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(12) **United States Patent**
Kim et al.

(10) **Patent No.:** **US 12,612,632 B2**
(45) **Date of Patent:** ***Apr. 28, 2026**

(54) **COMPOSITIONS FOR INDUCING MODIFICATIONS OF TARGET ENDOGENOUS NUCLEIC ACID SEQUENCES IN NUCLEUSES OF EUKARYOTIC CELLS**

(71) Applicant: **ToolGen Incorporated**, Seoul (KR)

(72) Inventors: **Jin-Soo Kim**, Seoul (KR); **Seung Woo Cho**, Seoul (KR); **Sojung Kim**, Seoul (KR)

(73) Assignee: **ToolGen Incorporated**, Seoul (KR)

(*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 124 days.

This patent is subject to a terminal disclaimer.

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(65) **Prior Publication Data**

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Related U.S. Application Data

(63) Continuation of application No. 17/004,338, filed on Aug. 27, 2020, which is a continuation of application No. 14/685,568, filed on Apr. 13, 2015, now Pat. No. 10,851,380, which is a continuation of application No. PCT/KR2013/009488, filed on Oct. 23, 2013.

(60) Provisional application No. 61/837,481, filed on Jun. 20, 2013, provisional application No. 61/803,599, filed on Mar. 20, 2013, provisional application No. 61/717,324, filed on Oct. 23, 2012.

(51) **Int. Cl.**

C12N 9/22 (2006.01)
C12N 9/16 (2006.01)
C12N 15/10 (2006.01)
C12N 15/11 (2006.01)
C12N 15/52 (2006.01)
C12N 15/63 (2006.01)
C12N 15/82 (2006.01)
C12N 15/85 (2006.01)
C12N 15/90 (2006.01)

(52) **U.S. Cl.**

CPC **C12N 15/52** (2013.01); **C12N 9/16** (2013.01); **C12N 9/22** (2013.01); **C12N 15/102** (2013.01); **C12N 15/111** (2013.01); **C12N 15/63** (2013.01); **C12N 15/8216** (2013.01); **C12N 15/85** (2013.01); **C12N 15/907** (2013.01); **C12Y 301/21** (2013.01); **C12N 2310/10** (2013.01); **C12N 2310/20** (2017.05); **C12N 2310/531** (2013.01)

(58) **Field of Classification Search**

None
See application file for complete search history.

(56) **References Cited**

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Primary Examiner — Catherine Konopka

(74) *Attorney, Agent, or Firm* — Gemini Law LLP

(57) **ABSTRACT**

The present disclosure relates to targeted genome editing in eukaryotic cells or organisms. More particularly, the present disclosure provides for compositions that may induce modifications in target endogenous nucleic acid sequences in nucleuses of eukaryotic cells. The composition may comprise a single-chain guide RNA (sgRNA) and a *Streptococcus pyogenes* Cas9 protein. In some embodiments, the sgRNA and the Cas9 protein may be present in a molar ratio ranging from 29:14.0 to 29:1.4.

13 Claims, 61 Drawing Sheets

Specification includes a Sequence Listing.

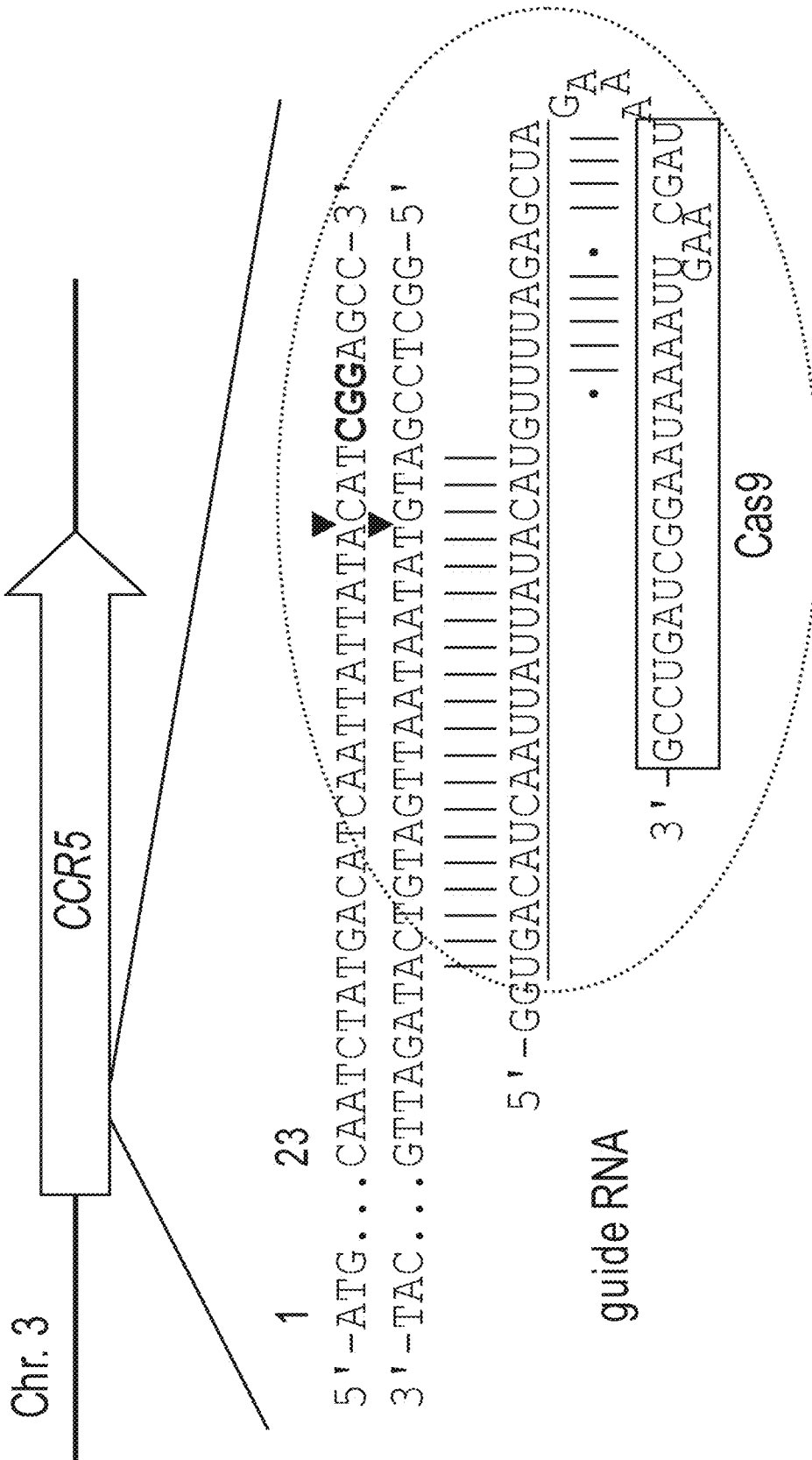
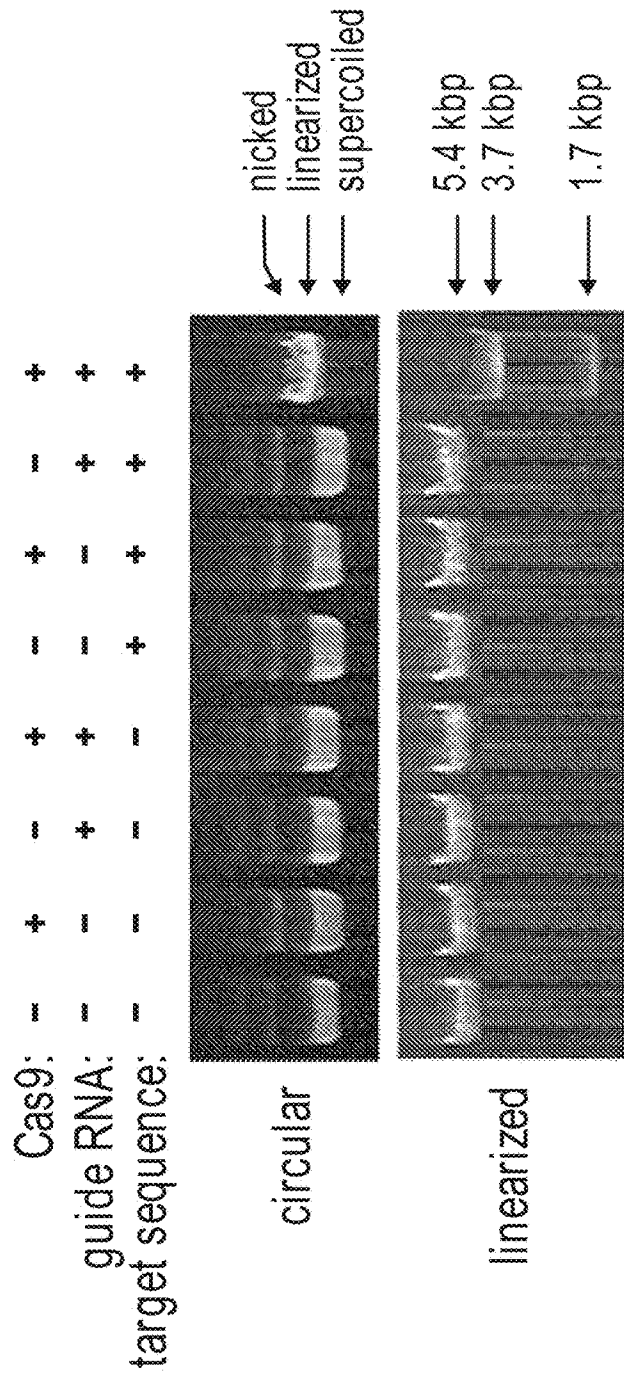
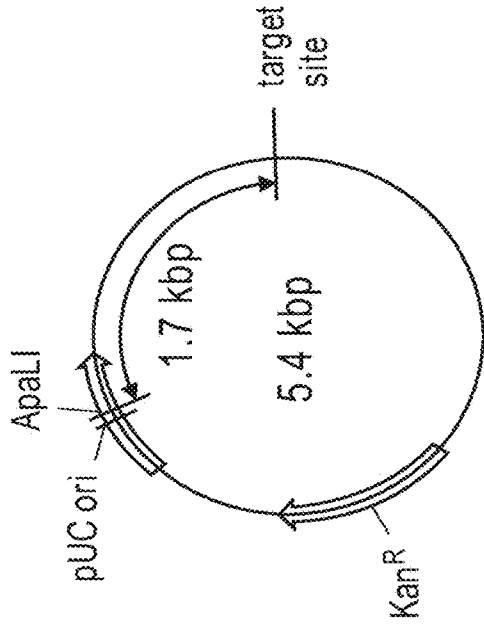


FIG. 1A

FIG. 1B



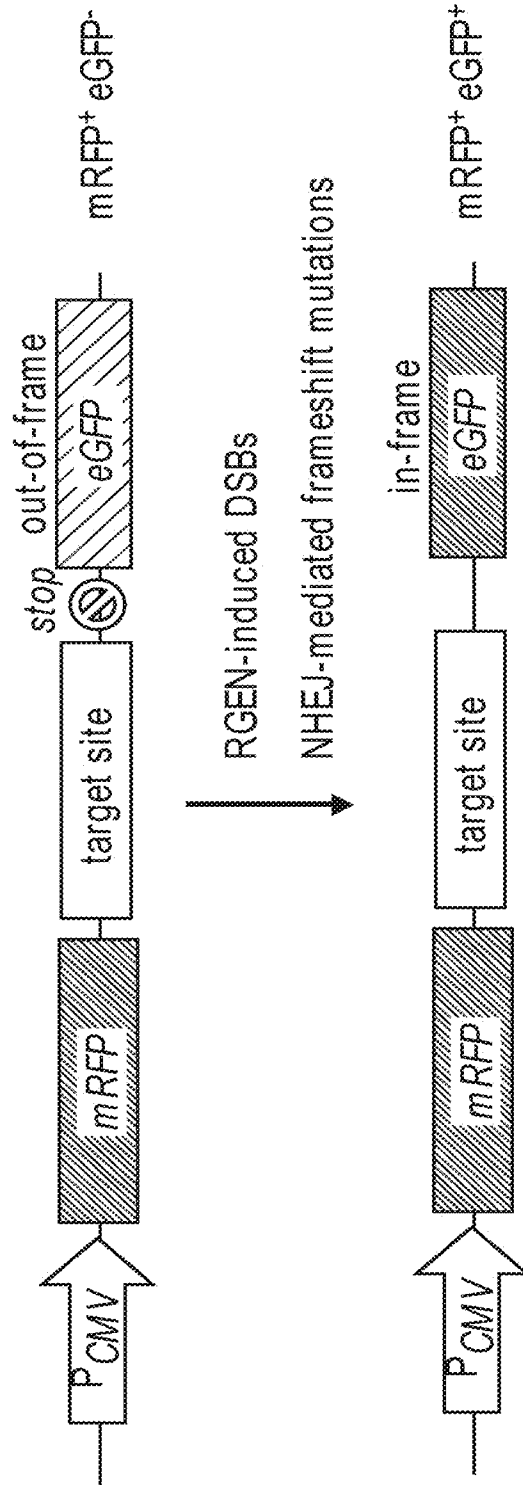


FIG. 2A

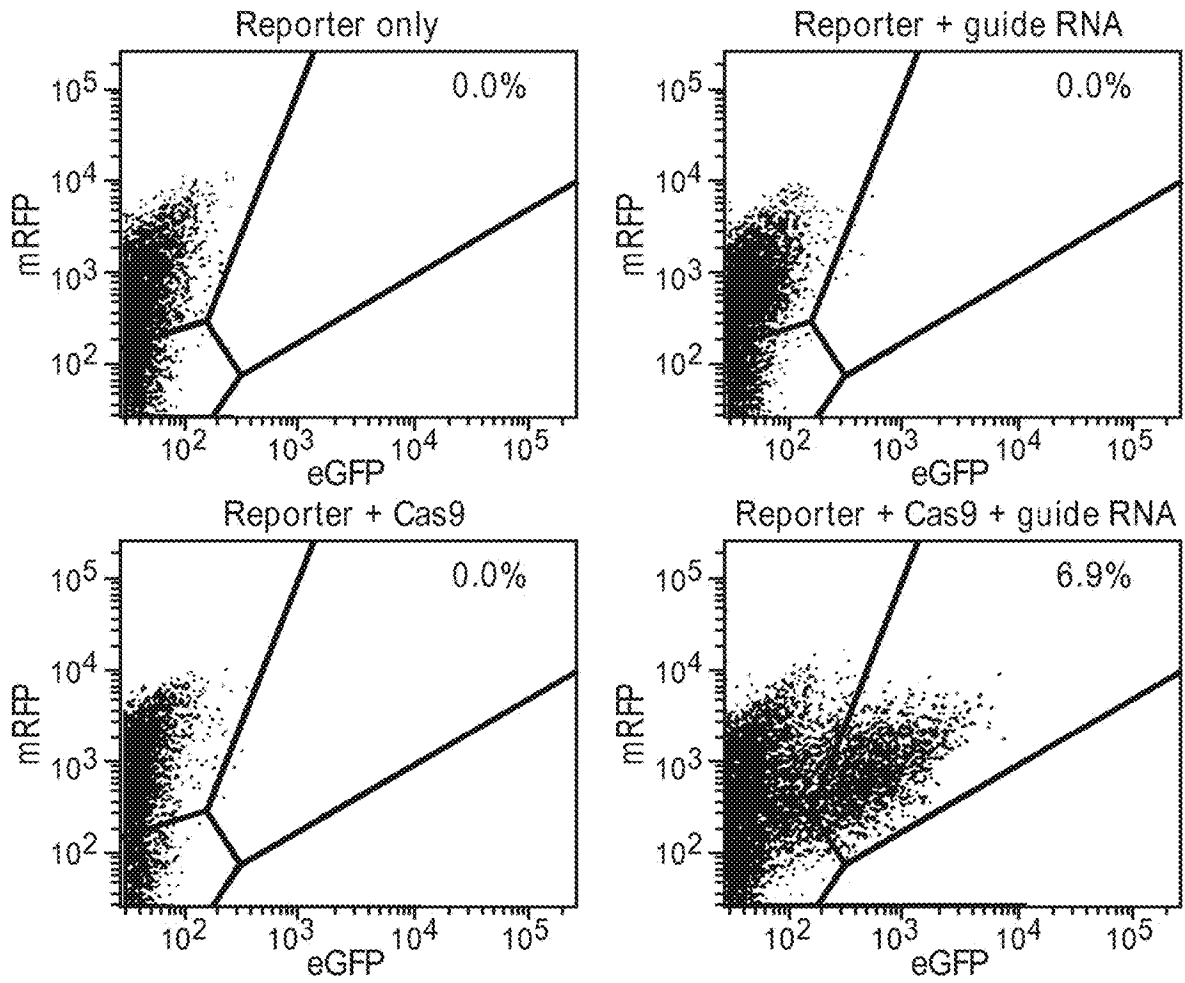


FIG. 2B

CCR5

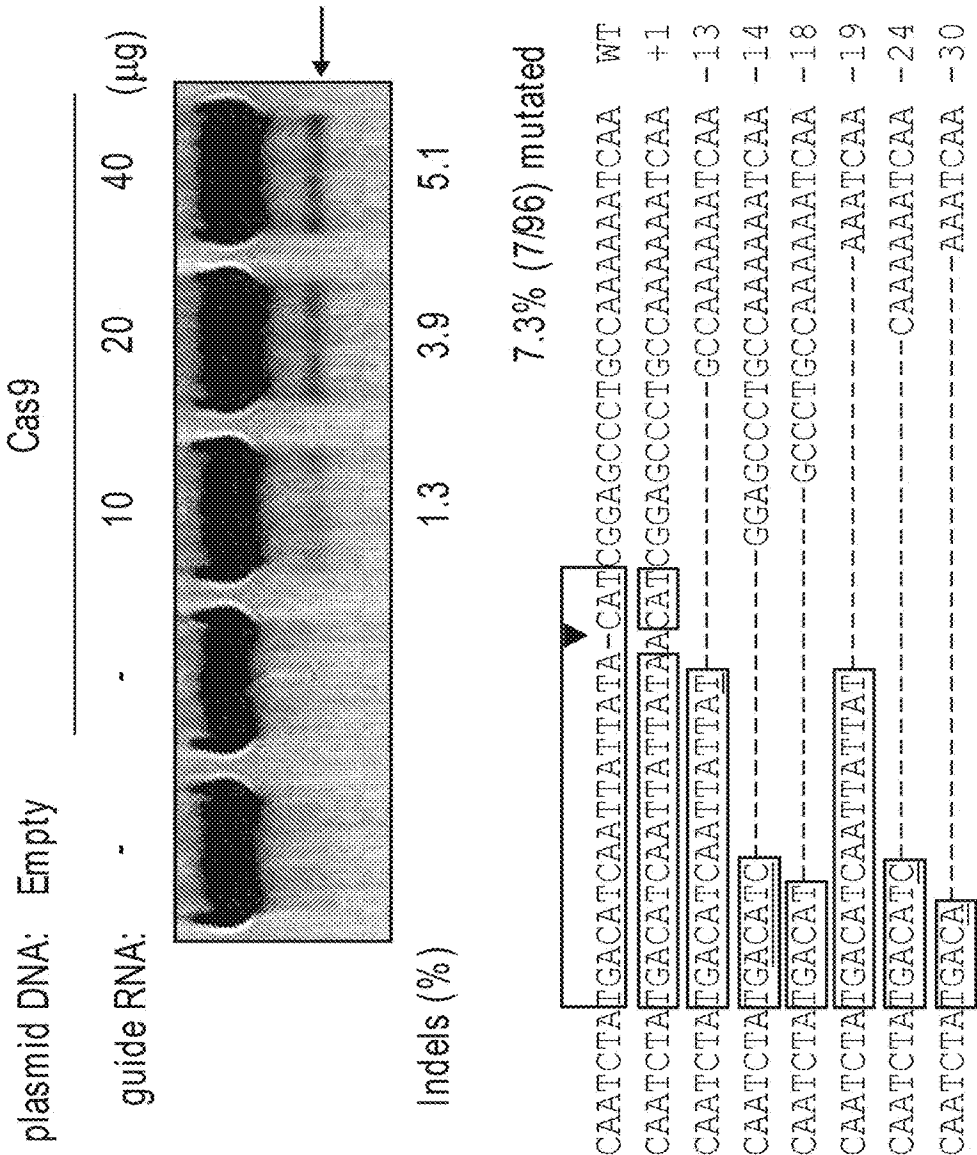
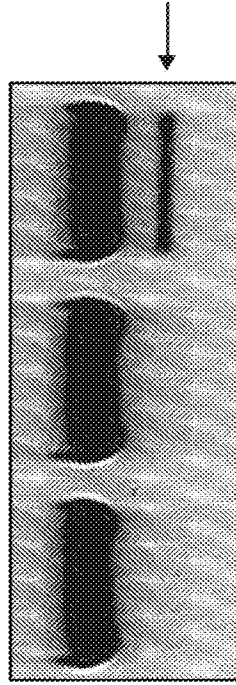


FIG. 3A

C4BPB

plasmid DNA: Empty Cas9

guide RNA: - +



Indels (%)

