcgat tttgaaagat ggaagcgaaa tagcaagtta aaataaggct agtccgttat 60
 caacttgaaa aagtgccacc gagtcggtgc acacggcgga agattg 106

SEQ ID NO: 182 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 182
 ggggtgggggg agtttgetcc tgg 23

SEQ ID NO: 183 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

-continued

source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 183
 ggatggaggg agtttgctcc tgg 23

SEQ ID NO: 184 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 184
 ggggagggga agtttgctcc tgg 23

SEQ ID NO: 185 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 185
 gacccctcc accccgctc cgg 23

SEQ ID NO: 186 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 186
 gaccccccc accccgcccc cgg 23

SEQ ID NO: 187 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 187
 gccccaccc accccgctc tgg 23

SEQ ID NO: 188 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 188
 ctccccacc accccgctc agg 23

SEQ ID NO: 189 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 189
 ggtgagtgag tgtgtgcgtg tgg 23

SEQ ID NO: 190 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..23
 mol_type = other DNA

-continued

organism = synthetic construct
 SEQUENCE: 190
 tgtgggtgag tgtgtgcgtg agg 23

SEQ ID NO: 191 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 191
 gagtccgagc agaagaagaa ggg 23

SEQ ID NO: 192 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 192
 gagttagagc agaagaagaa agg 23

SEQ ID NO: 193 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 193
 gaacctgagc tgctctgacg cgg 23

SEQ ID NO: 194 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 194
 ttggcagggg gtgggagggg agg 23

SEQ ID NO: 195 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 195
 gggagggaga gcttggcagg ggg 23

SEQ ID NO: 196 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 196
 gatggagcca gagagatcc tgg 23

SEQ ID NO: 197 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 197

-continued

cctgccaagc tctccctccc agg 23

SEQ ID NO: 198 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 198
 ctccctccca ggatcctctc tgg 23

SEQ ID NO: 199 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 199
 cctctaaggt ttgcttacga tgg 23

SEQ ID NO: 200 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 200
 ggttctggca aggagagaga tgg 23

SEQ ID NO: 201 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 201
 tctaaccccc acctcctggtt agg 23

SEQ ID NO: 202 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 202
 ttggcagggg gtgggagggg tgg 23

SEQ ID NO: 203 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 203
 ttggtagggg gtgggagggg tgg 23

SEQ ID NO: 204 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 204
 gggaagggga gcttggcagg tgg 23

-continued

SEQUENCE: 212
ggccgggaat caagagtcac ccagtgacca accatccctg taagcaaacc ttagaggttc 60
tggcaaggag agagatg 77

SEQ ID NO: 213 moltype = DNA length = 27
FEATURE Location/Qualifiers
misc_feature 1..27
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..27
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 213
ggccgggaat caagagtcac ccaggaa 27

SEQ ID NO: 214 moltype = DNA length = 79
FEATURE Location/Qualifiers
misc_feature 1..79
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..79
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 214
ggccgggaat caagagtcac ccagacctct ctggctccat cgtaagcaaa ccttagagggt 60
tctggcaagg agagagatg 79

SEQ ID NO: 215 moltype = DNA length = 28
FEATURE Location/Qualifiers
misc_feature 1..28
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..28
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 215
ggccgggaat caagagtcac cctaacag 28

SEQ ID NO: 216 moltype = DNA length = 66
FEATURE Location/Qualifiers
misc_feature 1..66
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..66
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 216
ggccgggaat caagagcctg gctccatcgt aagcaaacct tagaggttct ggcaaggaga 60
gagatg 66

SEQ ID NO: 217 moltype = DNA length = 47
FEATURE Location/Qualifiers
misc_feature 1..47
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..47
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 217
ggctccatcg taagcaaacc ttagaggttc tggcaaggag agagatg 47

SEQ ID NO: 218 moltype = DNA length = 78
FEATURE Location/Qualifiers
misc_feature 1..78
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..78
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 218
ggccgggaat caagagtcac ccagactctc tggctccatc gtaagcaaac ccttagagggt 60
ctggcaaggag gagagatg 78

SEQ ID NO: 219 moltype = DNA length = 80
FEATURE Location/Qualifiers
misc_feature 1..80
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..80

-continued

```

mol_type = other DNA
organism = synthetic construct
SEQUENCE: 219
ggccgggaat caagagtcac ccagagacag tgaccaacca tcgtaagcaa accttagagg 60
ttctggcaag gagagagatg 80

SEQ ID NO: 220      moltype = DNA length = 46
FEATURE           Location/Qualifiers
misc_feature      1..46
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source            1..46
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 220
ggtccatcgt aagcaaacct tagaggttct ggcaaggaga gagatg 46

SEQ ID NO: 221      moltype = DNA length = 69
FEATURE           Location/Qualifiers
misc_feature      1..69
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source            1..69
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 221
ggccgggaat caagagtcac ccactctccat cgtaagcaaa ccttagagggt tctggcaagg 60
agagagatg 69

SEQ ID NO: 222      moltype = DNA length = 76
FEATURE           Location/Qualifiers
misc_feature      1..76
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source            1..76
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 222
ggccgggaat caagagtcac ccagactctg gctccatcgt aagcaaacct tagaggttct 60
ggcaaggaga gagatg 76

SEQ ID NO: 223      moltype = DNA length = 50
FEATURE           Location/Qualifiers
misc_feature      1..50
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source            1..50
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 223
ggccgggaat caagagtcac ccagagacag tgaccaacca tcccatatca 50

SEQ ID NO: 224      moltype = DNA length = 59
FEATURE           Location/Qualifiers
misc_feature      1..59
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source            1..59
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 224
ggccgggaat caagagtcac cgtaagcaaa ccttagagggt tctggcaagg agagagatg 59

SEQ ID NO: 225      moltype = DNA length = 24
FEATURE           Location/Qualifiers
misc_feature      1..24
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source            1..24
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 225
aatgaccact acatccttca aggg 24

SEQ ID NO: 226      moltype = DNA length = 25
FEATURE           Location/Qualifiers
misc_feature      1..25
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide

```

-continued

source 1..25
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 226
 aatgaccact acatcctttc aaggg 25

SEQ ID NO: 227 moltype = DNA length = 26
 FEATURE Location/Qualifiers
 misc_feature 1..26
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..26
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 227
 aatgaccact acatcctttt caaggg 26

SEQ ID NO: 228 moltype = DNA length = 22
 FEATURE Location/Qualifiers
 misc_feature 1..22
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..22
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 228
 aatgaccact acatcctaag gg 22

SEQ ID NO: 229 moltype = DNA length = 21
 FEATURE Location/Qualifiers
 misc_feature 1..21
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..21
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 229
 aatgaccact acatcctagg g 21

SEQ ID NO: 230 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 230
 aatgaccact acatcctggg 20

SEQ ID NO: 231 moltype = DNA length = 42
 FEATURE Location/Qualifiers
 source 1..42
 mol_type = unassigned DNA
 organism = Homo sapiens

SEQUENCE: 231
 tatgtgcaat gaccactaca tctcgaaggg cagcaatcgg ag 42

SEQ ID NO: 232 moltype = DNA length = 45
 FEATURE Location/Qualifiers
 misc_feature 1..45
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..45
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 232
 tatgtgcaat gaccactaca tctcctcaa gggcagcaat cggag 45

SEQ ID NO: 233 moltype = DNA length = 30
 FEATURE Location/Qualifiers
 misc_feature 1..30
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..30
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 233
 tatgtgcaat gaccactaca tcaatcggag 30