14

8.3% (4/48) mutated

TATGTGCAATGACCACTACATCCT--CAAAGGGCAGCAATCGGAGCCAG WT
 TATGTGCAATGACCACTACATCCTT-CAAAGGGCAGCAATCGGAGCCAG +1
 TATGTGCAATGACCACTACATCCTCTCAAAGGGCAGCAATCGGAGCCAG +2
 TATGTGCAATG AATG-----GAGCCAG -30
 TATGTGCAATGAC-----180 bp-----180

FIG. 3B

TGACATCAATTATTATACAT	▼	CGG	CCR5				
TGACATCAATTATTATA	g	AT	g	Ga	ADCY5		
TGACATCA	c	TTATTAT	g	CAT	g	GG	KCNJ6
TGACAT	a	AATTATT	c	TACAT	g	GG	CNTNAP2
TGA	a	ATCAATTAT	c	ATA	g	ATCGG	Chr. 5 N/A

FIG. 4A

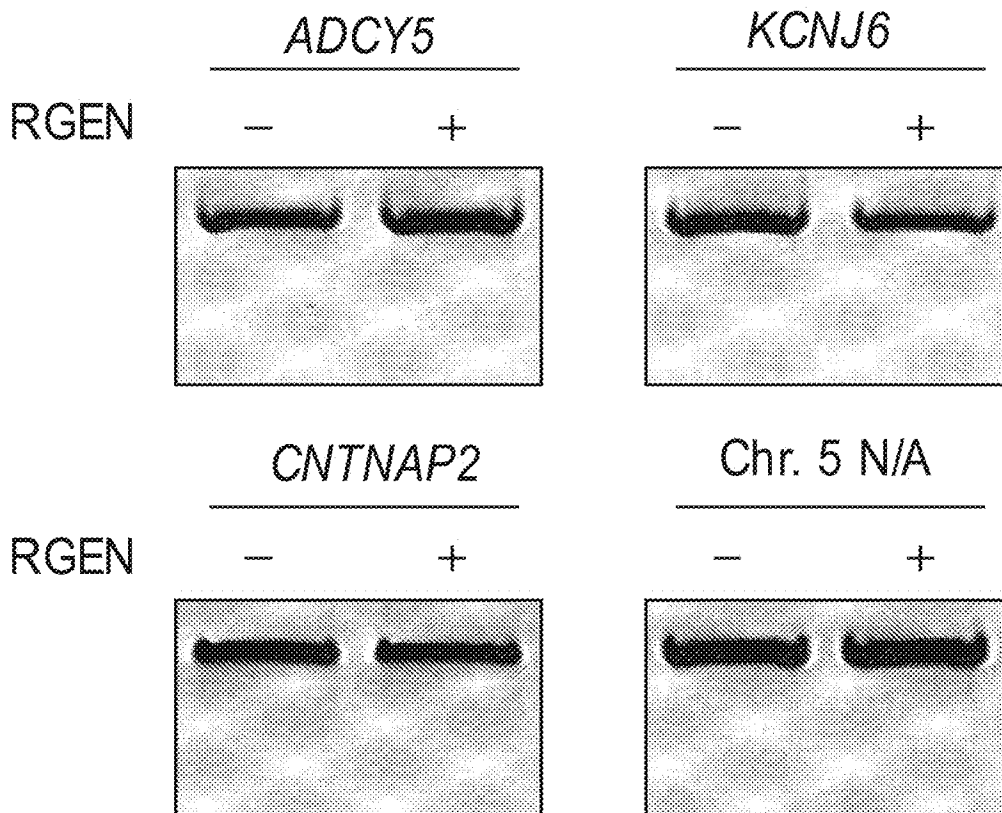


FIG. 4B

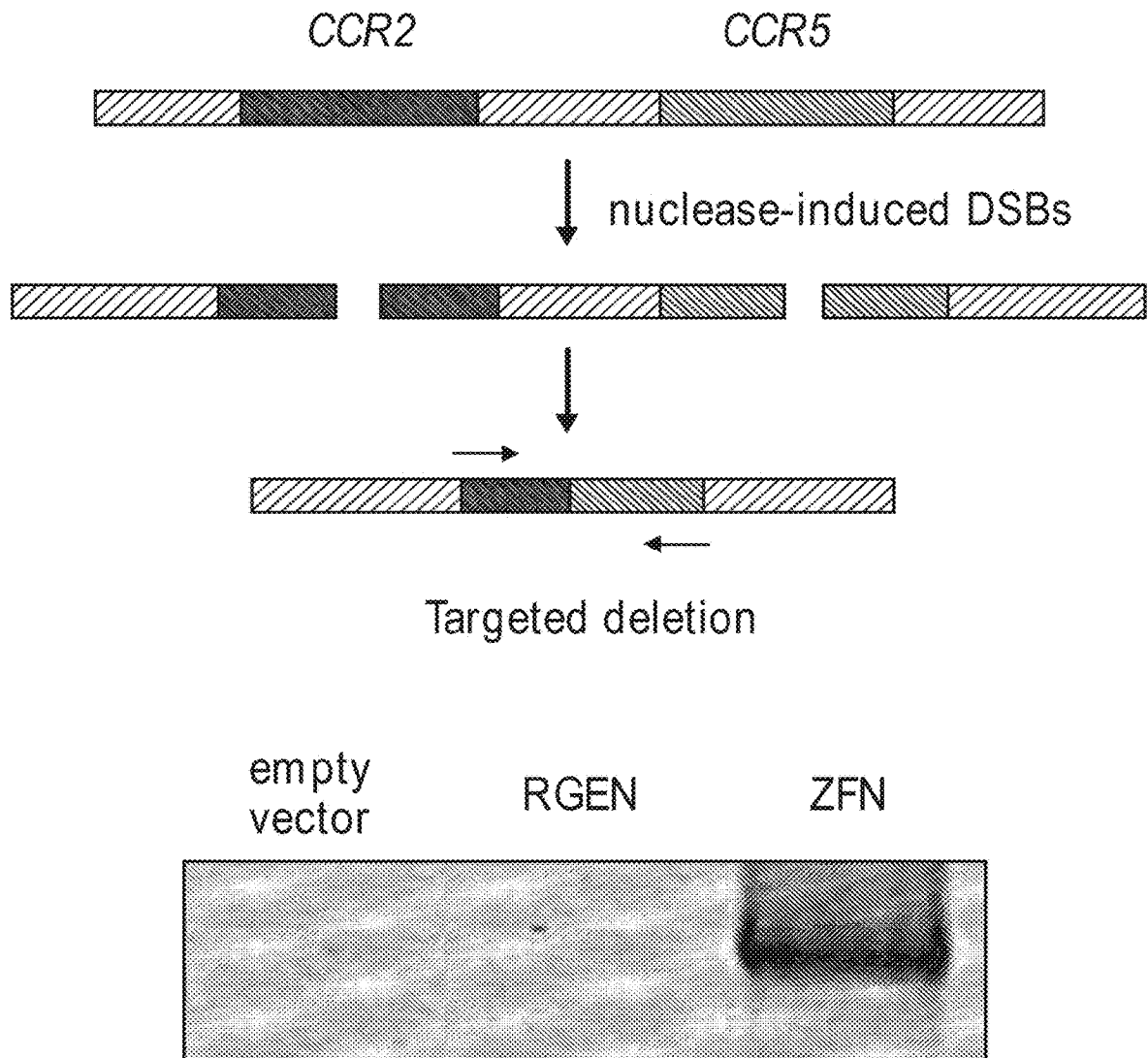


FIG. 4C

Foxn1

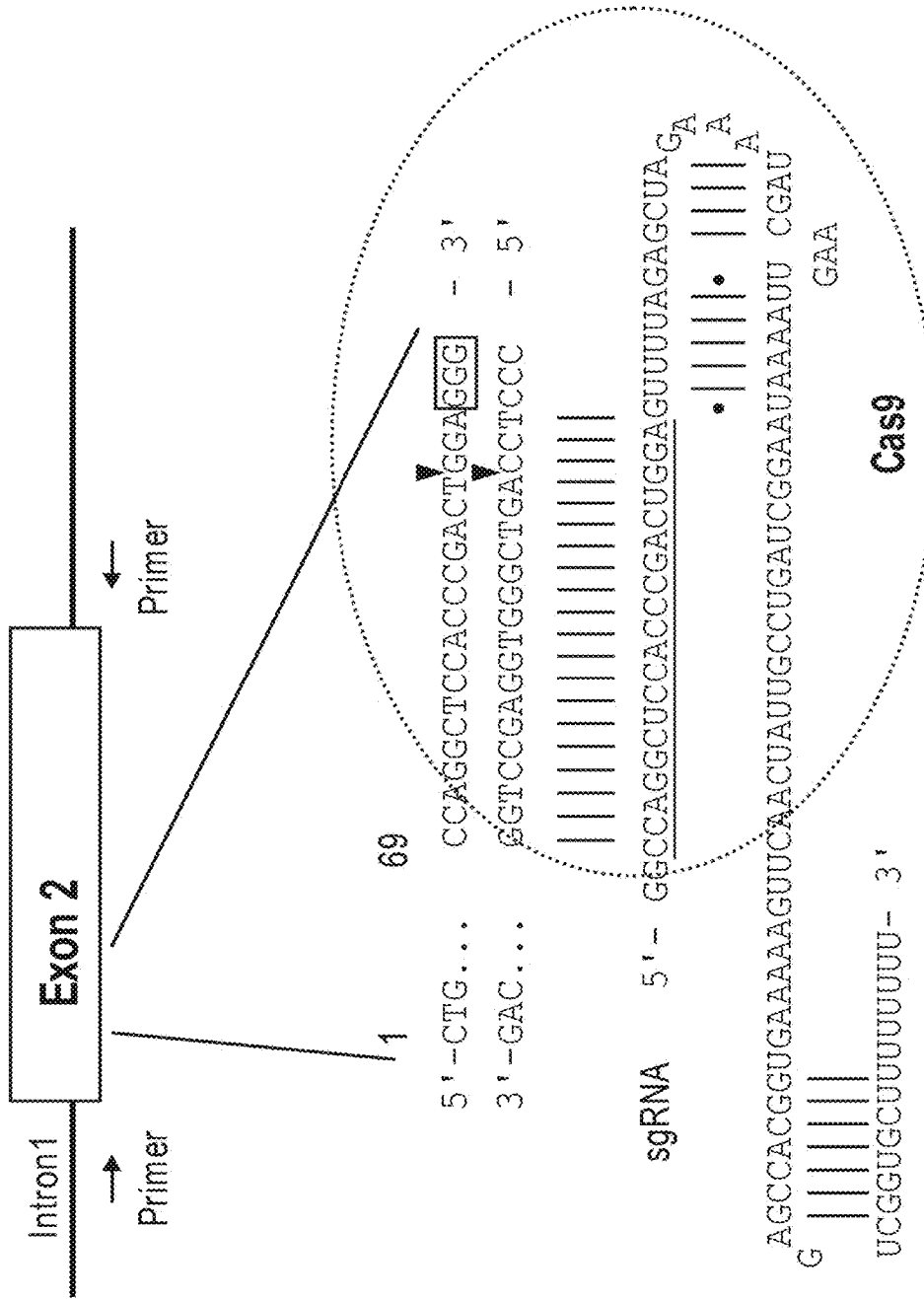


FIG. 5A

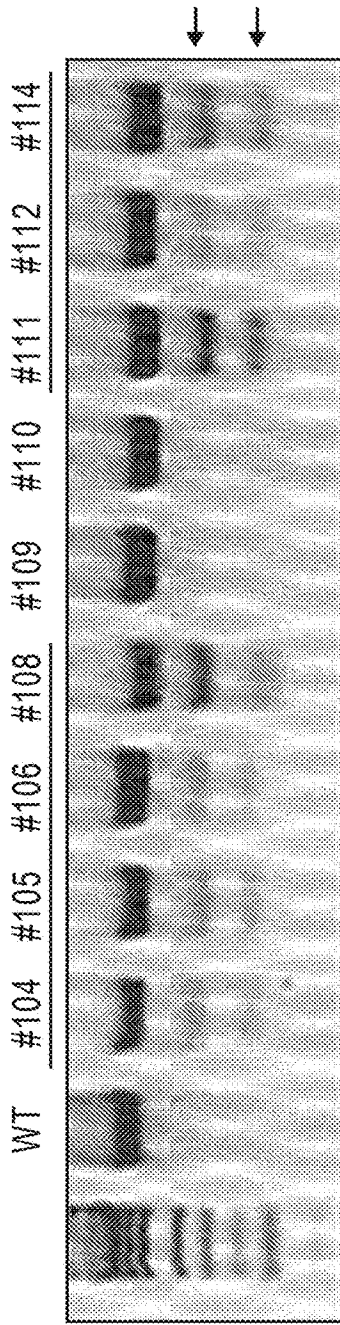


FIG. 5B

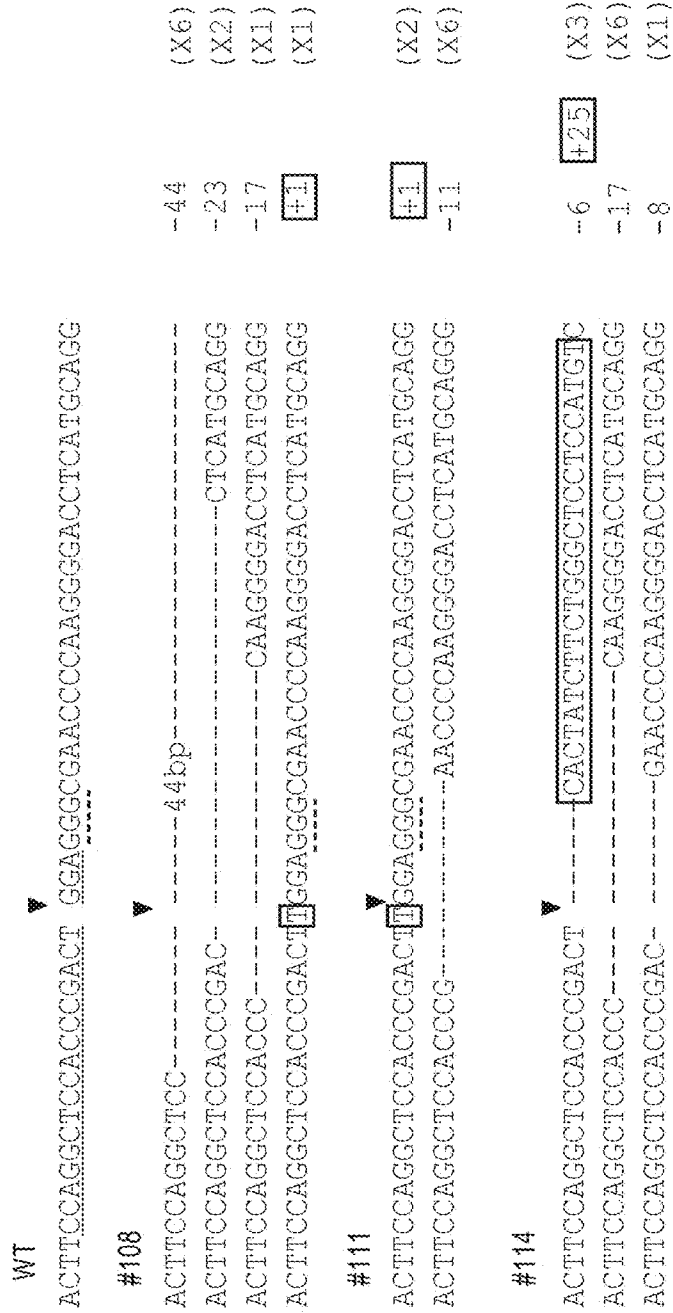


FIG. 5C

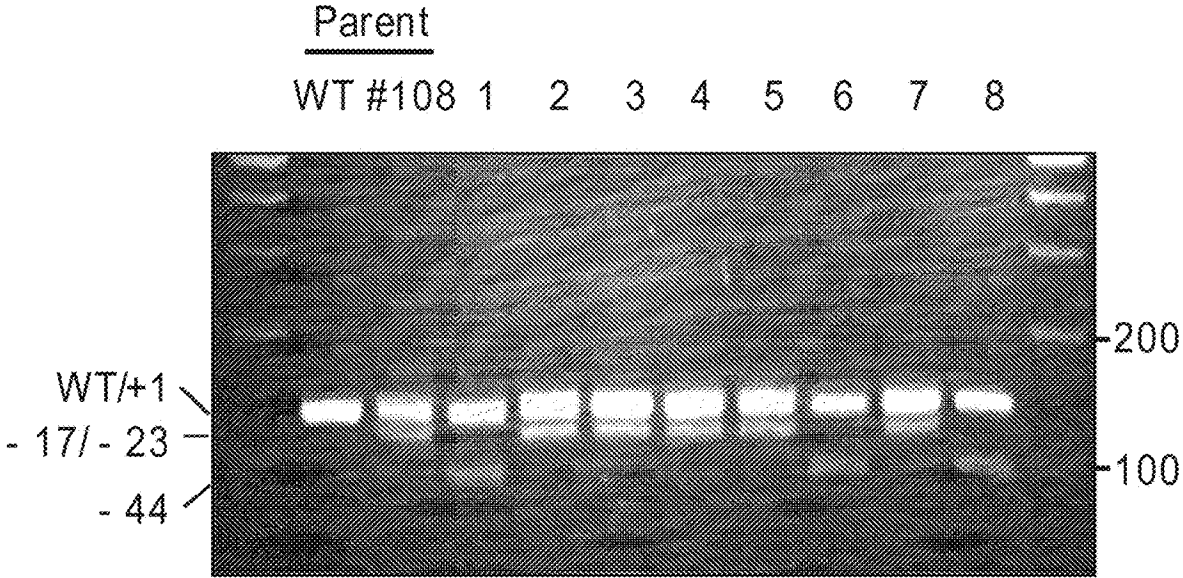


FIG. 5D

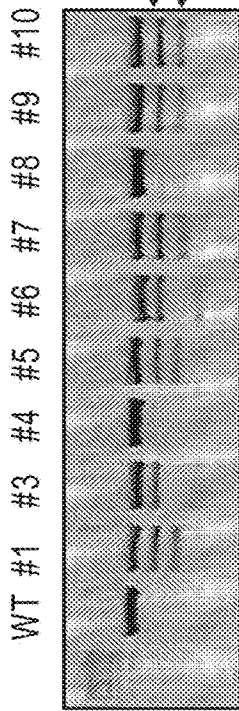


FIG. 6A

Cas9 mRNA (ng/μl)	sgRNA (ng/μl)	Tested embryos	Mutants (%)
10	1	27	9 (33)
10	10	49	28 (57)
10	100	45	41 (91)

FIG. 6B

sgRNA (ng/μl)	L	P	G	S	T	R	L	E	G	E	P	Q	G	D	L	M	Q	A	Mutants
100	A	CTT	CCA	GGC	TCC	ACC	CGA	CT	G	GAA	CCC	CAA	GGG	GAC	CTC	ATG	CAG	GCT	CC WT
	#1	A	CTT	CCA	GGC	TCC	ACC	CGA	---	--A	CCC	CAA	GGG	GAC	CTC	ATG	CAG	GCT	CC Δ11
	#3	A	CTT	CCA	GGC	TCC	ACC	CGA	---	--A	CCC	CAA	GGG	GAC	CTC	ATG	CAG	GC-	-- Δ11+Δ17
	#2	A	CTT	CCA	GGC	TCC	ACC	CGA	C-	---	---	---	---	---	---	---	---	---	-- Δ57
	#5	A	CTT	CCA	GGC	TCC	ACC	---	---	GAA	CCC	CAA	GGG	GAC	CTC	ATG	CAG	GCT	CC Δ11
	#11	A	CTT	CCA	GGC	TCC	ACC	CGA	CTG	---	---	CAA	GGG	GAC	CTC	ATG	CAG	GCT	CC Δ17
	#9	A	CTT	CCA	GGC	TCC	ACC	---	---	GAA	CCC	CAA	GGG	GAC	CTC	ATG	CAG	GCT	CC +1
	#11	A	CTT	CCA	GGC	TCC	A	---	---	GAA	CCC	CAA	GGG	GAC	CTC	ATG	CAG	GCT	CC Δ12
	#9	A	CTT	CCA	GGC	---	---	---	---	GAA	CCC	CAA	GGG	GAC	CTC	ATG	CAG	GCT	CC Δ72
	#1	A	CTT	CCA	GGC	T	---	---	---	---	---	AA	GGG	GAC	CTC	ATG	CAG	GCT	CC Δ25
		A	CTT	CCA	GGC	T	---	---	---	---	---	AA	GGG	GAC	CTC	ATG	CAG	GCT	CC Δ24

FIG. 6C

Pronucleus injection

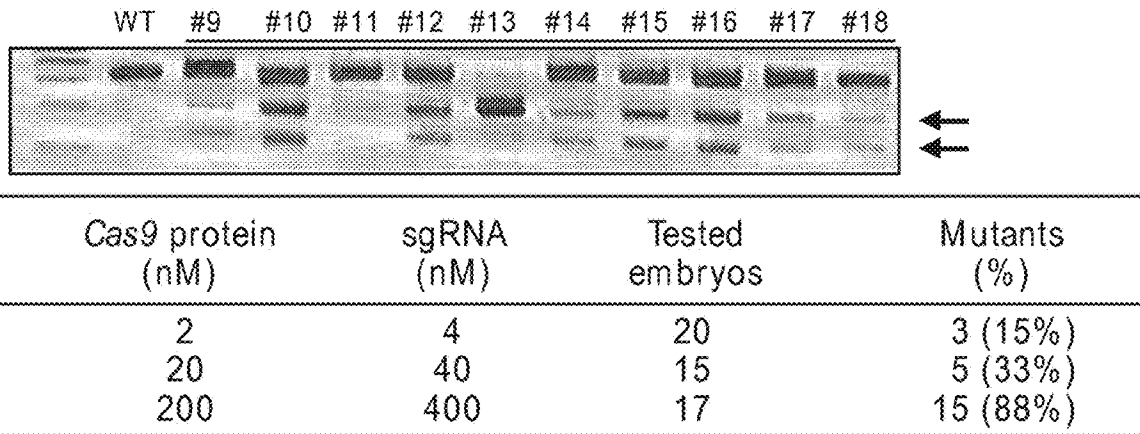


FIG. 7A

Intra-cytoplasmic injection

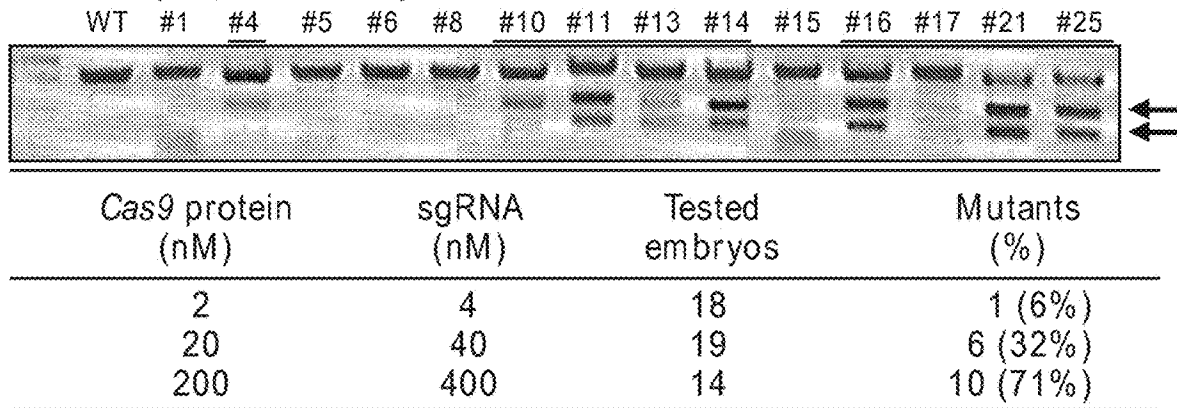


FIG. 7B

Sequence	Indels	Embryo no.
ACTTCCAGGCTCCACCCGACTGGAGGGCGGAACCCCAAGGGACCTCATGCAG	WT	
ACTTCCAGGCGAACCC-----AAGGGACCTCATGCAG	Δ18	2
ACTTCCAGGCTCCAC-----AAGGGACCTCATGCAG	Δ20	1
ACTTCCAGGCTCCACCC-----AAGGGACCTCATGCC	Δ19	1
ACTTCCAGGCTCCACCC-----CAAGGGACCTCATGCAG	Δ17	1
ACTTCCAGGCTCCACCCGA-----ACCCCAAGGGACCTCATGCAG	Δ11	3
ACTTCCAGGCTCCACCCGAA--GGAGGGCGAACCCCAAGGGACCTCATGCAG	Δ3+1	1
ACTTCCAGGCTCCACCCGACT--AGGGCGAACCCCAAGGGACCTCATGCAG	Δ2	1
ACTTCCAGGCTCCACCCGACTGGAGGGCGAACCCCAAGGGACCTCATGCAG	+1	1
ACTTCCAGGCTCCACCCGACTTGGAGGGCGAACCCCAAGGGACCTCATGCAG	+1	10
ACTTCCAGGCTCCACCCGA-----GGCGAACCCCAAGGGACCTCATGCAG	Δ6	1
ACTTCCAGGCTCCACCCGA-----GGCGAACCCCAAGGGACCTCATGCAG	Δ5	2
ACTTCCAGGCTCCACC-----TCATGCAG	Δ28	1
-----AGGGCGAACCCCAAGGGACCTCATGCAG	Δ126	1
Total		26

FIG. 7C

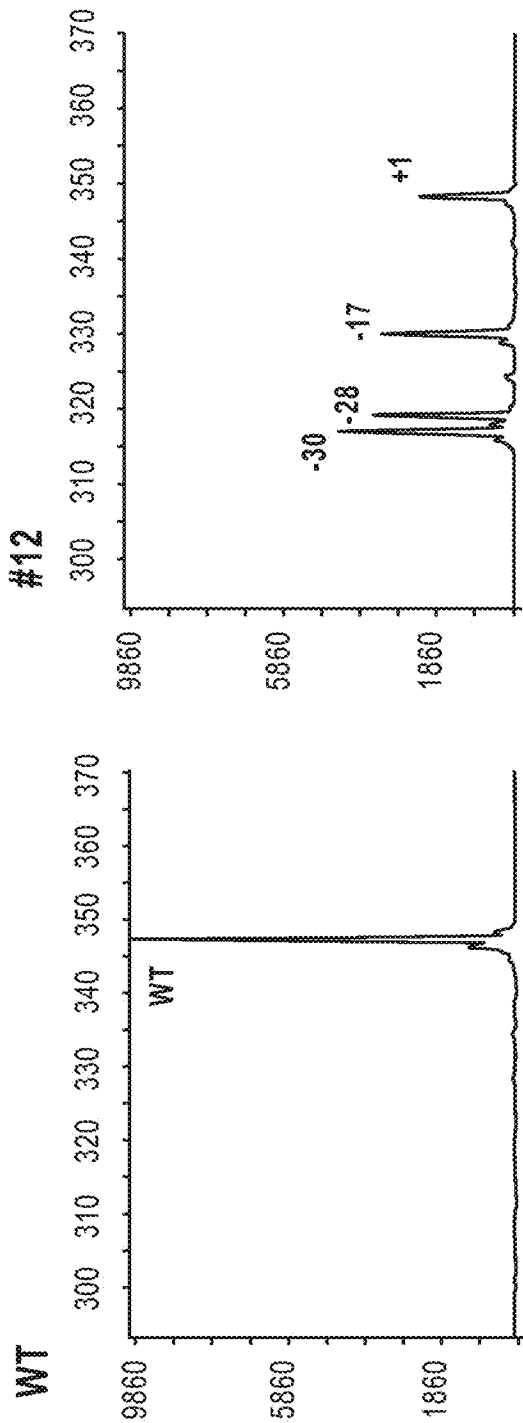
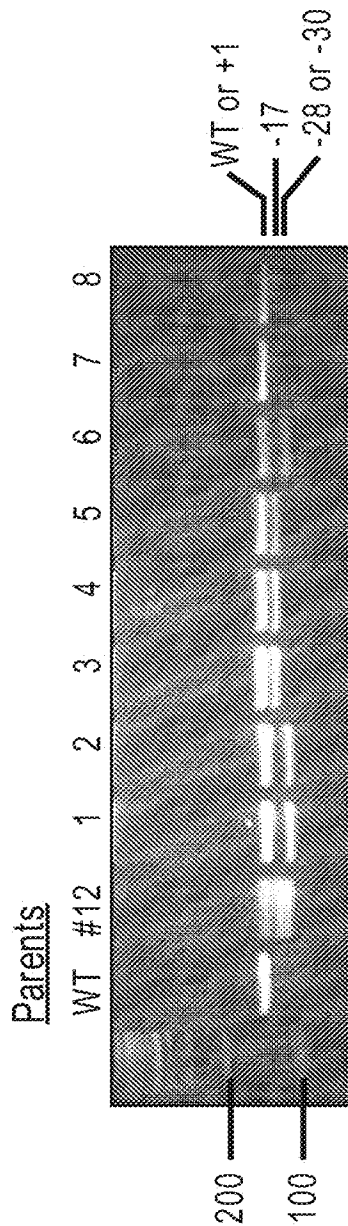


FIG. 8B



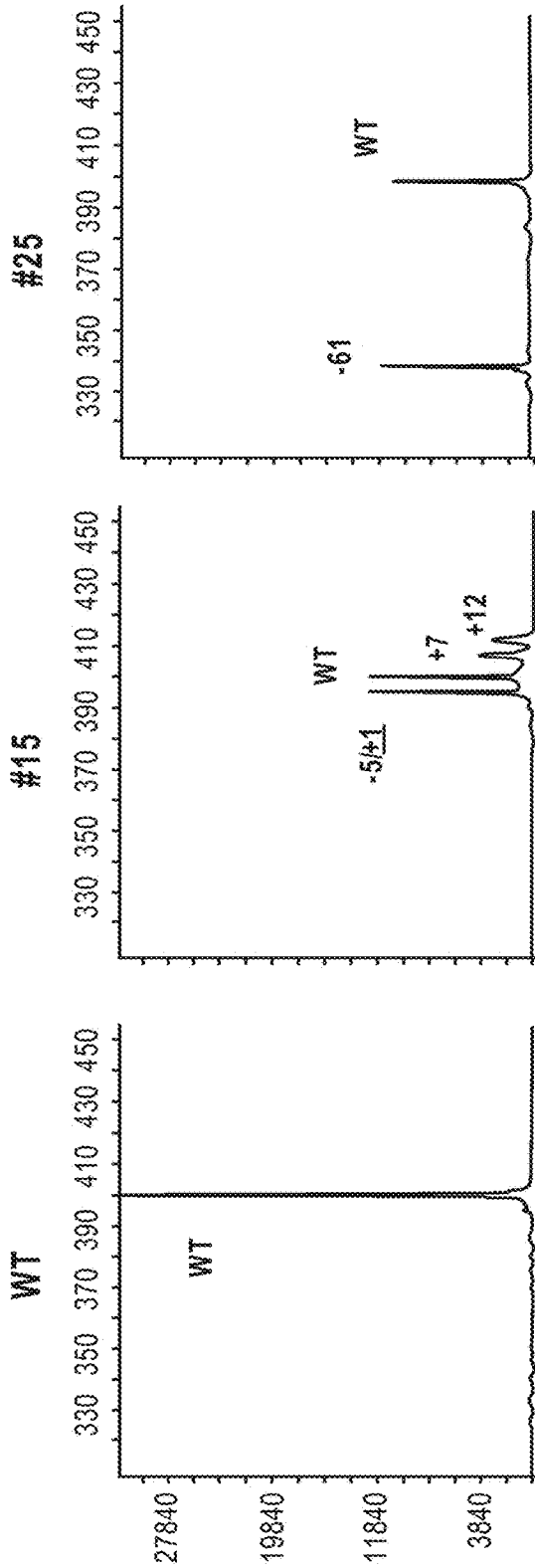


FIG. 9A

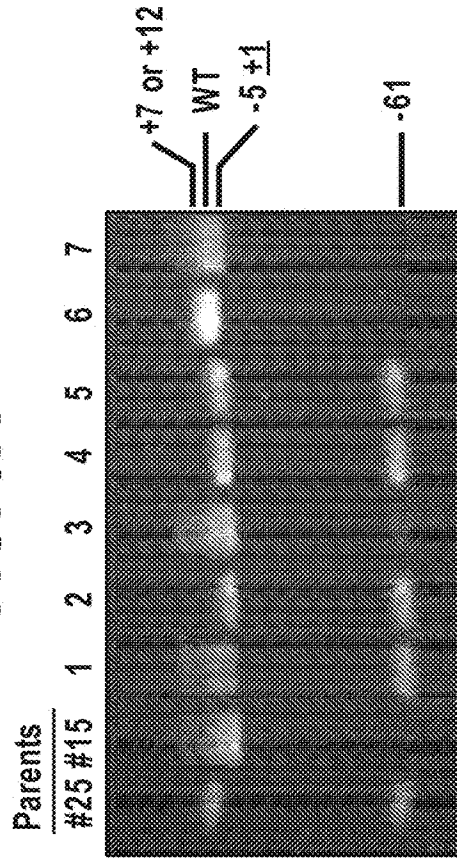
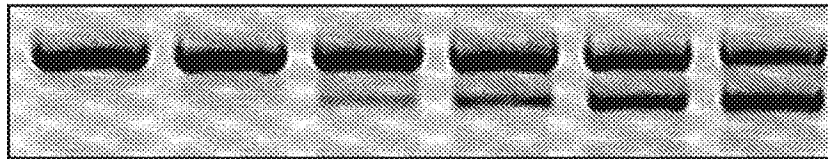


FIG. 9B

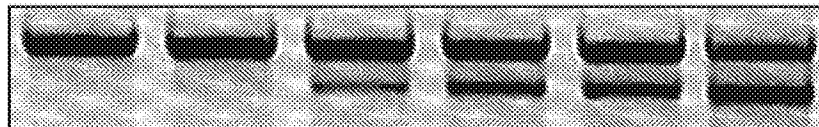
CCR5 #4

sgRNA	-	100 (29)	10(2.9)	30(8.8)	100(29)	50(15)	μg (μM)
Cas9 protein	225 (14)	-	22.5(1.4)	75(4.5)	225(14)	-	μg (μM)
Cas9 plasmid	-	-	-	-	-	5	μg



4.8 18 38 47 indel (%)

sgRNA	-	100 (29)				50(15)	μg (μM)
Cas9 protein	225 (14)	-	22.5(1.4)	75(4.5)	225(14)	-	μg (μM)
Cas9 plasmid	-	-	-	-	-	5	μg



12 30 33 43 indel (%)

crRNA	-	40(29)	20(15)	μg (μM)	
tracrRNA	-	80(29)	40(15)	μg (μM)	
Cas9 protein	225(14)	-	225(14)	-	μg (μM)
Cas9 plasmid	-	-	-	5	μg



9.4 42 indel (%)

FIG. 10A

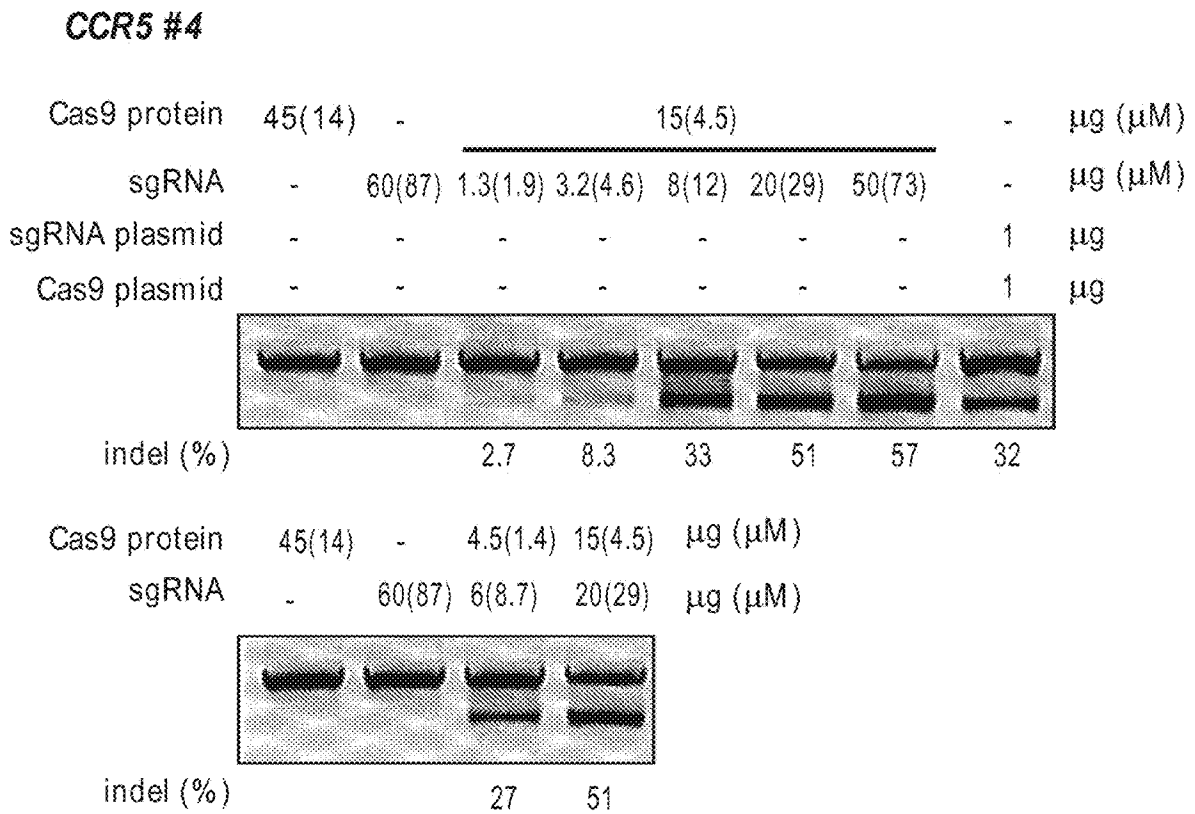


FIG. 10B

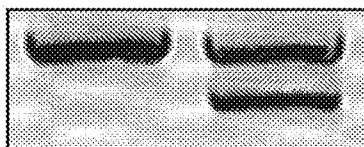
CCR5

CAATCTATGACATCAATTATTATA-CATCGGAGCCCTGCCAAAAAATCAA WT
 CAATCTATGACATCAATTATTAT-----CGGAGCCCTGCCAAAAAATCAA -4
 CAATCTATGACATCAATTAT-----CATCGGAGCCCTGCCAAAAAATCAA -4
 CAATCTATGACATCAATTAT-----CGGAGCCCTGCCAAAAAATCAA -7
 CAATCTATGACATCAATTATTAT--CATCGGAGCCCTGCCAAAAAATCAA -1
 CAATCTATGACATCAATTATTATAACATCGGAGCCCTGCCAAAAAATCAA +1
 CAATCTATGACAA-----GAGCCCTGCCAAAAAATCAA -17, +1

FIG. 10C

ABCC11

Cas9 protein - 15(4.5) μg (μM)
 sgRNA - 20(29) μg (μM)



Indel (%) 35

FIG. 10D

ABCC11

TTCTCAAGGCAGCATCATACTTCCCCCACGGTGGGACAGCTGCCCTCCCTGG WT
 TTCTCAAGGCAGCATCATACTTCC-----CTGGGACAGCTGCCCTCCCTGG -6
 TTCTCAAGGCAGCATCATACTT---CACGGTGGGACAGCTGCCCTCCCTGG -3
 TTCTCAAGGCAGC-----TGCCCTCCCTGG -29
 TTCTCAAGGCAGCATCATACTT-----CCCTCCCTGG -20
 TTCTCAAGGCAGCATCATACTT-----CCCTCCCTGG -20
 TTCTC----- -256