-continued

| | | |
|--|--|----|
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..21 | |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..21 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 241 | | |
| tatgtgcaat gaccactaca t | | 21 |
| SEQ ID NO: 242 | moltype = DNA length = 53 | |
| FEATURE | Location/Qualifiers | |
| source | 1..53 | |
| | mol_type = unassigned DNA | |
| | organism = Homo sapiens | |
| SEQUENCE: 242 | | |
| tcatacagat gatgtctcat catcagagga gcgagaaggt aaagtcaaaa tca | | 53 |
| SEQ ID NO: 243 | moltype = DNA length = 32 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..32 | |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..32 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 243 | | |
| tcatacagat gatacaggta aagtcaaaaat ca | | 32 |
| SEQ ID NO: 244 | moltype = DNA length = 32 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..32 | |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..32 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 244 | | |
| tcatacaggt gatgaaggta aagtcaaaaat ca | | 32 |
| SEQ ID NO: 245 | moltype = DNA length = 50 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..50 | |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..50 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 245 | | |
| tcatacagat gatgtctcat catcagagcg agaaggtaaa gtcaaaaatca | | 50 |
| SEQ ID NO: 246 | moltype = DNA length = 48 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..48 | |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..48 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 246 | | |
| tcatacagat gatgtctcat catcagcgag aaggtaaagt caaaatca | | 48 |
| SEQ ID NO: 247 | moltype = DNA length = 52 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..52 | |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..52 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 247 | | |
| tcatacagat gatgtctcat catcaggagg cgagaaggta aagtcaaaaat ca | | 52 |
| SEQ ID NO: 248 | moltype = DNA length = 41 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..41 | |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..41 | |

-continued

```

mol_type = other DNA
organism = synthetic construct
SEQUENCE: 248
tcatacacagat gatgtctcgc gagaaggtaa agtcaaaatc a 41

SEQ ID NO: 249      moltype = DNA length = 32
FEATURE            Location/Qualifiers
misc_feature        1..32
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
source              1..32
                    mol_type = other DNA
                    organism = synthetic construct
SEQUENCE: 249
tcatacacagat gatgaaggta aagtcaaaat ca 32

SEQ ID NO: 250      moltype = DNA length = 29
FEATURE            Location/Qualifiers
misc_feature        1..29
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
source              1..29
                    mol_type = other DNA
                    organism = synthetic construct
SEQUENCE: 250
tcatacacagat gaaggtaaag tcaaaatca 29

SEQ ID NO: 251      moltype = DNA length = 42
FEATURE            Location/Qualifiers
misc_feature        1..42
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
source              1..42
                    mol_type = other DNA
                    organism = synthetic construct
SEQUENCE: 251
tcatacacagat gatgtctaca gatgaaggta aagtcaaaat ca 42

SEQ ID NO: 252      moltype = DNA length = 51
FEATURE            Location/Qualifiers
misc_feature        1..51
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
source              1..51
                    mol_type = other DNA
                    organism = synthetic construct
SEQUENCE: 252
tcatacacagat gatgtctcat catcaggagc gagaaggtaa agtcaaaatc a 51

SEQ ID NO: 253      moltype = DNA length = 34
FEATURE            Location/Qualifiers
misc_feature        1..34
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
source              1..34
                    mol_type = other DNA
                    organism = synthetic construct
SEQUENCE: 253
gtcatcctca tcctgataaa ctgcaaaagg ctga 34

SEQ ID NO: 254      moltype = DNA length = 606
FEATURE            Location/Qualifiers
source              1..606
                    mol_type = unassigned DNA
                    organism = Homo sapiens
SEQUENCE: 254
gctggtgtct ggggtctgtg ccccttcccc accccagccc accccagggt tctgttccat 60
tctcaggctg gtcacatggg tggctctagg gtgtcccatg agagatgcaa agcgcctgaa 120
ttttctgact cttcccatca gaccccccaa agacacatgt gaccaccac cccatctctg 180
accatgaggc caccctgagg tgetggggccc tgggcttcta ccctgcccag atcacactga 240
cctggcagcg ggatggcgag gaccaaactc aggacaccga gcttggggag accagaccag 300
caggagatag aaccttccag aagtgggcag ctgtgggtgt gccttctgga gaagagcaga 360
gatacacatg ccatgtacag catgagggggc tgcgaagccc cctcaccctg agatggggta 420
aggaggggga tgaggggtca tatctgttca tatctgttct cagggaaagc aggagccctt 480
ctggagccct tcagcagggt cagggcccct catcttcccc tcctttccca gagccatctt 540
cccagtcac catcccacatc gtgggcattg ttgtggcct ggctgtccta gcagttgtgg 600
tcacg 606

SEQ ID NO: 255      moltype = DNA length = 26