FIG. 10E

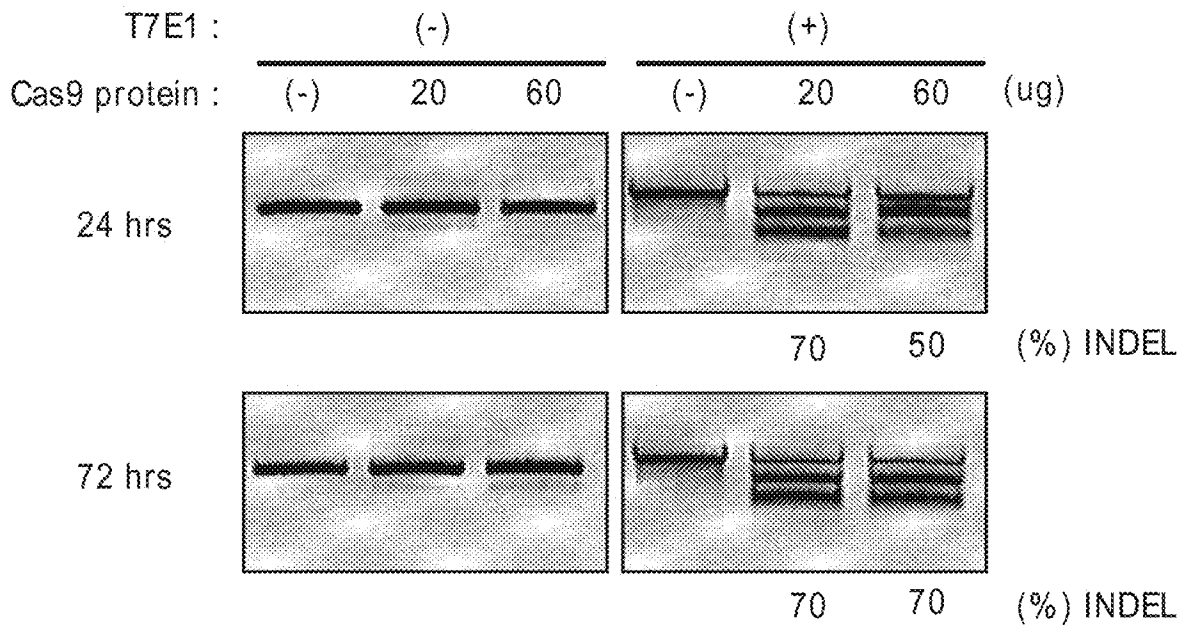


FIG. 11

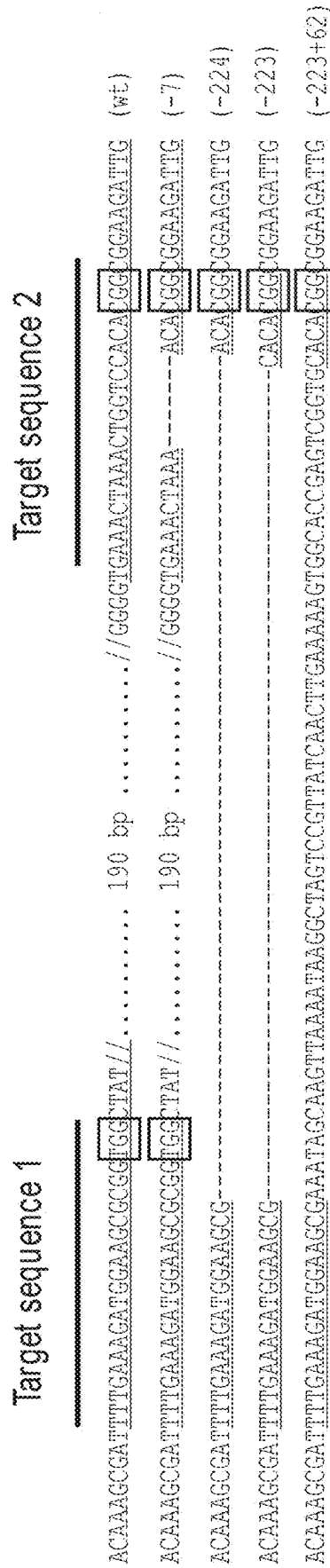


FIG. 12

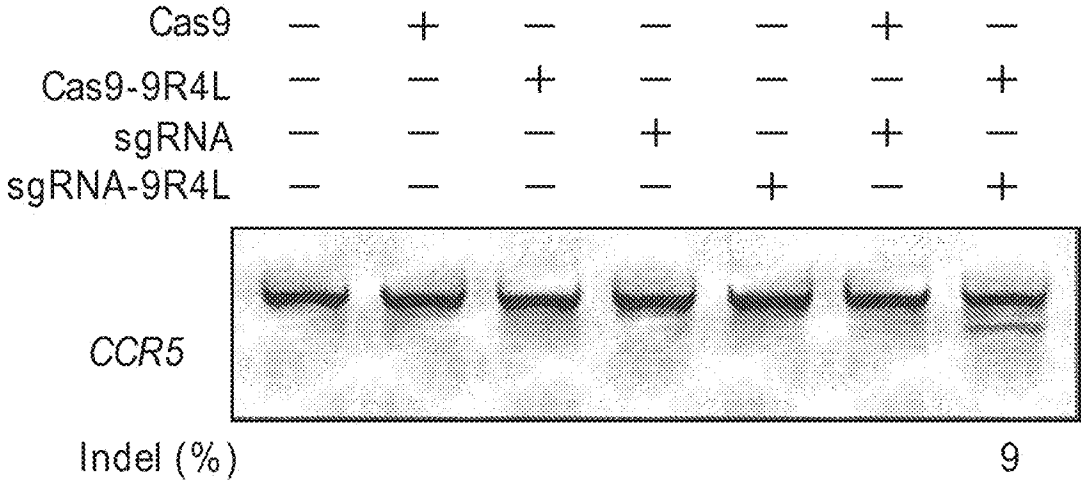


FIG. 13

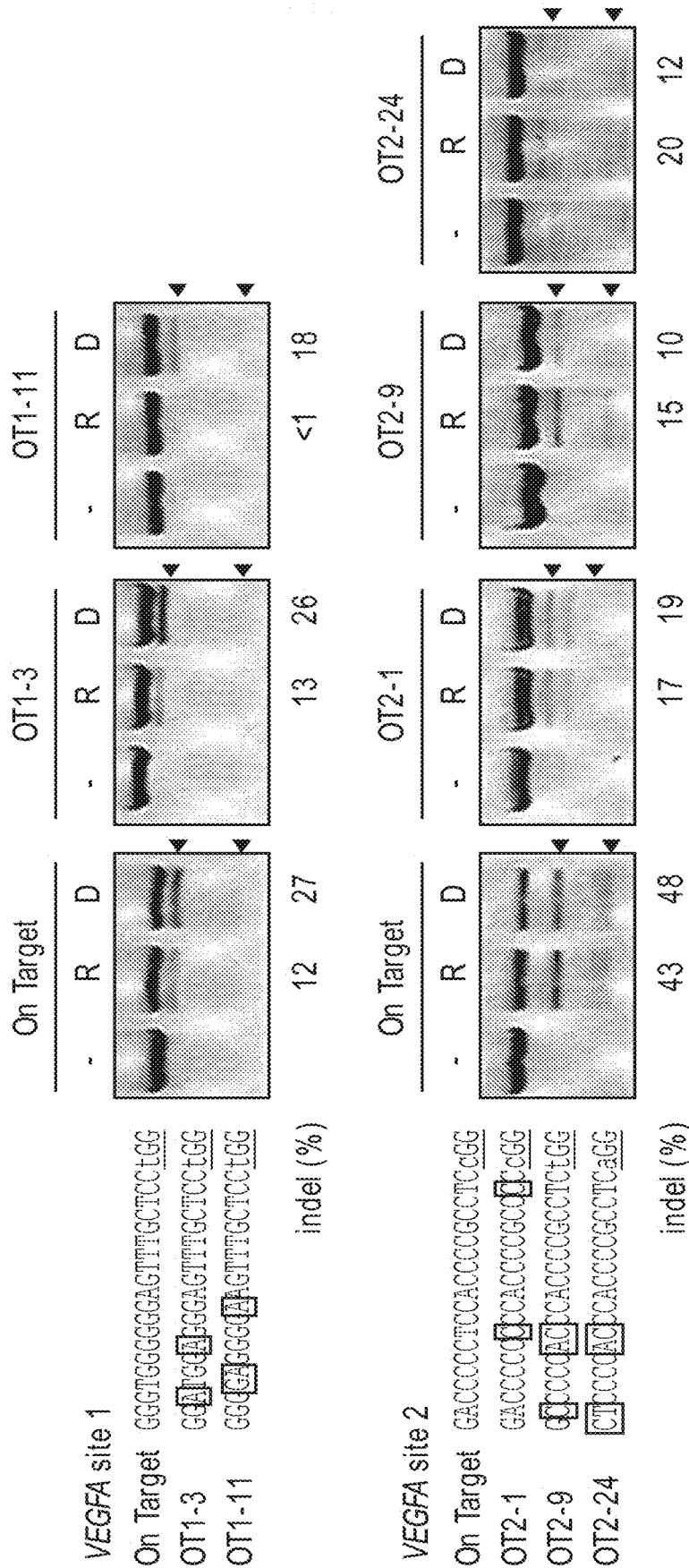


FIG. 14A

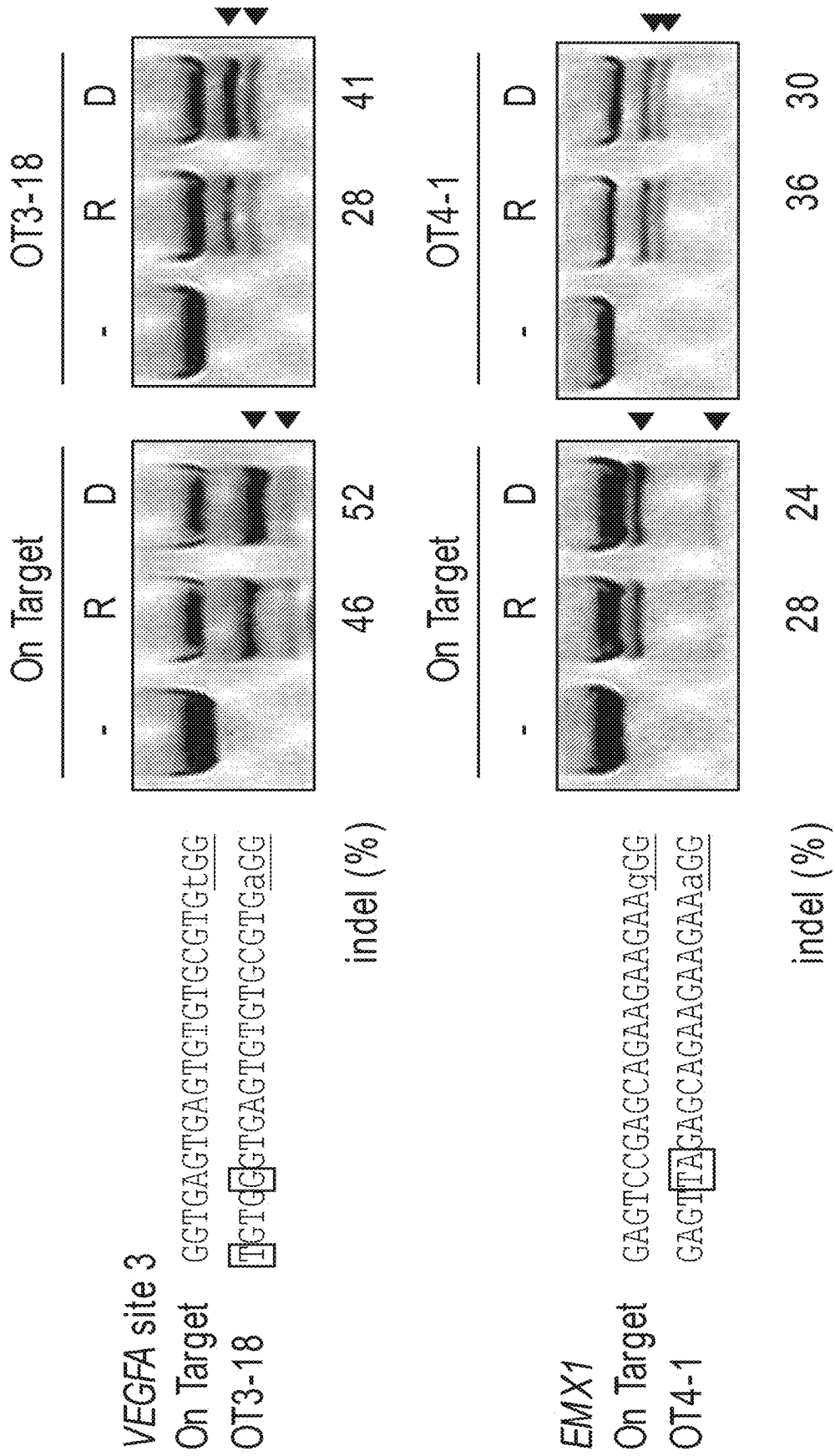


FIG. 14B

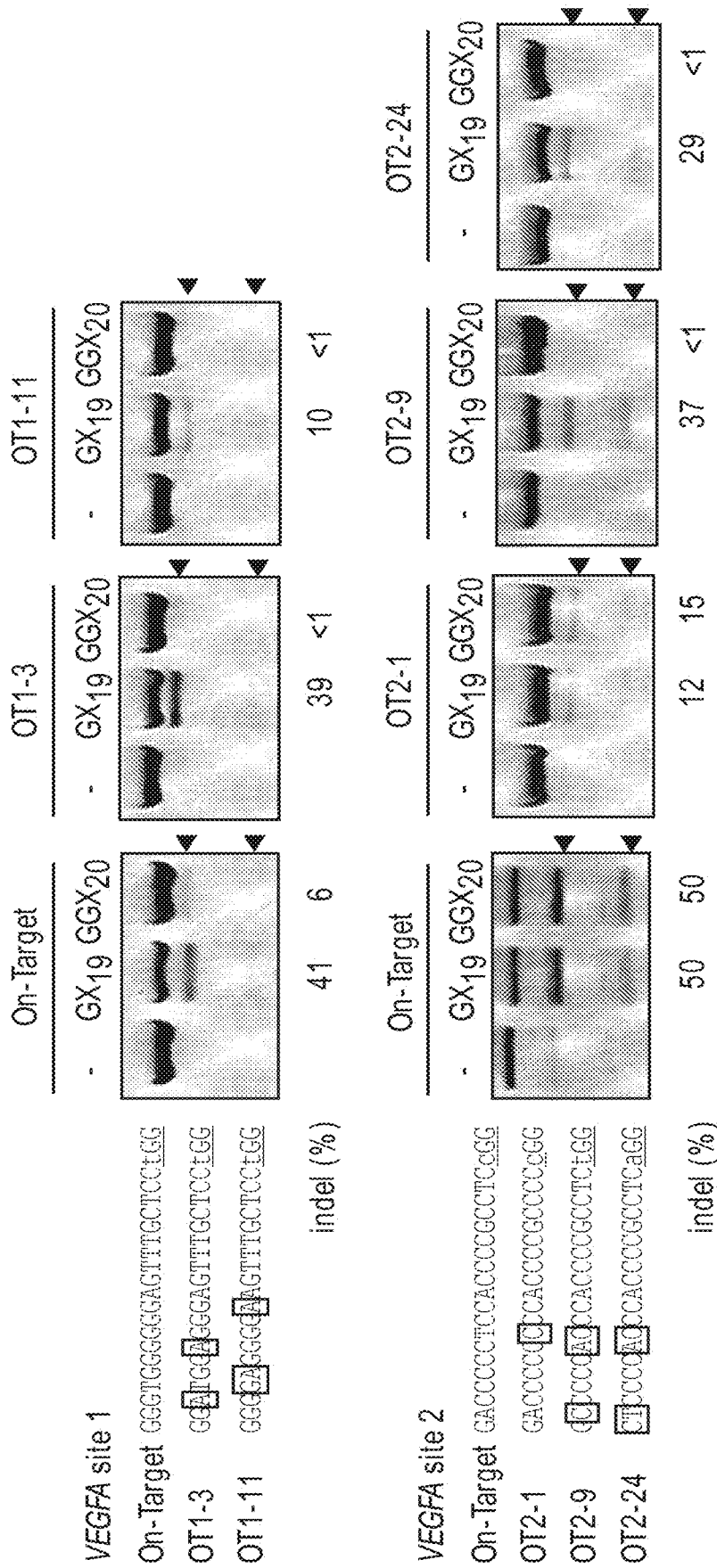


FIG. 15A

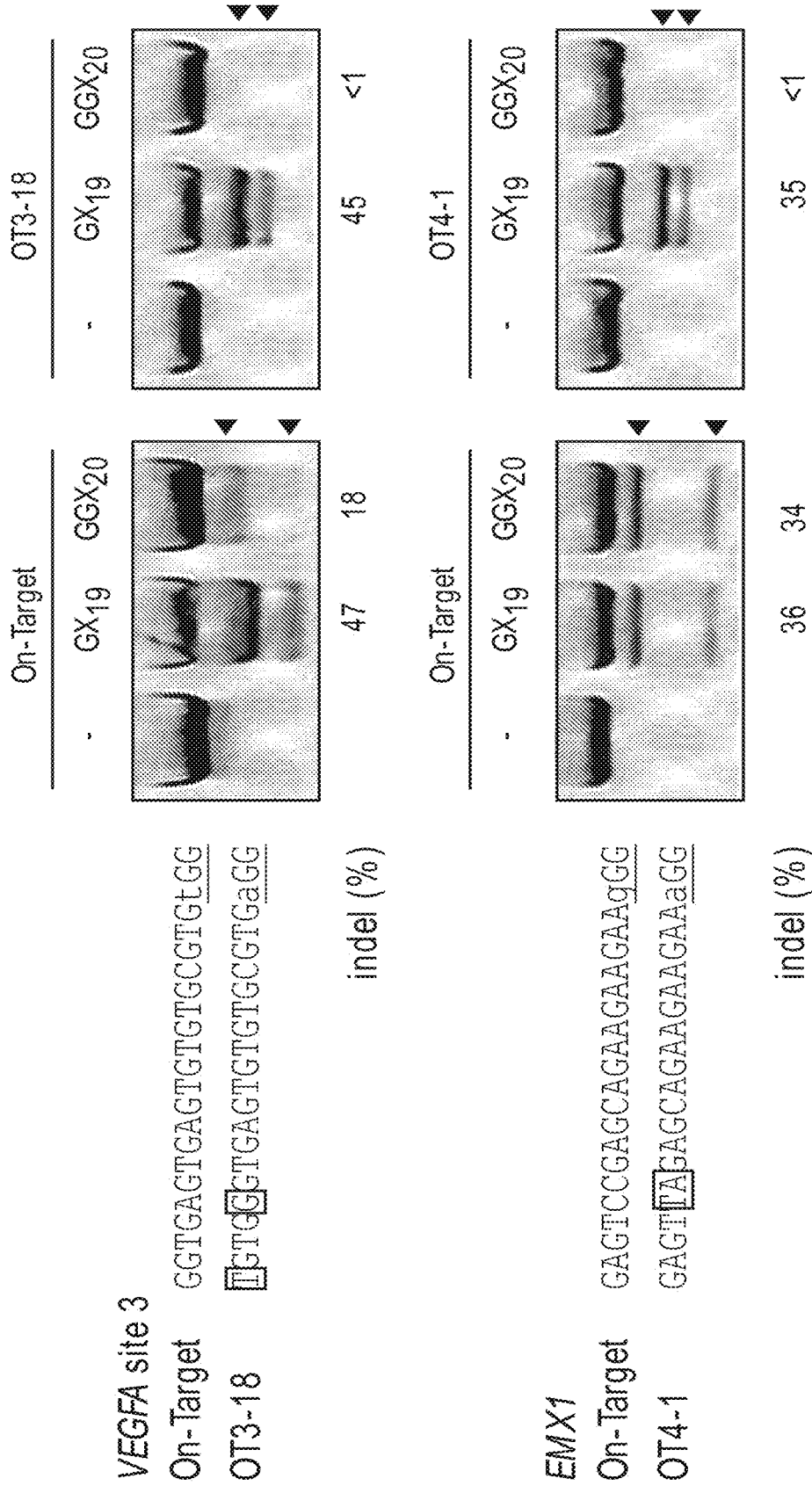


FIG. 15B

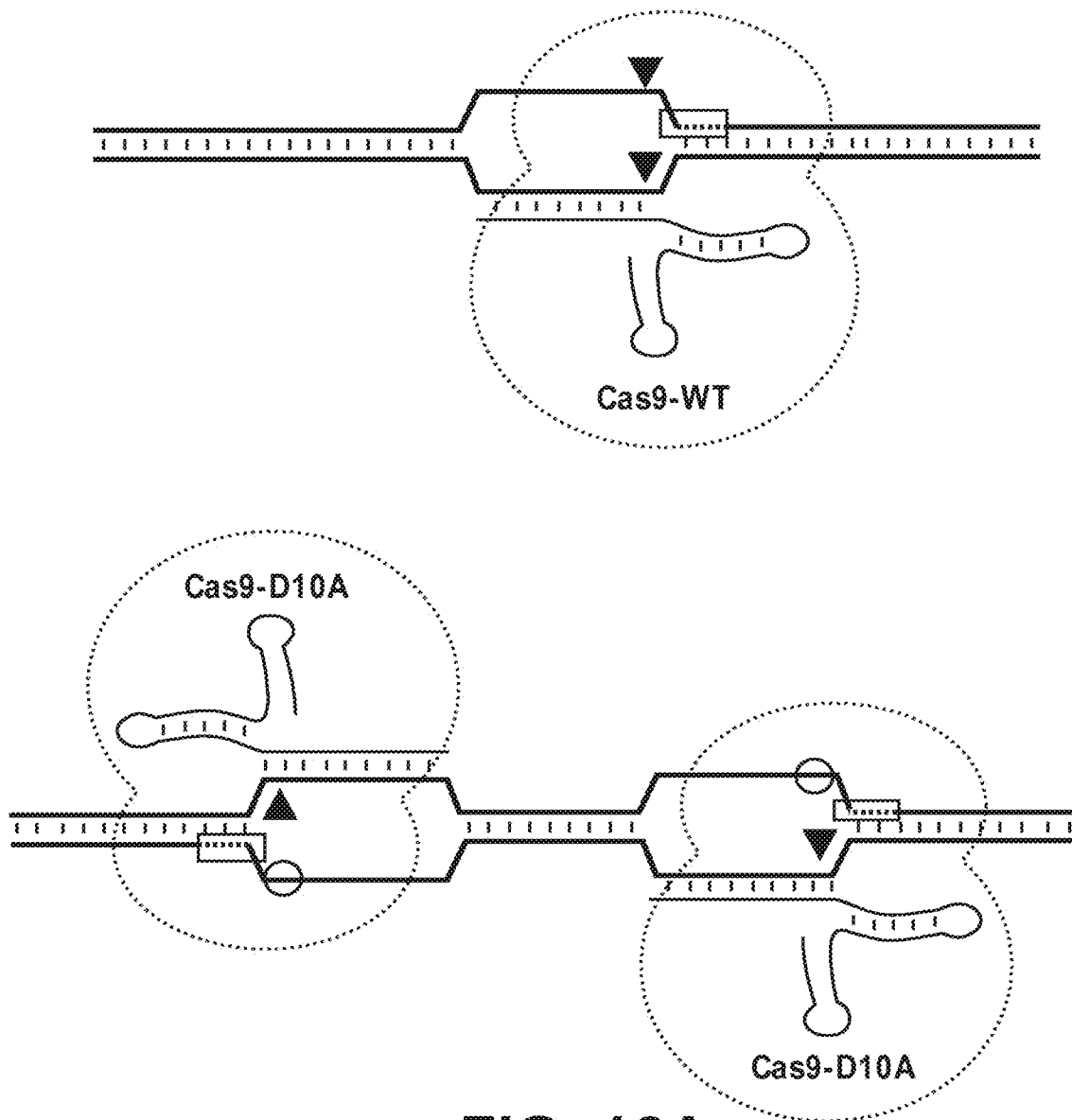


FIG. 16A

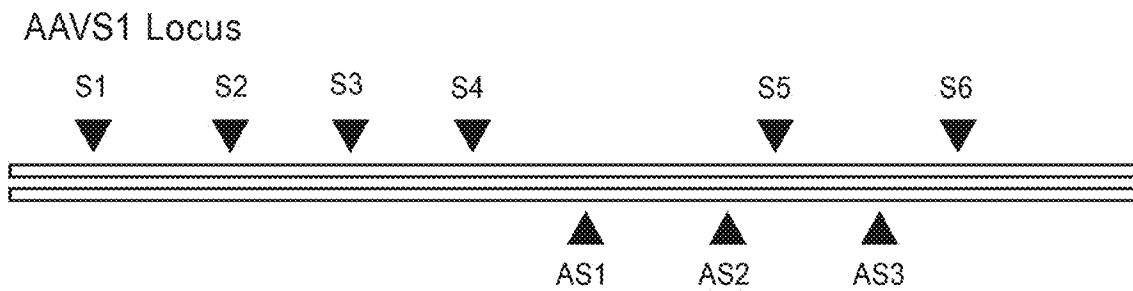


FIG. 16B

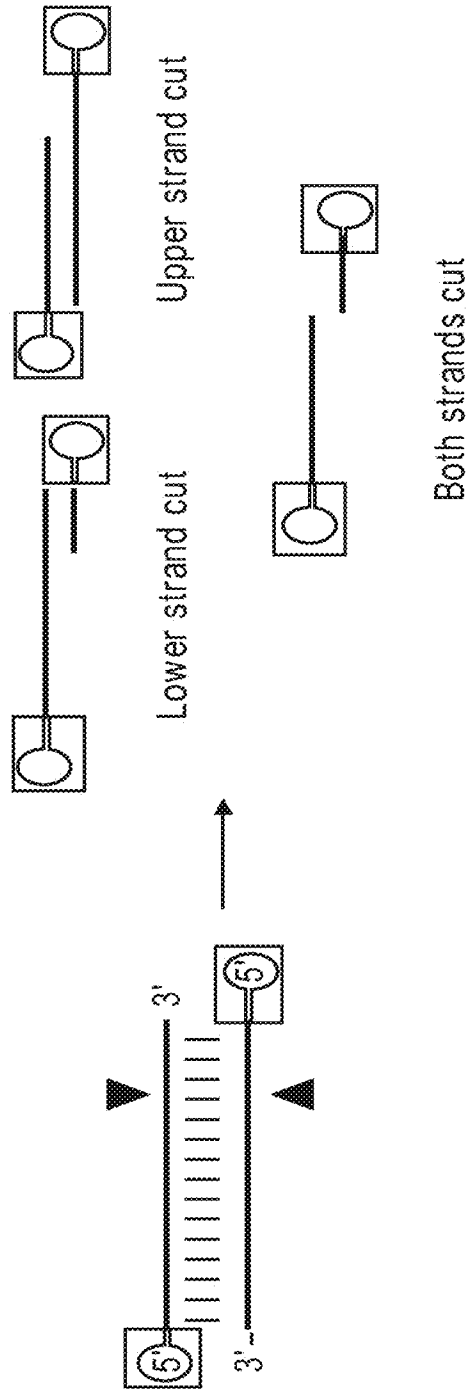


FIG. 16C