```

-continued

| | | |
|---|--|----|
| FEATURE | Location/Qualifiers | |
| source | 1..26 mol_type = unassigned DNA organism = Homo sapiens | |
| SEQUENCE: 255 | | |
| actaccacag ctccttctct gagtgg | | 26 |
| SEQ ID NO: 256 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..23 note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..23 mol_type = other DNA organism = synthetic construct | |
| SEQUENCE: 256 | | |
| actaccacag ctcctctgag tgg | | 23 |
| SEQ ID NO: 257 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| source | 1..23 mol_type = unassigned DNA organism = Homo sapiens | |
| SEQUENCE: 257 | | |
| gtagttggag ctggcggcgt agg | | 23 |
| SEQ ID NO: 258 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..23 note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..23 mol_type = other DNA organism = synthetic construct | |
| SEQUENCE: 258 | | |
| gtagttggag ctagcggcgt agg | | 23 |
| SEQ ID NO: 259 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| source | 1..23 mol_type = unassigned DNA organism = Homo sapiens | |
| SEQUENCE: 259 | | |
| gtagttggag ctggtggcgt agg | | 23 |
| SEQ ID NO: 260 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..23 note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..23 mol_type = other DNA organism = synthetic construct | |
| SEQUENCE: 260 | | |
| gtagttggag ctagtggcgt agg | | 23 |
| SEQ ID NO: 261 | moltype = DNA length = 28 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..28 note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..28 mol_type = other DNA organism = synthetic construct | |
| SEQUENCE: 261 | | |
| ccatacatTA aagatagTca tcttgggg | | 28 |
| SEQ ID NO: 262 | moltype = DNA length = 60 | |
| FEATURE | Location/Qualifiers | |
| source | 1..60 mol_type = unassigned DNA organism = Homo sapiens | |
| SEQUENCE: 262 | | |
| ccatacagtc agtatcaatt ctggaagaat ttccagacat taaagatagt catcttgggg | | 60 |
| SEQ ID NO: 263 | moltype = DNA length = 55 | |
| FEATURE | Location/Qualifiers | |
| source | 1..55 mol_type = unassigned DNA | |

-continued

organism = Homo sapiens
 SEQUENCE: 263
 ccatacagtc agtatcaatt ctggaagaat ttocagacat taaagatagt catct 55

SEQ ID NO: 264 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 264
 ccatacatta aagatagtca tct 23

SEQ ID NO: 265 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 source 1..23
 mol_type = unassigned DNA
 organism = Homo sapiens

SEQUENCE: 265
 agatgactat ctttaatgtc tgg 23

SEQ ID NO: 266 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 source 1..23
 mol_type = unassigned DNA
 organism = Homo sapiens

SEQUENCE: 266
 agatgactat ctttaatgta tgg 23

SEQ ID NO: 267 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 267
 gtagttggag ctgatggcgt agg 23

SEQ ID NO: 268 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 268
 gtagttggag ctggtagcgt agg 23

SEQ ID NO: 269 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 269
 gtagttggag ctggtgacgt agg 23

SEQ ID NO: 270 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 270
 gtagttggag ctaatggcgt agg 23