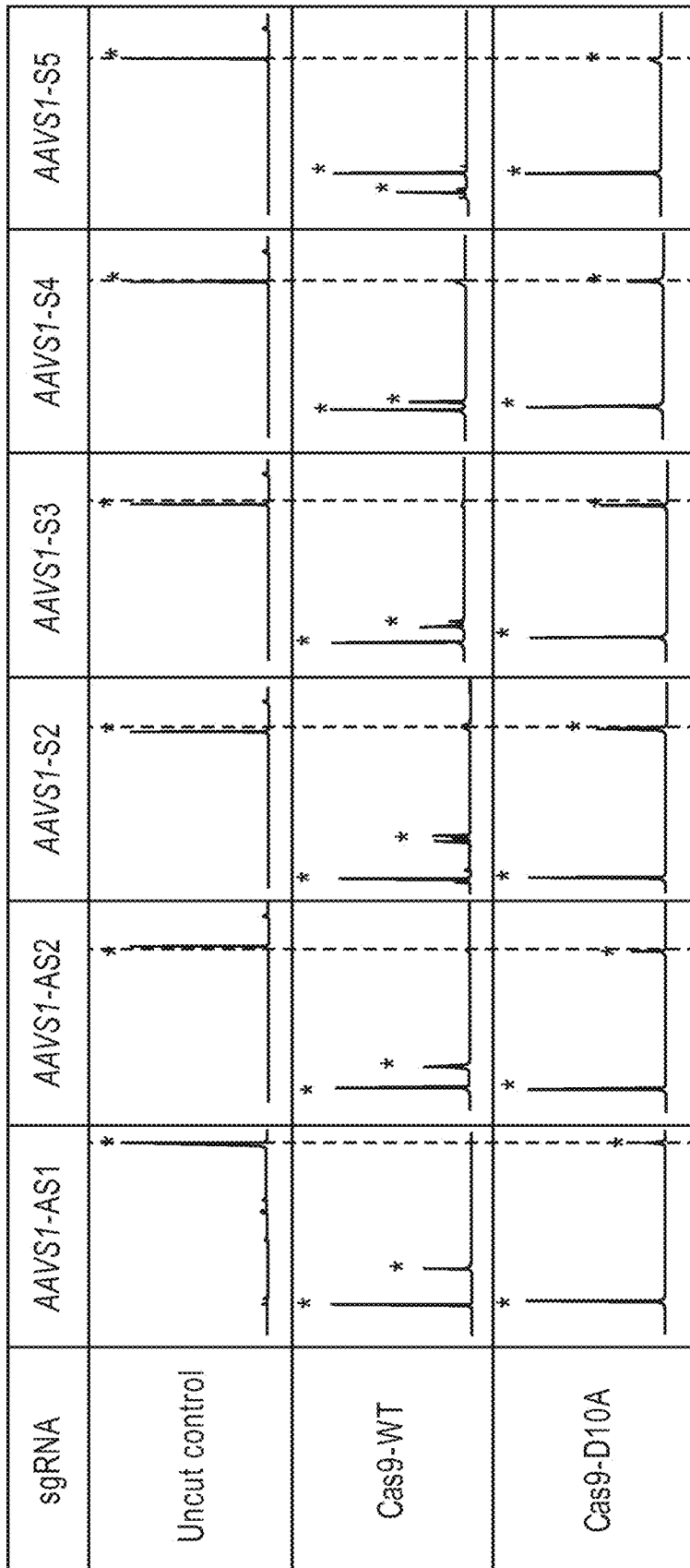


FIG. 16D

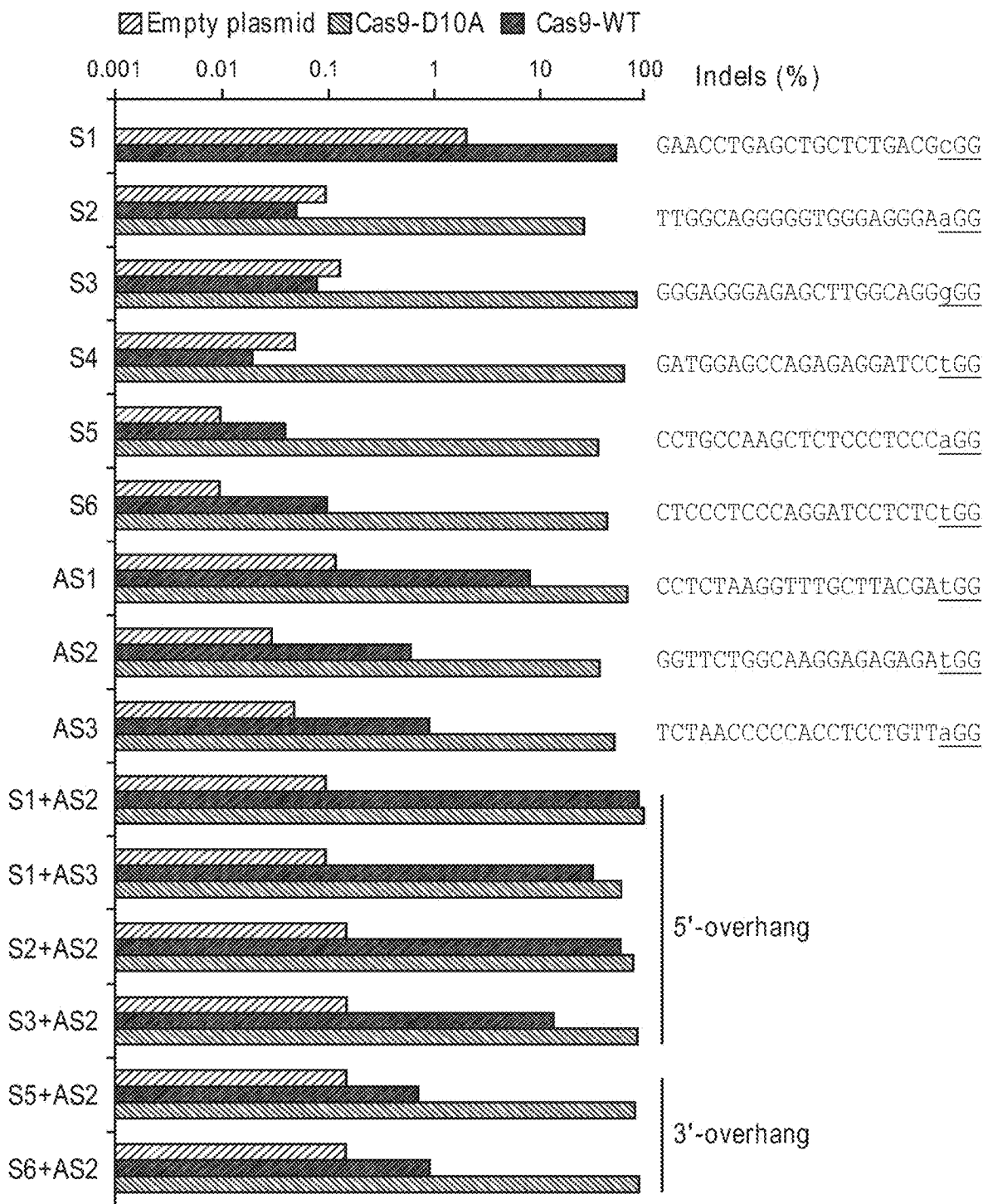


FIG. 17A

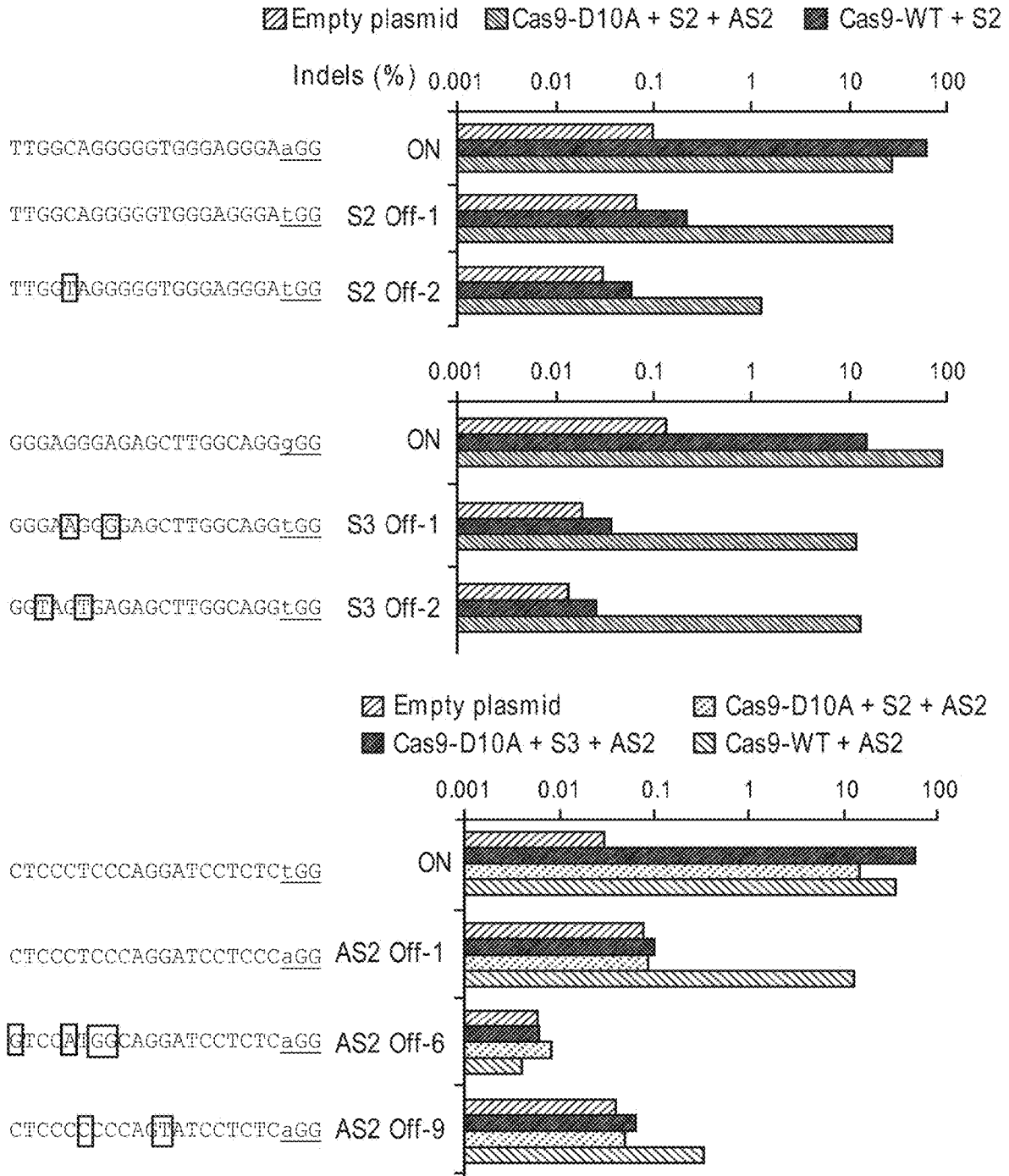


FIG. 17B

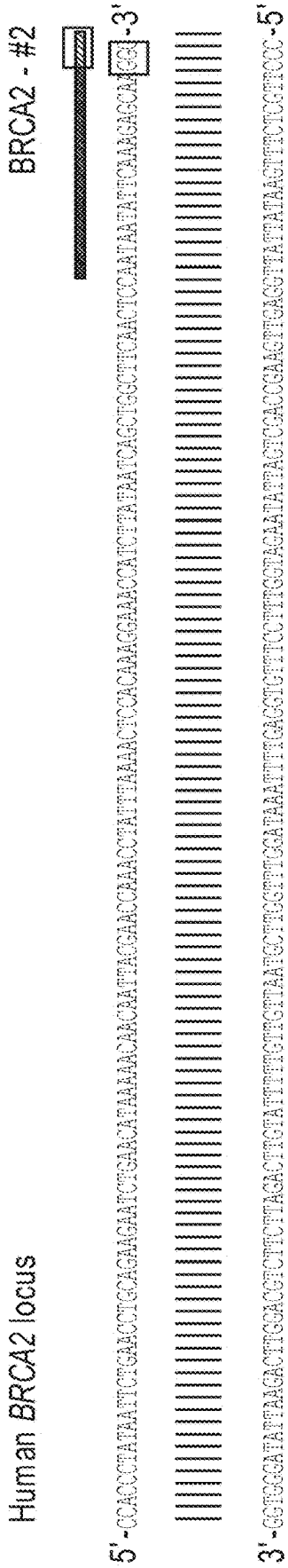


FIG. 18C

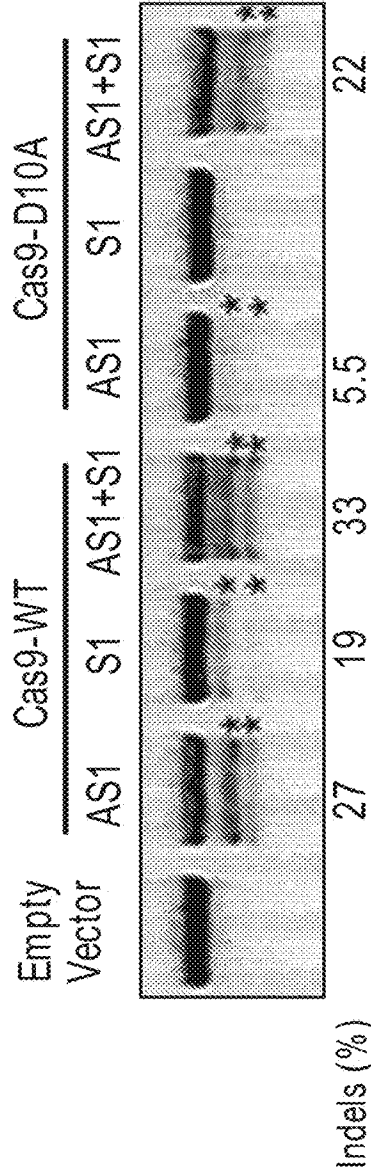


FIG. 18D

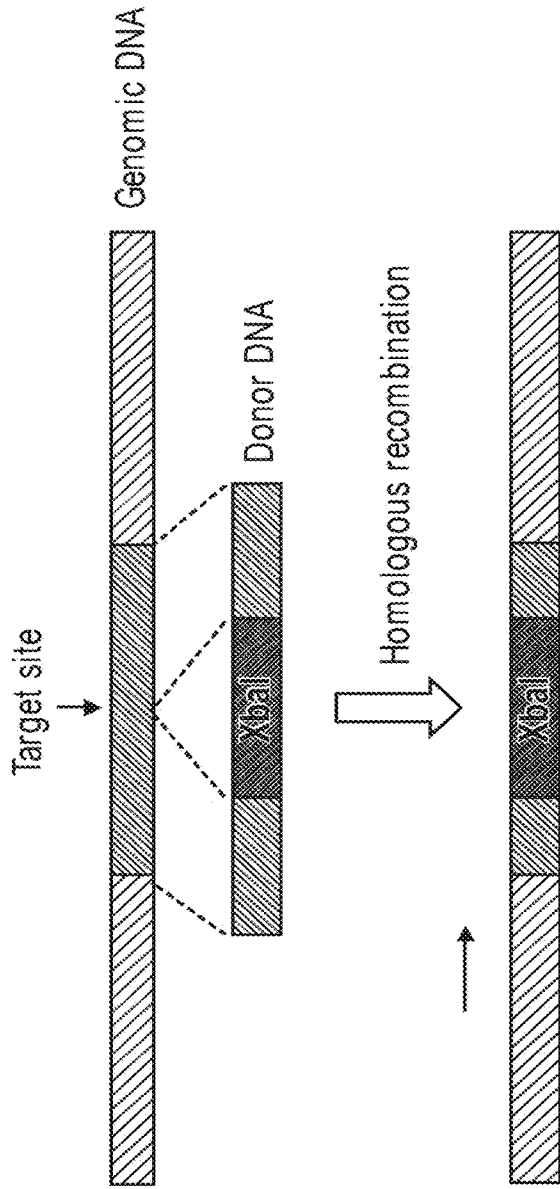


FIG. 19A

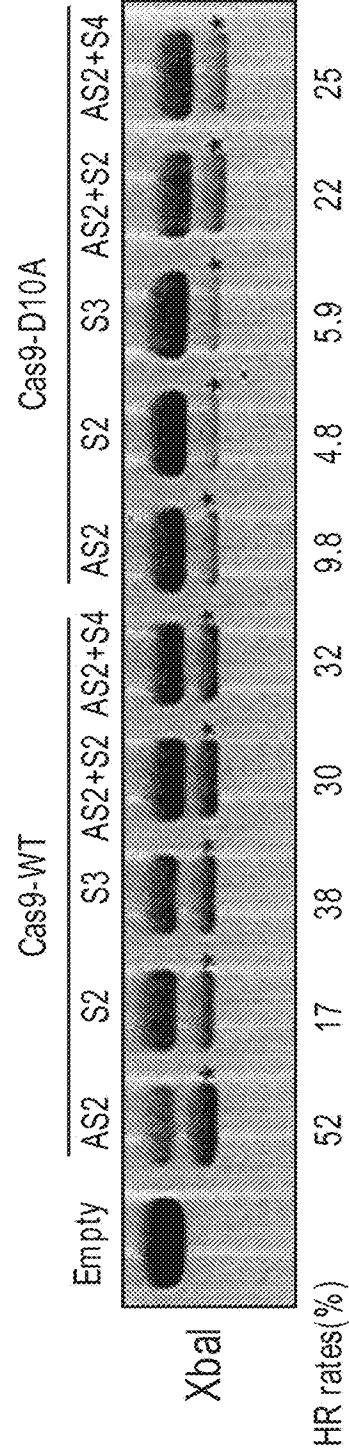


FIG. 19B

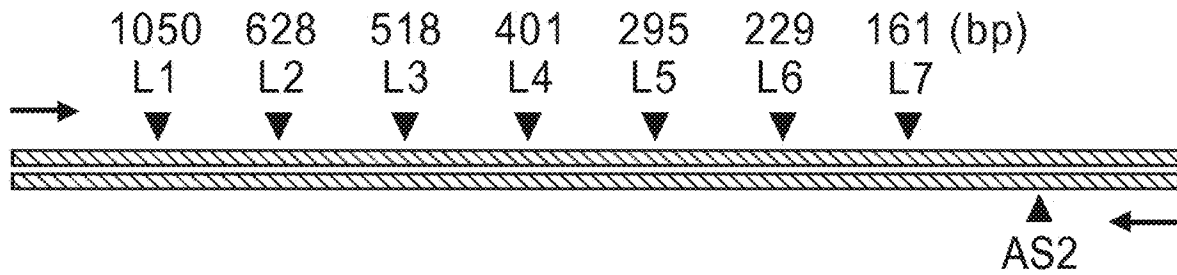


FIG. 20A

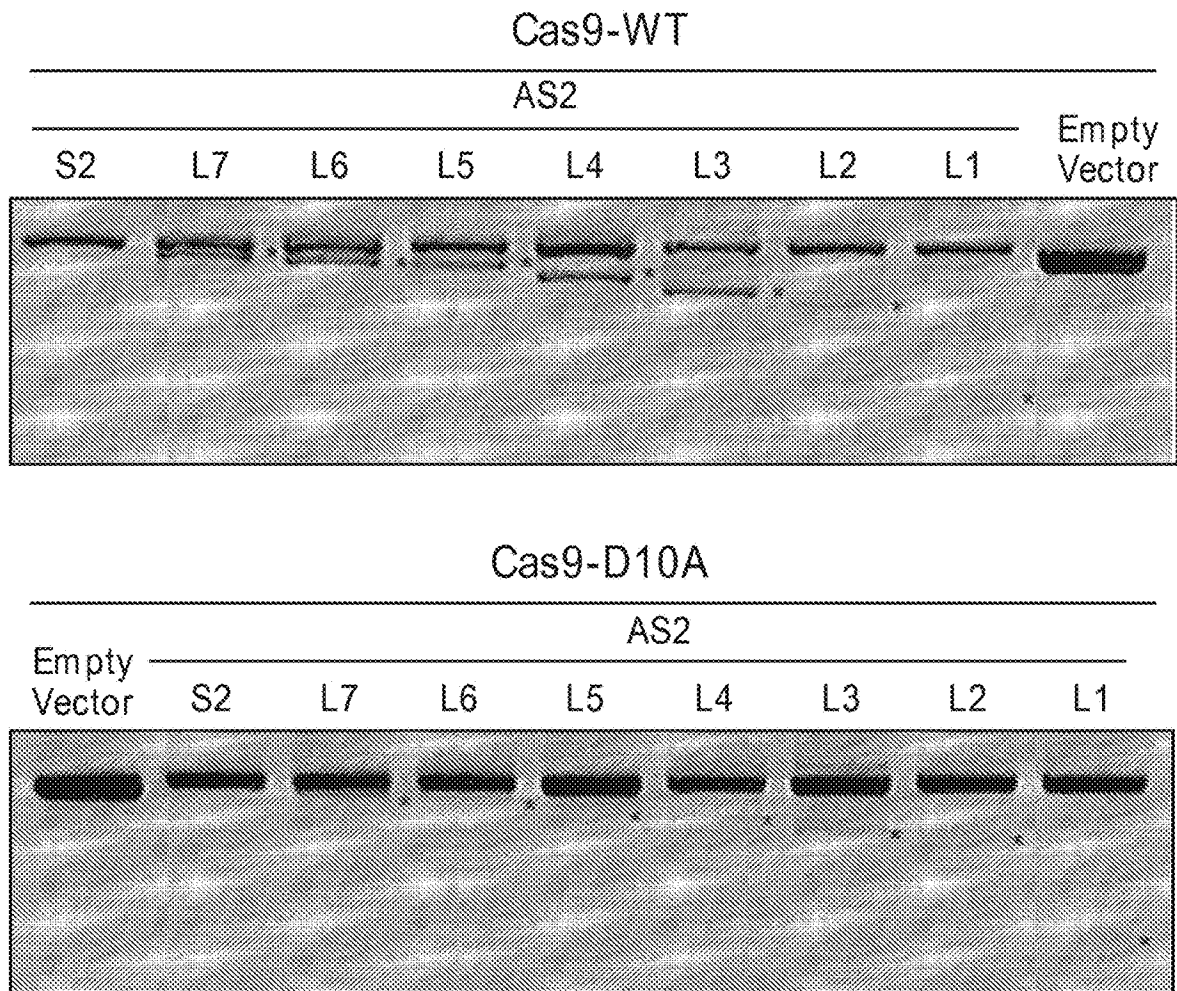


FIG. 20B

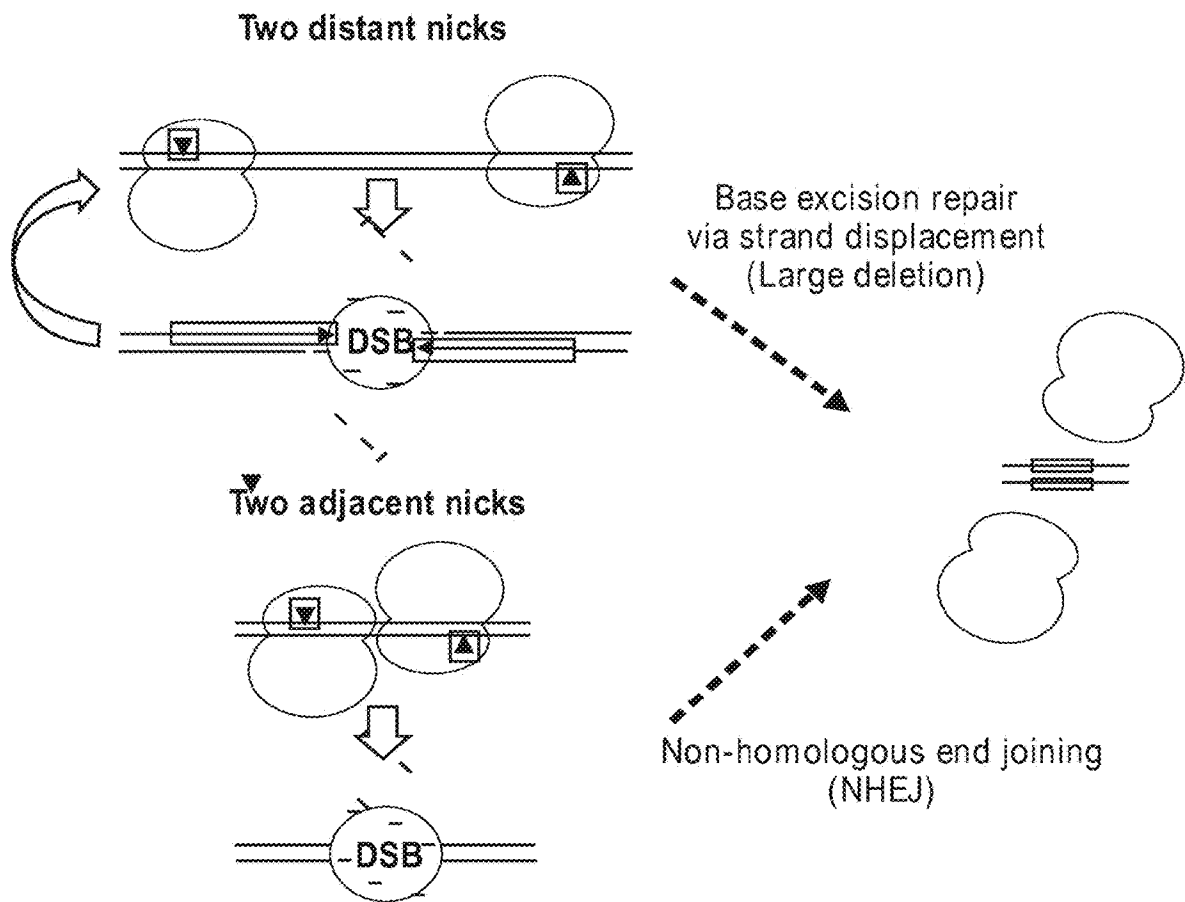


FIG. 20D

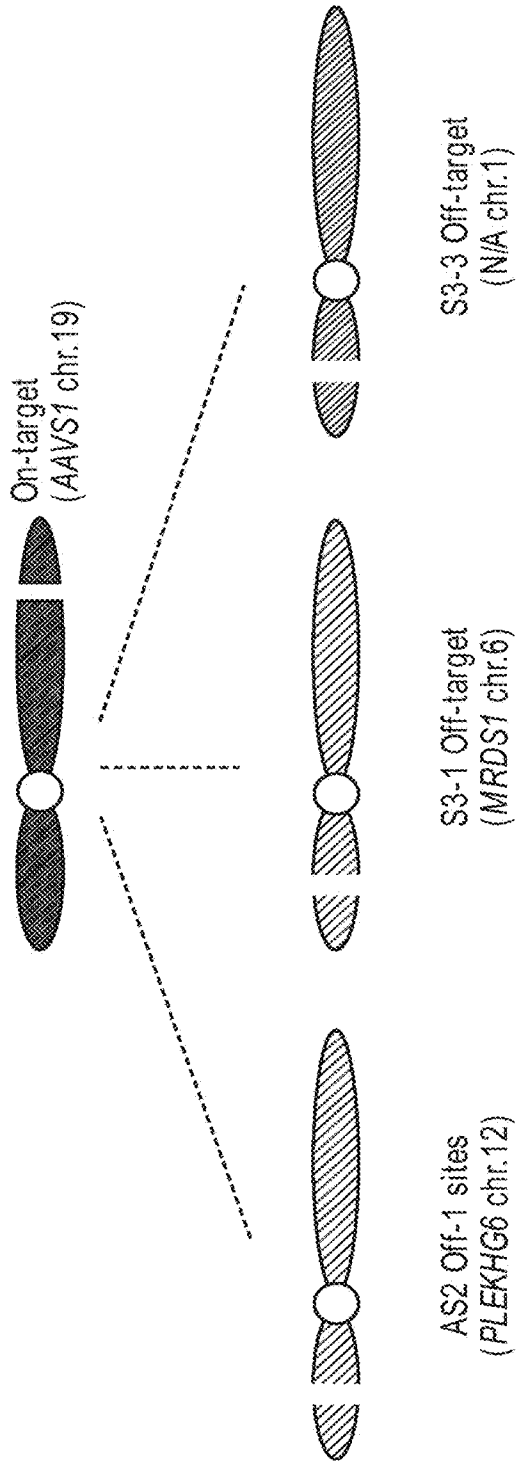


FIG. 21A

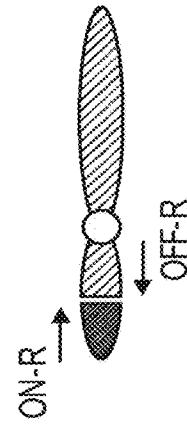
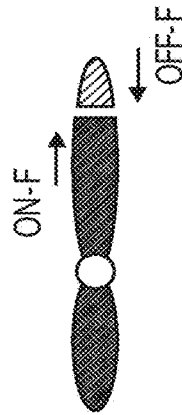


FIG. 21B

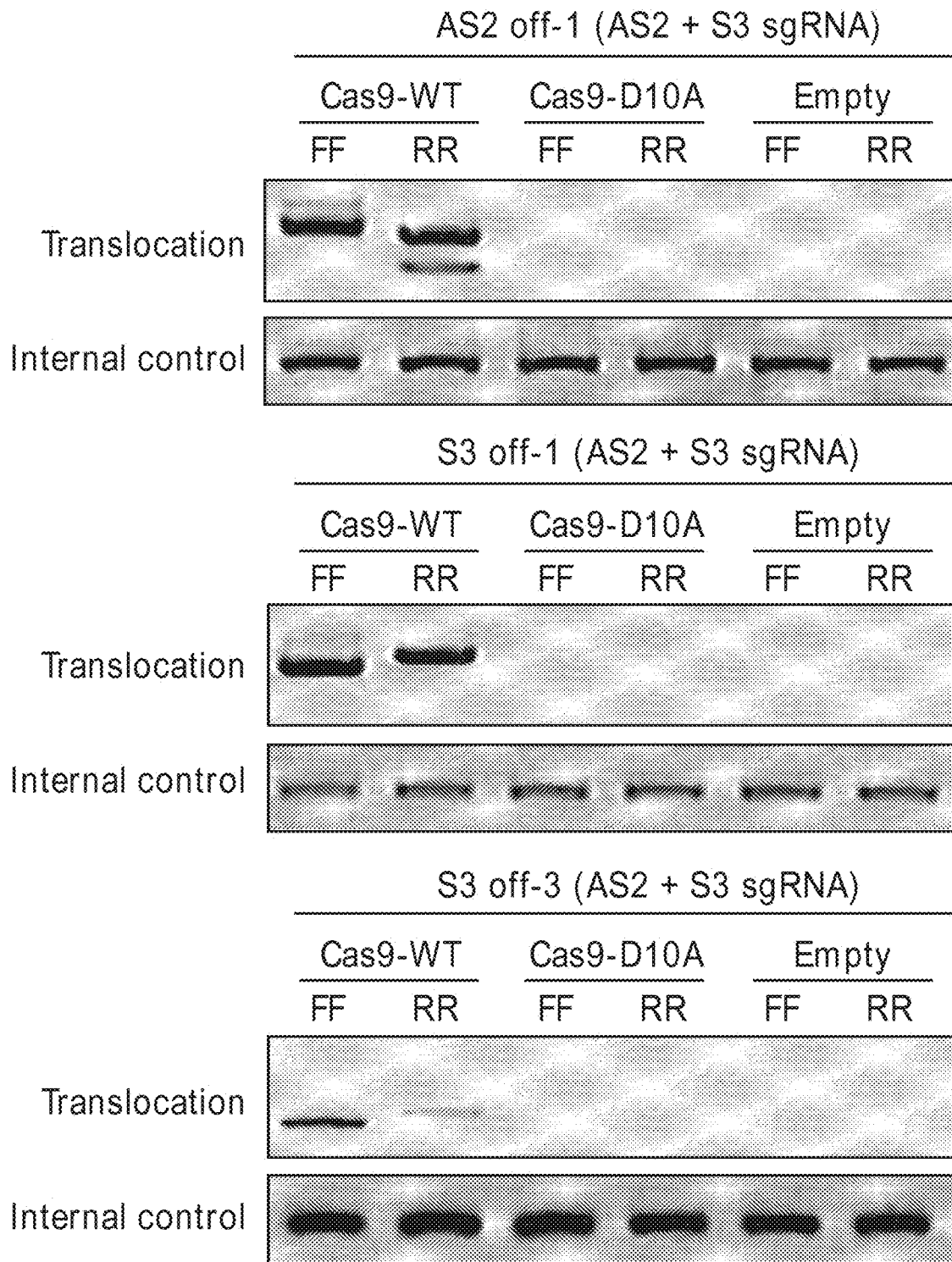


FIG. 21C

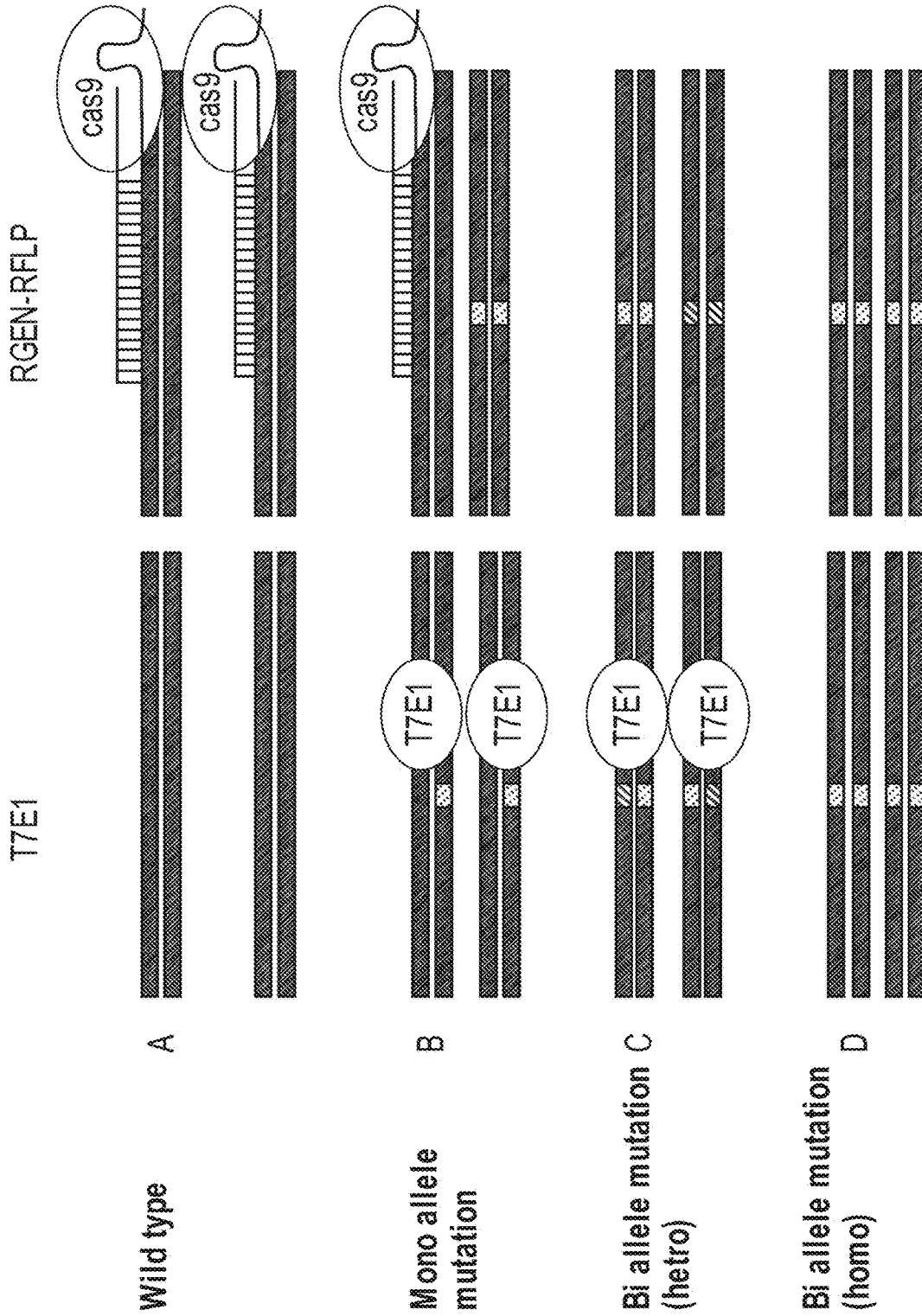


FIG. 22A

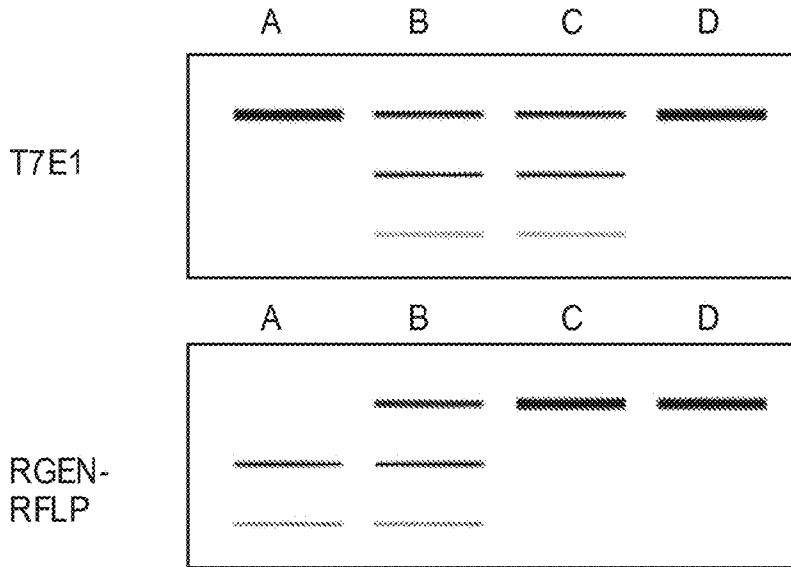


FIG. 22B

Plasmid target sequence

AATGACCACTACATCCT---	CAAGGG	WT	
AATGACCACTACATCCT	<u>T</u> ---	CAAGGG	I1
AATGACCACTACATCCT	<u>TT</u> ---	CAAGGG	I2
AATGACCACTACATCCT	<u>TTT</u>	CAAGGG	I3
AATGACCACTACATCCT----	AAGGG	D1	
AATGACCACTACATCCT-----	AGGG	D2	
AATGACCACTACATCCT-----	GGG	D3	