SEQ ID NO: 271 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic

-continued

| | |
|---------------------------|--|
| caaatgaatg gtgcacatca tgg | 23 |
| SEQ ID NO: 279 | moltype = DNA length = 23 |
| FEATURE | Location/Qualifiers |
| misc_feature | 1..23 |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide |
| source | 1..23 |
| | mol_type = other DNA |
| | organism = synthetic construct |
| SEQUENCE: 279 | |
| caaatgagtg atgcacatca tgg | 23 |
| SEQ ID NO: 280 | moltype = DNA length = 23 |
| FEATURE | Location/Qualifiers |
| misc_feature | 1..23 |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide |
| source | 1..23 |
| | mol_type = other DNA |
| | organism = synthetic construct |
| SEQUENCE: 280 | |
| caaaagaatg atgcacatca tgg | 23 |
| SEQ ID NO: 281 | moltype = DNA length = 23 |
| FEATURE | Location/Qualifiers |
| misc_feature | 1..23 |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide |
| source | 1..23 |
| | mol_type = other DNA |
| | organism = synthetic construct |
| SEQUENCE: 281 | |
| cgaatgaatg atgcacatca tgg | 23 |
| SEQ ID NO: 282 | moltype = DNA length = 23 |
| FEATURE | Location/Qualifiers |
| misc_feature | 1..23 |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide |
| source | 1..23 |
| | mol_type = other DNA |
| | organism = synthetic construct |
| SEQUENCE: 282 | |
| caaatgaatg atgcatgtca tgg | 23 |
| SEQ ID NO: 283 | moltype = DNA length = 23 |
| FEATURE | Location/Qualifiers |
| misc_feature | 1..23 |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide |
| source | 1..23 |
| | mol_type = other DNA |
| | organism = synthetic construct |
| SEQUENCE: 283 | |
| caaatgaatg atgcgctca tgg | 23 |
| SEQ ID NO: 284 | moltype = DNA length = 23 |
| FEATURE | Location/Qualifiers |
| misc_feature | 1..23 |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide |
| source | 1..23 |
| | mol_type = other DNA |
| | organism = synthetic construct |
| SEQUENCE: 284 | |
| caaatgaatg atgtacgtca tgg | 23 |
| SEQ ID NO: 285 | moltype = DNA length = 23 |
| FEATURE | Location/Qualifiers |
| misc_feature | 1..23 |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide |
| source | 1..23 |
| | mol_type = other DNA |
| | organism = synthetic construct |
| SEQUENCE: 285 | |
| caaatgaatg gtgcacgtca tgg | 23 |

-continued

| | | |
|---------------------------|--|----|
| SEQ ID NO: 286 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..23 | |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..23 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 286 | | |
| caaatgagtg atgcacgtca tgg | | 23 |
| SEQ ID NO: 287 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..23 | |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..23 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 287 | | |
| caaaagaatg atgcacgtca tgg | | 23 |
| SEQ ID NO: 288 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..23 | |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..23 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 288 | | |
| cgaatgaatg atgcacgtca tgg | | 23 |
| SEQ ID NO: 289 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| source | 1..23 | |
| | mol_type = unassigned DNA | |
| | organism = Homo sapiens | |
| SEQUENCE: 289 | | |
| atcataggtc gtcacgtta tgg | | 23 |
| SEQ ID NO: 290 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..23 | |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..23 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 290 | | |
| atcataggtt gtcacgtta tgg | | 23 |
| SEQ ID NO: 291 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| source | 1..23 | |
| | mol_type = unassigned DNA | |
| | organism = Homo sapiens | |
| SEQUENCE: 291 | | |
| atcataggtc gtctcgtta tgg | | 23 |
| SEQ ID NO: 292 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| misc_feature | 1..23 | |
| | note = Description of Artificial Sequence: Synthetic oligonucleotide | |
| source | 1..23 | |
| | mol_type = other DNA | |
| | organism = synthetic construct | |
| SEQUENCE: 292 | | |
| atcataggtt gtctcgtta tgg | | 23 |
| SEQ ID NO: 293 | moltype = DNA length = 23 | |
| FEATURE | Location/Qualifiers | |
| source | 1..23 | |
| | mol_type = unassigned DNA | |
| | organism = Homo sapiens | |
| SEQUENCE: 293 | | |
| ctggacaaga agagtacgt gcc | | 23 |