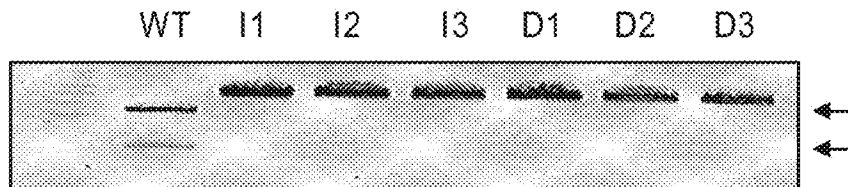


FIG. 23

#1 (+/-)

TATGTGC	AATGACCACTACATCCT	---	CAA	GGGCAGCAATCGGAG	WT
TATGTGC	AATGACCACTACATCCT	CCT	CAA	GGGCAGCAATCGGAG	+3

#2 (+/-)

TATGTGC	AATGACCACTACATCCT	CAAGGGCAGCAATCGGAG	WT
TATGTGC	AATGACCACTACATC	-----AATCGGAG	-12

#5 (+/-)

TATGTGC	AATGACCACTACATCCT	CAAGGGCAGCAATCGGAG	WT
TATGTGC	AATGACCACTACAT	-----CAGCAATCGGAG	-9

#6 (+/-)

TATGTGC	AATGACCACTACATCCT	CAAGGGCAGCAATCGGAG	WT
TATGTGC	AATGACCACTACATCC	-----AGCAATCGGAG	-8

#12 (-/-)

				-----CAGCAATCGG	-36
TATGTGC	AATGACCACTACATCCT	-----	CAA	GGGCAGCAATCGG	+1
TATGTGC	AATGACCACTACATCCT	-----	CAA	GGGCAGCAATCGG	+1
TATGTGC	AATGACCACTACATCCT	/67bp/	CAA	GGGCAGCAATCGG	+67

#28 (-/-)

TATGTGC	AATGACCACTACATCCT	CAAGGGCAGCAATCGG	+1
TATGTGC	AATGACCACTACAT	-----GGCAGCAATCGG	-7, +1
TATGTGC	AATGACCACTACAT	-----	-94

FIG. 24A

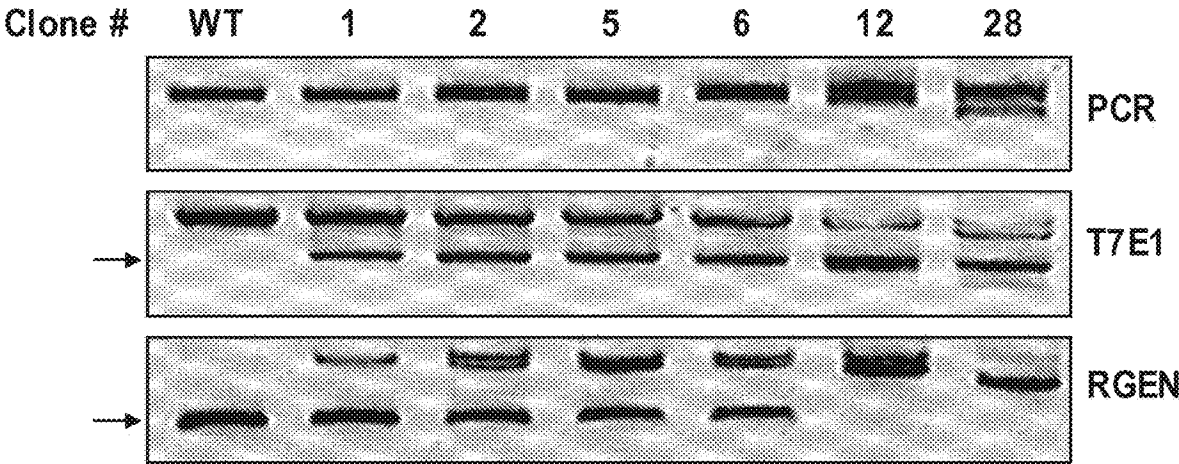


FIG. 24B

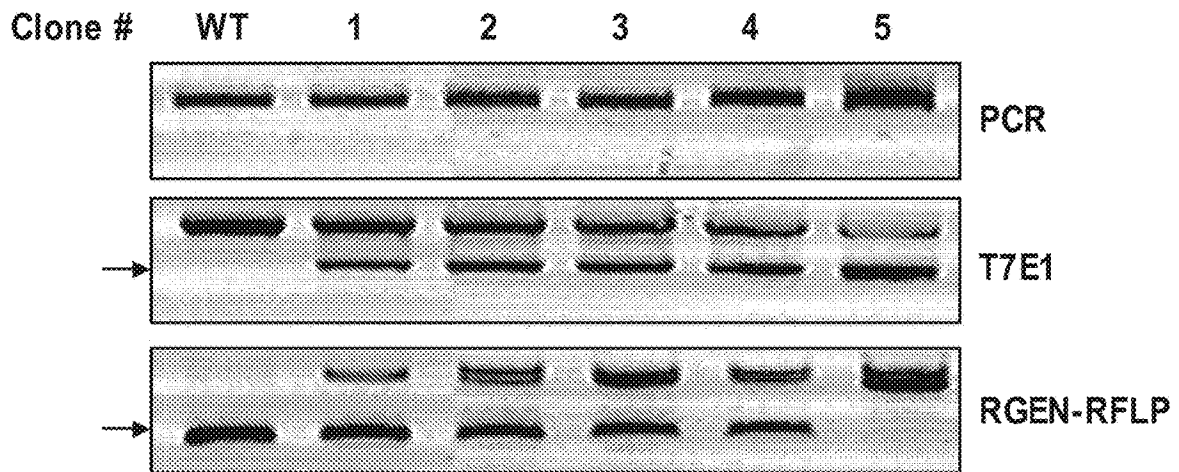


FIG. 25A

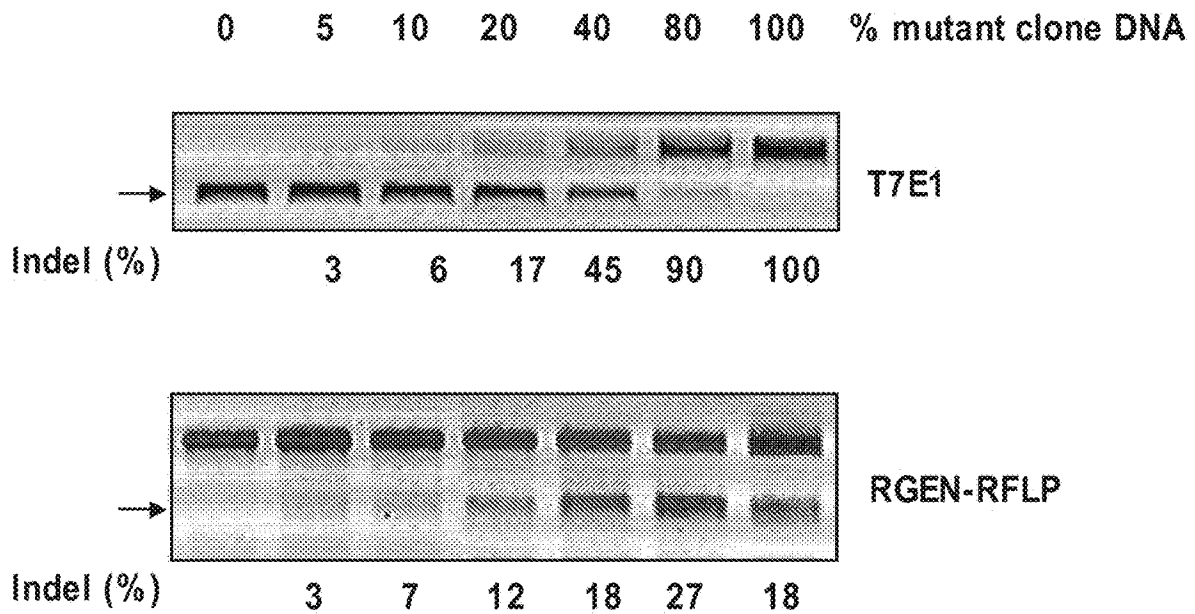


FIG. 25B

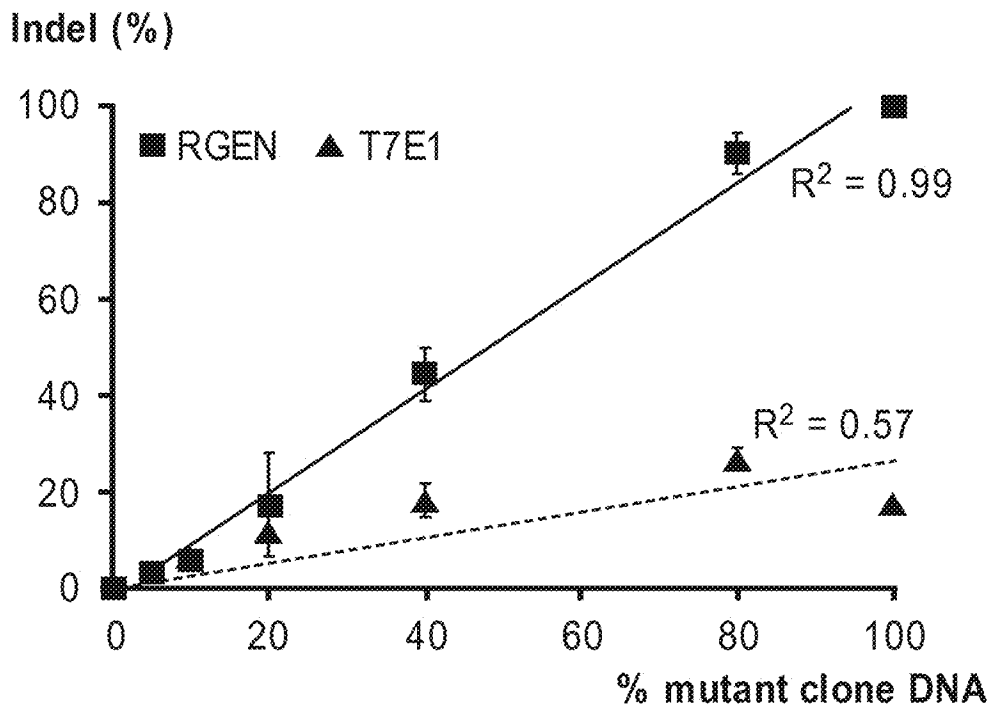
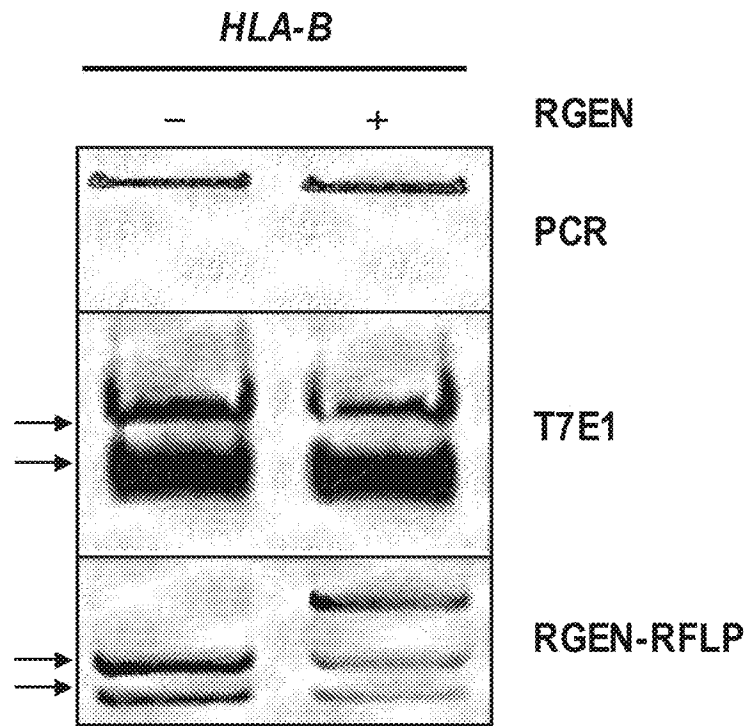


FIG. 25C

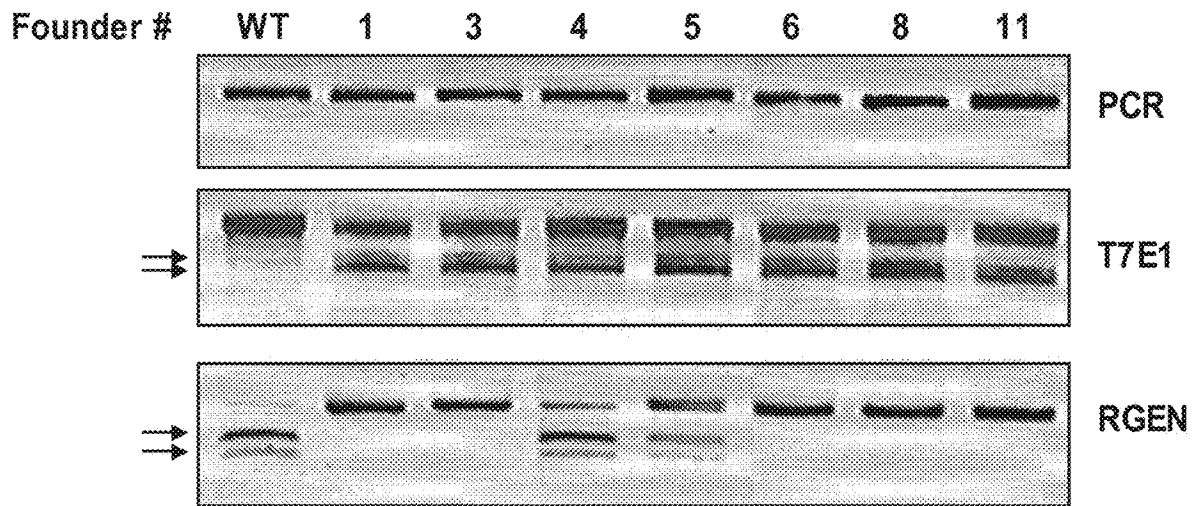


FIG. 26B

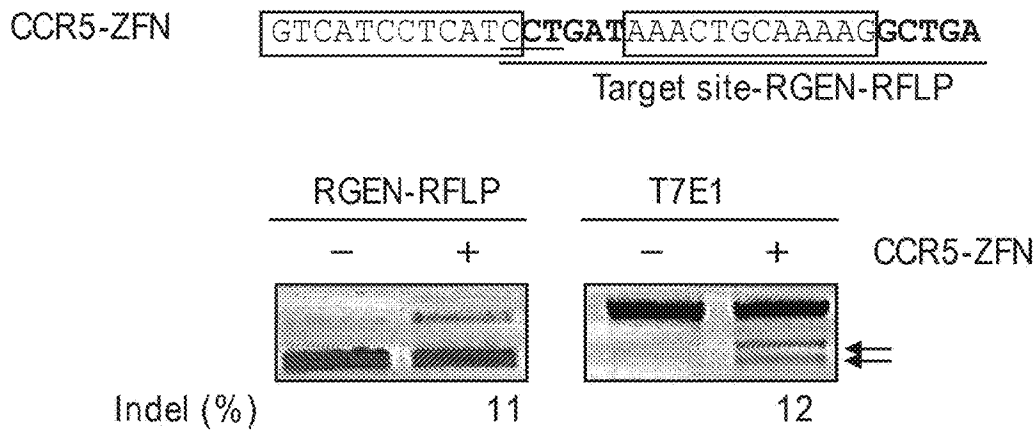


FIG. 27

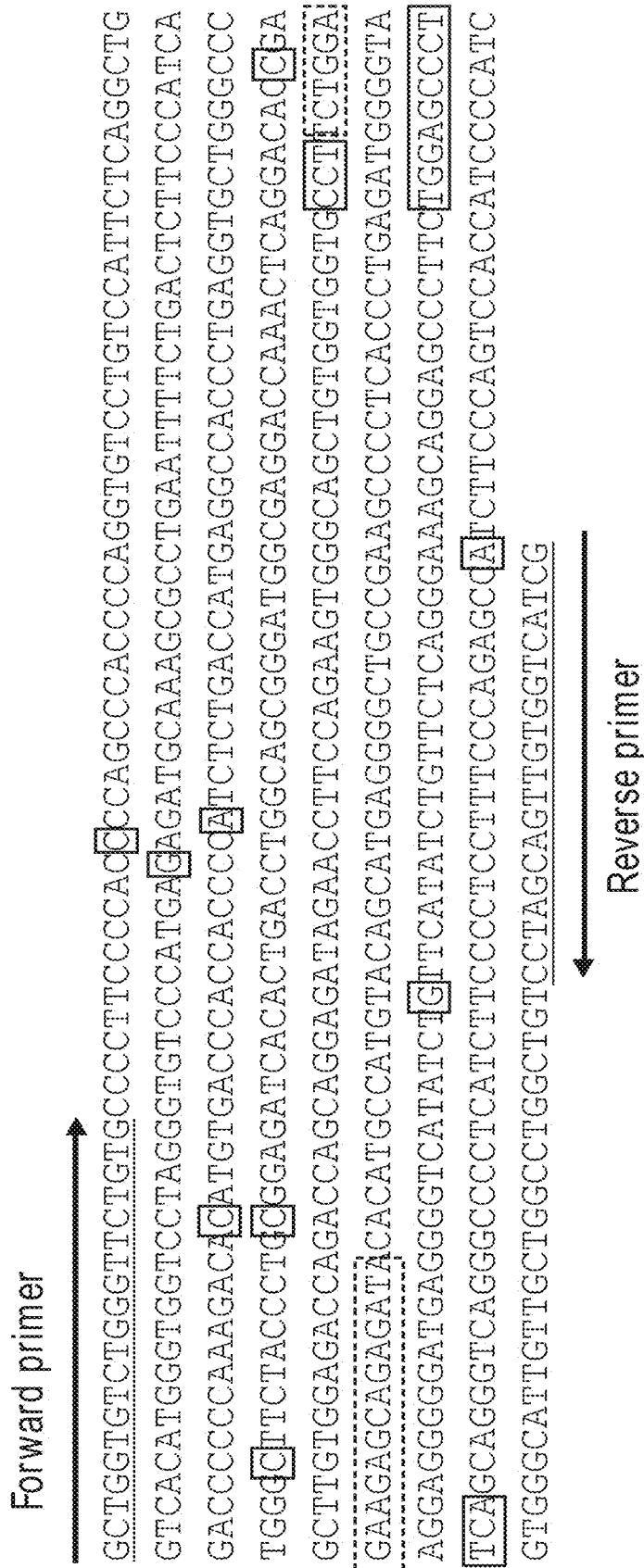
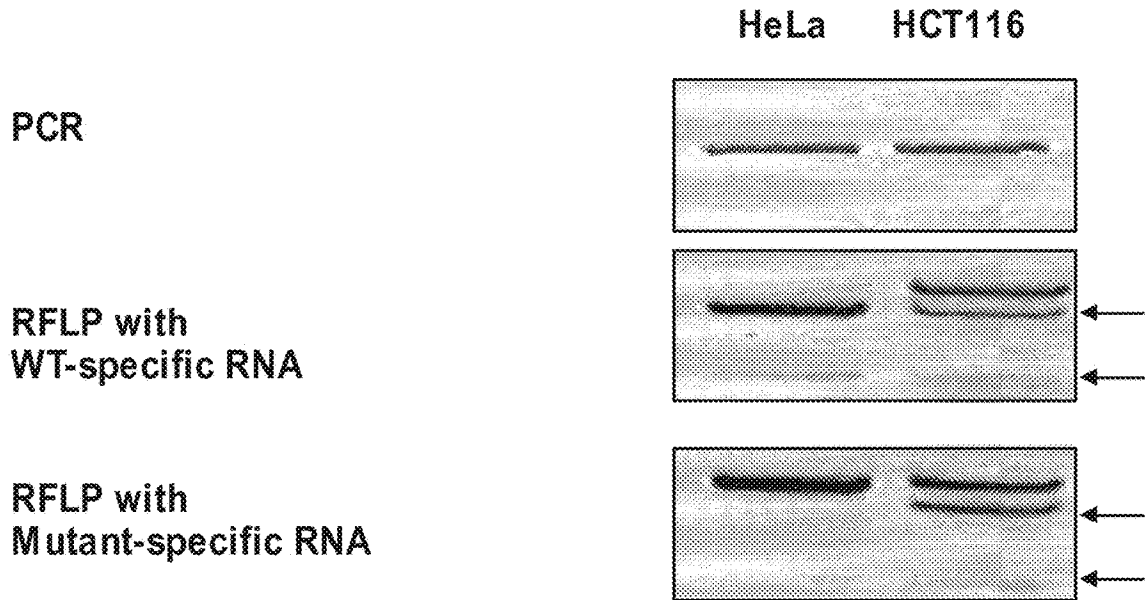