-continued

source 1..32
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 301
 acttccaggc tccaagggg acctcatgca gg 32

SEQ ID NO: 302 moltype = DNA length = 41
 FEATURE Location/Qualifiers
 misc_feature 1..41
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..41
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 302
 acttccaggc tccttaggag gcgaaccca agggacctc a 41

SEQ ID NO: 303 moltype = DNA length = 25
 FEATURE Location/Qualifiers
 misc_feature 1..25
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..25
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 303
 acttccaggc tccacctcat gcagg 25

SEQ ID NO: 304 moltype = DNA length = 32
 FEATURE Location/Qualifiers
 misc_feature 1..32
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..32
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 304
 acttccaggc tccacccccca agggacctca tg 32

SEQ ID NO: 305 moltype = DNA length = 35
 FEATURE Location/Qualifiers
 misc_feature 1..35
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..35
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 305
 acttccaggc tccaccaag gggacctcat gcagg 35

SEQ ID NO: 306 moltype = DNA length = 36
 FEATURE Location/Qualifiers
 misc_feature 1..36
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..36
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 306
 acttccaggc tccaccccaa gggacctca tgcagg 36

SEQ ID NO: 307 moltype = DNA length = 38
 FEATURE Location/Qualifiers
 misc_feature 1..38
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..38
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 307
 acttccaggc tccaccacc caaggggacc tcatgcag 38

SEQ ID NO: 308 moltype = DNA length = 38
 FEATURE Location/Qualifiers
 misc_feature 1..38
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

source 1..38
 mol_type = other DNA

-continued

organism = synthetic construct
 SEQUENCE: 308
 acttccaggc tccacccac ccaaggggac ctcatgca 38

SEQ ID NO: 309 moltype = DNA length = 40
 FEATURE Location/Qualifiers
 misc_feature 1..40
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..40
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 309
 acttccaggc tccacccacc ccaaggggac ctcatgcagg 40

SEQ ID NO: 310 moltype = DNA length = 45
 FEATURE Location/Qualifiers
 misc_feature 1..45
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..45
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 310
 acttccaggc tccacccggc gaacccaag gggacctcat gcagg 45

SEQ ID NO: 311 moltype = DNA length = 34
 FEATURE Location/Qualifiers
 misc_feature 1..34
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..34
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 311
 acttccaggc tccaccctgg ggacctcat cagg 34

SEQ ID NO: 312 moltype = DNA length = 42
 FEATURE Location/Qualifiers
 misc_feature 1..42
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..42
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 312
 acttccaggc tccacccgaa cccaagggg acctcatgca gg 42

SEQ ID NO: 313 moltype = DNA length = 31
 FEATURE Location/Qualifiers
 misc_feature 1..31
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..31
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 313
 acttccaggc tccacccgaa cctcatgcag g 31

SEQ ID NO: 314 moltype = DNA length = 35
 FEATURE Location/Qualifiers
 misc_feature 1..35
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..35
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 314
 acttccaggc tccacccgag gggacctcat gcagg 35

SEQ ID NO: 315 moltype = DNA length = 36
 FEATURE Location/Qualifiers
 misc_feature 1..36
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..36
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 315

-continued

```

acttcaggc tccaccccaa ggggacctca tgcagg                36

SEQ ID NO: 316      moltype = DNA length = 42
FEATURE            Location/Qualifiers
misc_feature       1..42
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..42
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 316
acttcaggc tccacccgaa ccccaagggg acctcatgca gg                42

SEQ ID NO: 317      moltype = DNA length = 43
FEATURE            Location/Qualifiers
misc_feature       1..43
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..43
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 317
acttcaggc tccacccgag accccaaggg gacctcatgc agg                43

SEQ ID NO: 318      moltype = DNA length = 48
FEATURE            Location/Qualifiers
misc_feature       1..48
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..48
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 318
acttcaggc tccacccgag ggcgaacccc aaggggacct catgcagg                48

SEQ ID NO: 319      moltype = DNA length = 30
FEATURE            Location/Qualifiers
misc_feature       1..30
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..30
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 319
acttcaggc tccacccgac ctcatgcagg                30

SEQ ID NO: 320      moltype = DNA length = 42
FEATURE            Location/Qualifiers
misc_feature       1..42
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..42
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 320
acttcaggc tccacccgac ccccaagggg acctcatgca gg                42

SEQ ID NO: 321      moltype = DNA length = 42
FEATURE            Location/Qualifiers
misc_feature       1..42
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..42
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 321
acttcaggc tccacccgac gaagggcccc aaggggacct ca                42

SEQ ID NO: 322      moltype = DNA length = 45
FEATURE            Location/Qualifiers
misc_feature       1..45
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..45
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 322
acttcaggc tccacccgac gaaccccaag gggacctcat gcagg                45

```

-continued

SEQ ID NO: 323 moltype = DNA length = 48
FEATURE Location/Qualifiers
misc_feature 1..48
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..48
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 323
acttccaggc tccaccggac ggcgaacccc aaggggacct catgcagg 48

SEQ ID NO: 324 moltype = DNA length = 51
FEATURE Location/Qualifiers
misc_feature 1..51
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..51
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 324
acttccaggc tccaccggac gtgcttgagg gcgaacccca aggggacctc a 51

SEQ ID NO: 325 moltype = DNA length = 47
FEATURE Location/Qualifiers
misc_feature 1..47
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..47
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 325
acttccaggc tccaccggac tcactatctt ctgggctcct ccatgtc 47

SEQ ID NO: 326 moltype = DNA length = 49
FEATURE Location/Qualifiers
misc_feature 1..49
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..49
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 326
acttccaggc tccaccggac ttggcgaacc ccaaggggac ctcatgcag 49

SEQ ID NO: 327 moltype = DNA length = 51
FEATURE Location/Qualifiers
misc_feature 1..51
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..51
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 327
acttccaggc tccaccggac ttgcagggcg aacccaagg ggacctcatg c 51

SEQ ID NO: 328 moltype = DNA length = 53
FEATURE Location/Qualifiers
misc_feature 1..53
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..53
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 328
acttccaggc tccaccggac ttggagggcg aacccaagg ggacctcatg cag 53

SEQ ID NO: 329 moltype = DNA length = 53
FEATURE Location/Qualifiers
misc_feature 1..53
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..53
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 329
acttccaggc tccaccggac ttggagggcg gaacccaagg gggacctcat gca 53

SEQ ID NO: 330 moltype = DNA length = 53
FEATURE Location/Qualifiers

-continued

```

misc_feature      1..53
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source            1..53
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 330
acttcaggc tccacccgac tgttgaggc cgaaccccaa ggggacctca tgc      53

SEQ ID NO: 331    moltype = DNA length = 502
FEATURE          Location/Qualifiers
misc_feature     1..502
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_difference  26..480
                  note = misc_feature - a, c, t, g, unknown or other
source           1..502
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 331
acttcaggc tccacccgac tggagnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 60
nnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 120
nnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 180
nnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 240
nnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 300
nnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 360
nnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 420
nnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 480
ggcgaacccc aaggggacct cc                                          502

SEQ ID NO: 332    moltype = DNA length = 72
FEATURE          Location/Qualifiers
source           1..72
                  mol_type = unassigned DNA
                  organism = Homo sapiens

SEQUENCE: 332
agctctccct cccaggatcc tctctggctc catcgtaagc aaaccttaga ggttctggca 60
aggagagaga tg                                                    72
    
```

- The invention claimed is:
1. A method for producing a genome-modified non-human animal, the method comprising:
 - providing a Cas9/RNA complex comprising a Cas9 protein and a guide RNA by providing the Cas9 protein and the guide RNA to a cell-free buffer wherein the guide RNA comprises a CRISPR RNA (crRNA) and a transactivating crRNA (tracrRNA), and wherein the guide RNA is provided in a molar excess over the Cas9 protein in the cell-free buffer;
 - introducing the Cas9/RNA complex into a non-human embryo, wherein the Cas9/RNA complex induces a modification at a target endogenous nucleic acid of the non-human embryo to provide for a genome modified embryo;
 - transferring the genome modified embryo into a foster mother; and
 - allowing the foster mother to produce a F0 animal having the modification at the target endogenous nucleic acid.
 2. The method of claim 1, further comprising:
 - mating the F0 animal with a wild-type non-human animal or another genome-modified non-human animal of the same species as the F0 animal having the modification at the target endogenous nucleic acid to provide a F1 offspring having the modification at the target endogenous nucleic acid.
 3. The method of claim 1, wherein introducing the Cas9/RNA complex into the non-human embryo comprises introducing the Cas9/RNA complex into a pronucleus of the non-human embryo.
 4. The method of claim 1, wherein introducing the Cas9/RNA complex into the non-human embryo comprises introducing the Cas9/RNA complex into a cytoplasm of the non-human embryo.
 5. The method of claim 1, wherein the Cas9/RNA complex is introduced into the non-human embryo by electroporation.
 6. The method of claim 1, wherein the Cas9/RNA complex is introduced into the non-human embryo by microinjection.
 7. The method of claim 1, wherein the crRNA is fused to the tracrRNA.
 8. The method of claim 1, wherein the crRNA comprises
 - i) a first portion to be hybridized with a portion of the tracrRNA, and
 - ii) a second portion complementary to the target endogenous nucleic acid of the non-human embryo; and
 wherein the target endogenous nucleic acid comprises
 - i) a first strand having a 20-base pair region complementary to the second portion of the crRNA and
 - ii) a second strand having a trinucleotide protospacer adjacent motif (PAM).
 9. The method of claim 1, wherein providing the Cas9/RNA complex comprises
 - preparing the Cas9 protein;
 - preparing the guide RNA; and
 - preparing the cell-free buffer.
 10. The method of claim 1, wherein the molar excess is at least two-fold molar excess.
 11. A method for producing a genome-modified non-human animal, the method comprising:

preparing a Cas9 protein;
preparing a single-guide RNA (sgRNA);
preparing a cell-free buffer;
preparing a Cas9/sgRNA complex by providing the Cas9
protein and the sgRNA to the cell-free buffer, wherein 5
the sgRNA is provided in a molar excess over the Cas9
protein; and
introducing the Cas9/sgRNA complex into a non-human
embryo, wherein the non-human embryo comprises a
target endogenous nucleic acid in the nucleus, and 10
wherein the Cas9/sgRNA complex induces modifica-
tion of the target endogenous nucleic acid of the
non-human embryo to provide for a genome modified
embryo.

12. The method of claim 11, wherein the molar excess is 15
an at least two-fold molar excess.

13. The method of claim 11, wherein introducing the
Cas9/sgRNA complex into the non-human embryo com-
prises introducing the Cas9/sgRNA complex into a pro-
nucleus of the non-human embryo. 20

14. The method of claim 11, wherein introducing the
Cas9/sgRNA complex into the non-human embryo com-
prises introducing the Cas9/sgRNA complex into a cyto-
plasm of the non-human embryo.

15. The method of claim 11, wherein the Cas9/sgRNA 25
complex is introduced into the non-human embryo by elec-
troporation.

16. The method of claim 11, wherein the Cas9/sgRNA
complex is introduced into the non-human embryo by
microinjection. 30

* * * * *