FIG. 28



HeLa

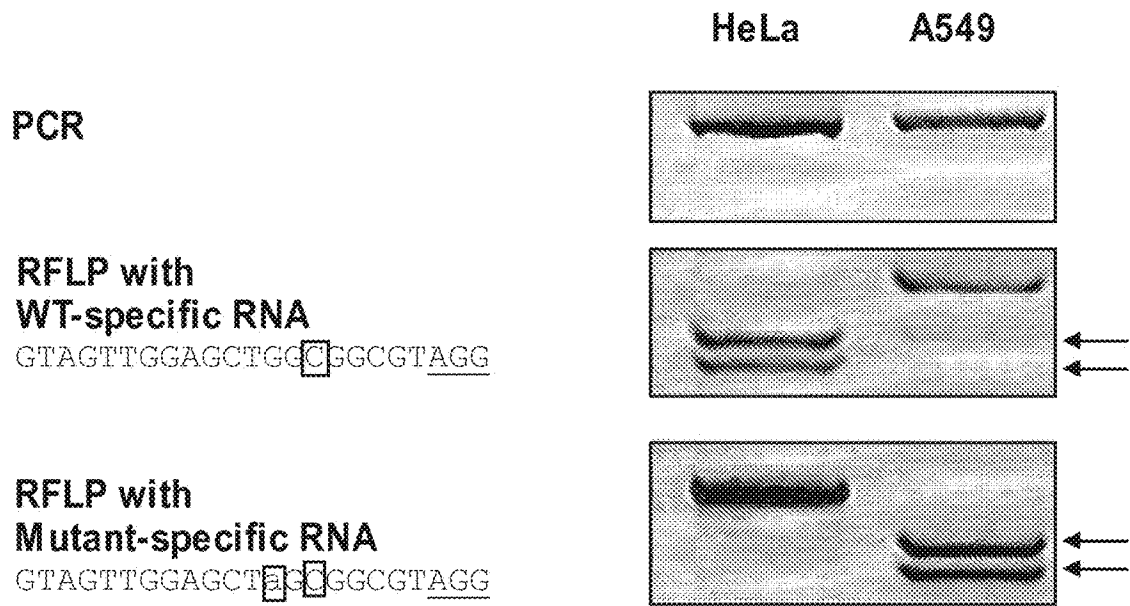
ACTACCACAGCTCCTTCTCTGAGTGG wild-type

HCT116

ACTACCACAGCTCCTTCTCTGAGTGG wild-type

ACTACCACAGCTCCT---CTGAGTGG c.133-135 del TCT

FIG. 29A



HeLa

GTAGTTGGAGCTGGTGGCGTAGG wild-type

A549

GTAGTTGGAGCTGGTGGCGTAGG c.34G>A

FIG. 29B

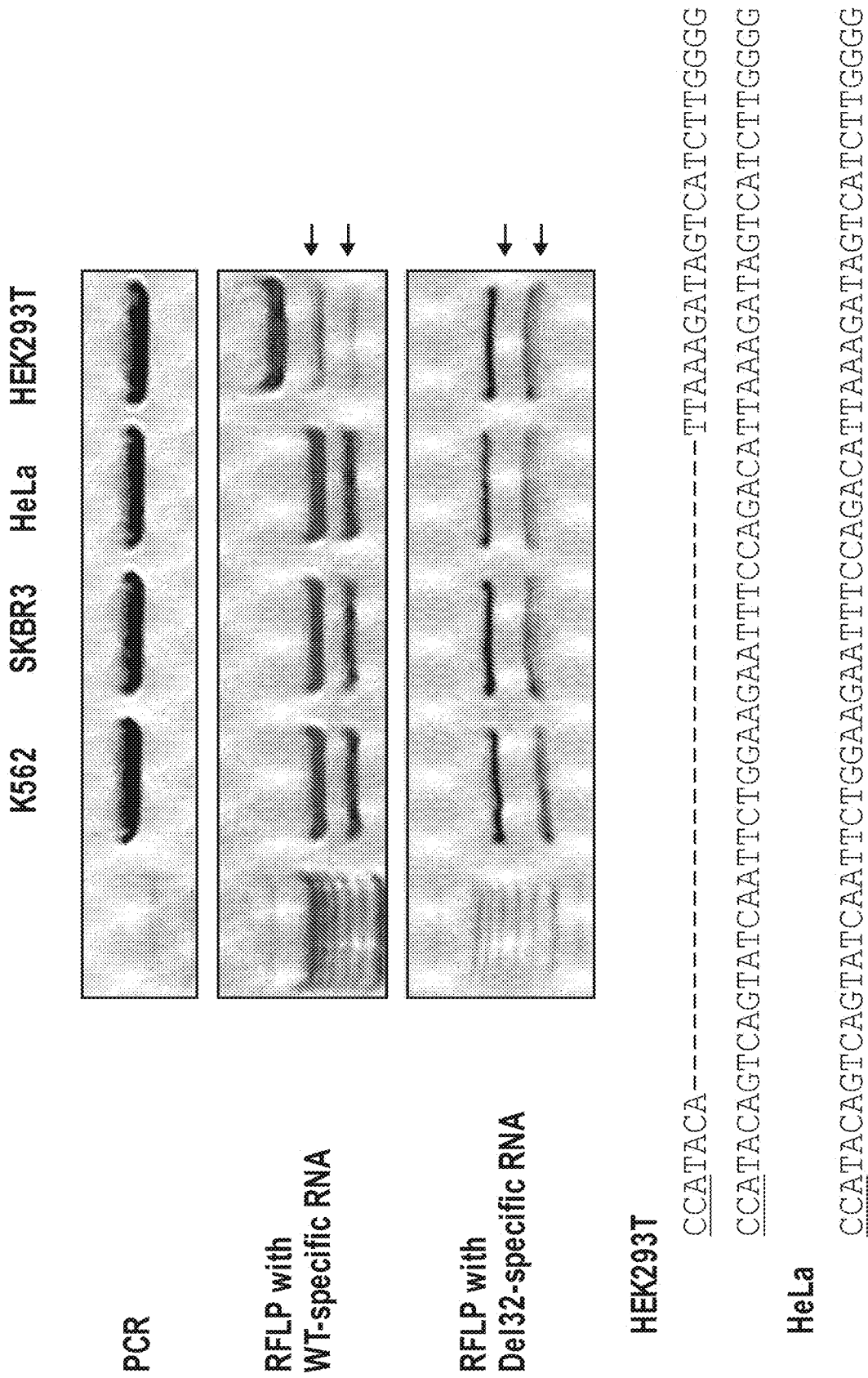


FIG. 30A

CCR5 WT
 ccatacagtcagtcattctggaagaatttccagacattaaagatagtcattct
 CCR5 WT target site
 CCR5 Δ32
 ccataca-----ttaagatagtcattct
 CCR5 Δ32 on-target site
 CCR5 Δ32 off-target site

FIG. 30B

WT Δ32 Linearized plasmid

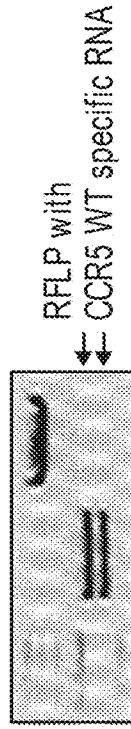
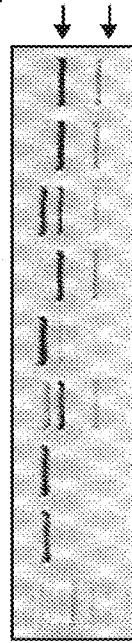


FIG. 30C

181 127 1/9
 on off on off on off on off
 Dilution (Cas9 0.1ug/crRNA 25ng / tracrRNA 60ng)
 Plasmid



On agatgactatctttaatgctcgg
 Off agatgactatctttaatgctcgg

FIG. 30D

KRAS

HeLa

GTAGTTGGAGCTGGTGGCGTAGG Wild-type

A549

GTAGTTGGAGCT^aGTGGCGTAGG c.34G>A

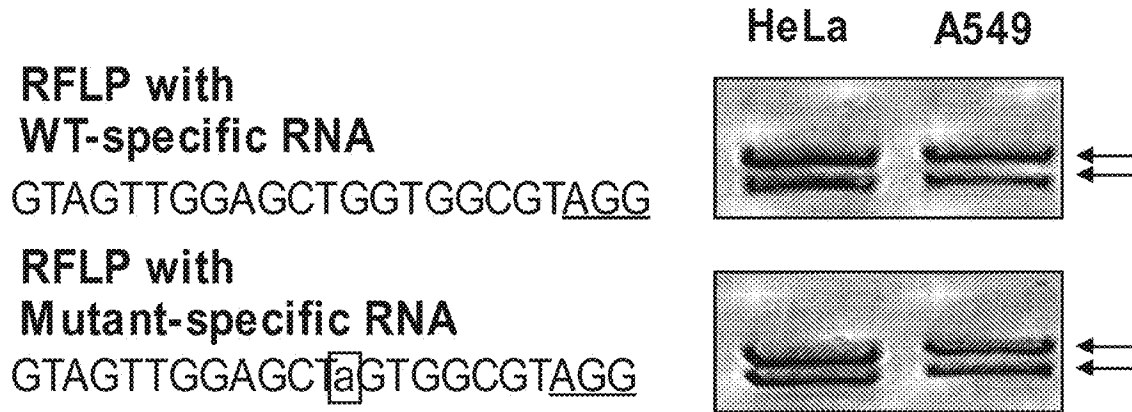


FIG. 31A

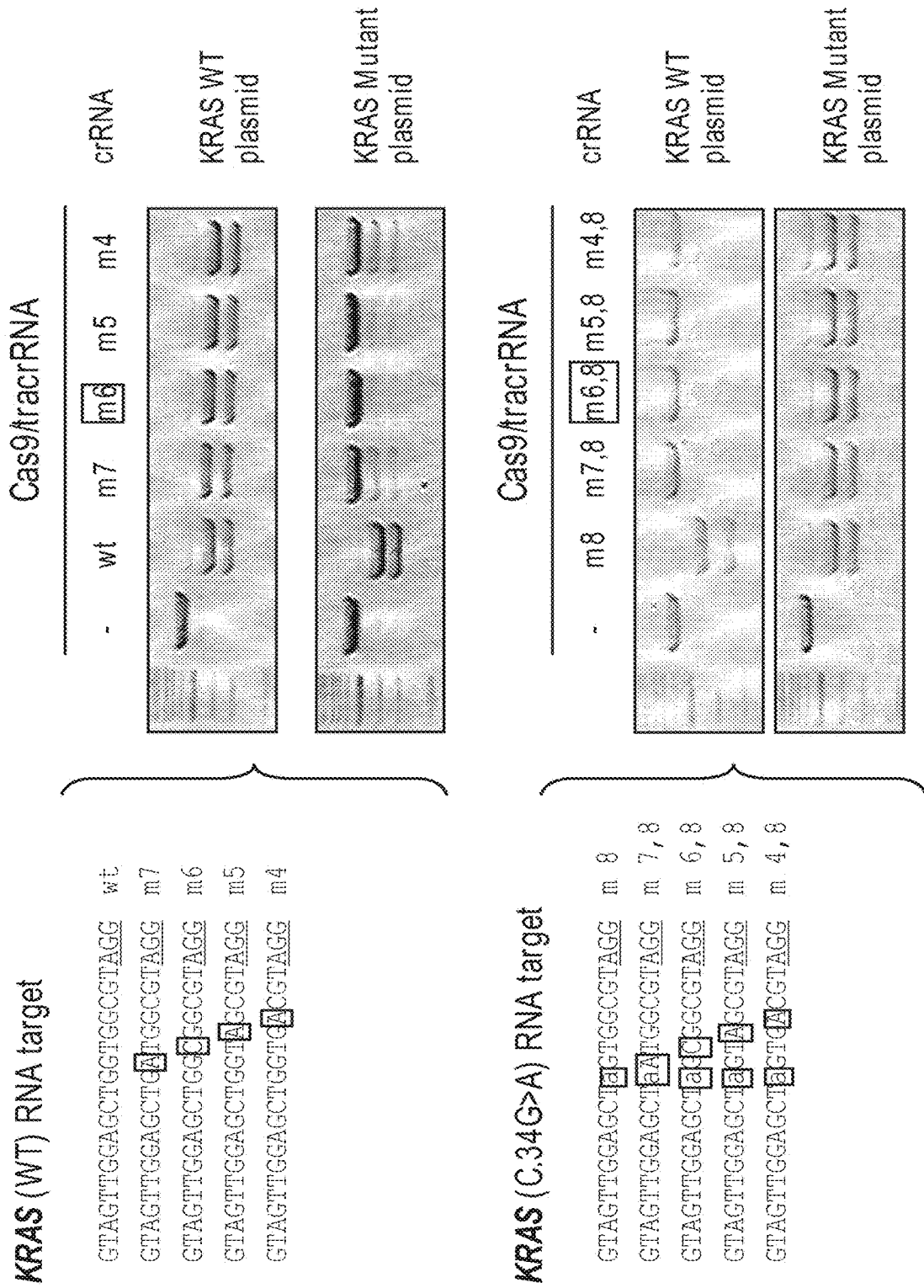


FIG. 31B

PIK3CA

HeLa

CAAATGAATGATGCACATCATGG Wild-type

HCT116

CAAATGAATGATGCACATCATGG Wild-type

CAAATGAATGATGCACAGTCATGG C.3140A>G

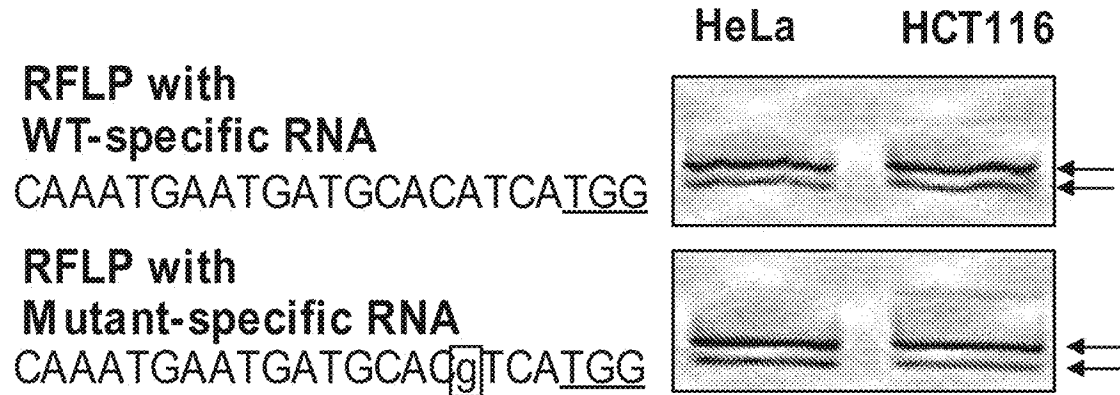


FIG. 32A

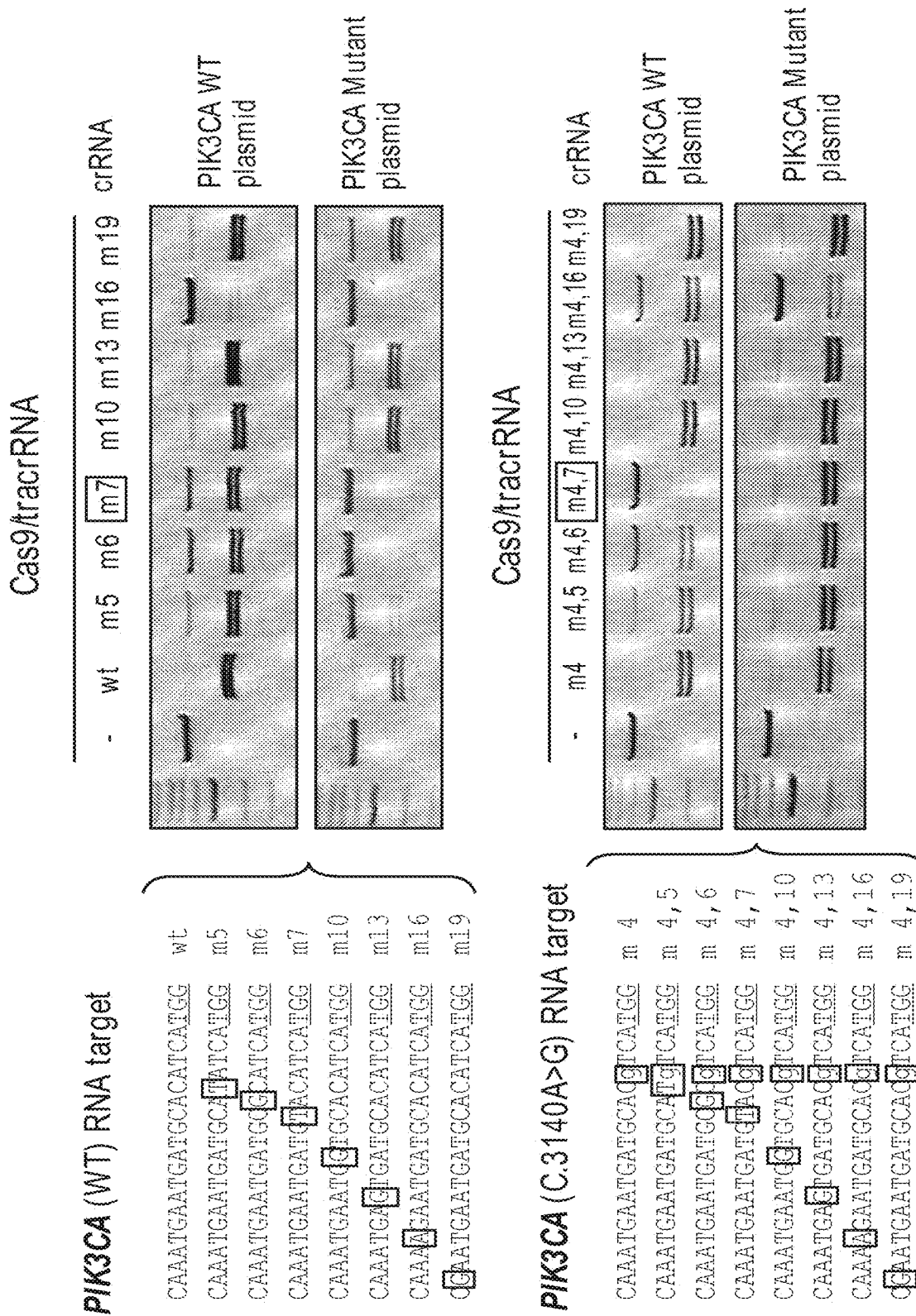


FIG. 32B

IDH1

HeLa

ATCATAGGTCGTCATGCTTATGG Wild-type

HT1080

ATCATAGGTCGTCATGCTTATGG Wild-typ

ATCATAGGTGTCATGCTTATGG c.394C>T

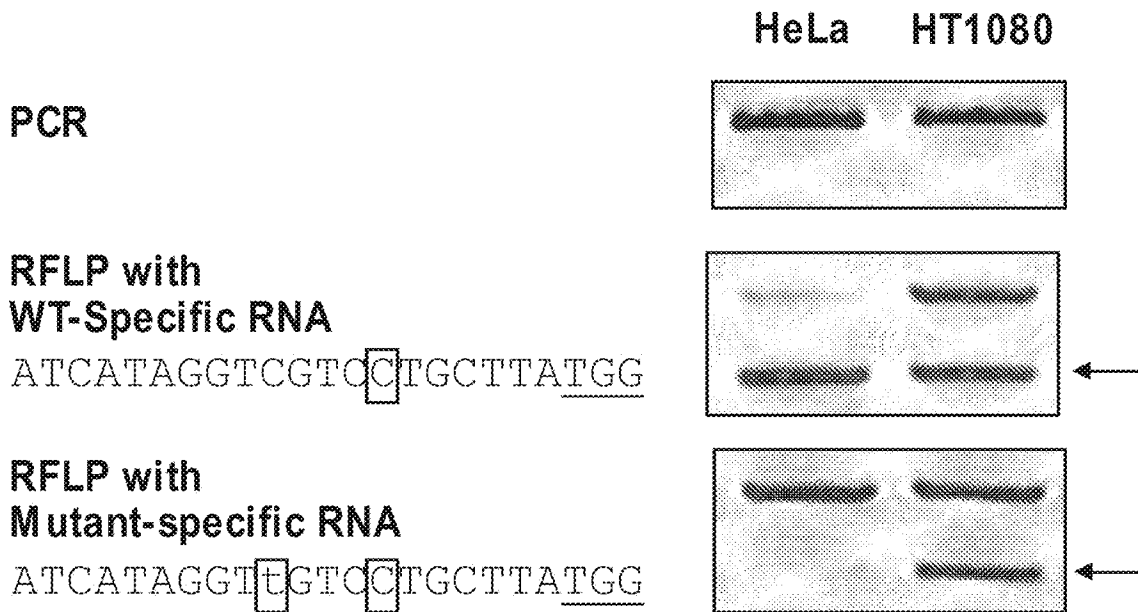


FIG. 33A

PIK3CA

HeLa

CAAATGAATGATGCACATCATGG Wild-type

HCT116

CAAATGAATGATGCACATCATGG Wild-type

CAAATGAATGATGCACgTCATGG C.3140A>G

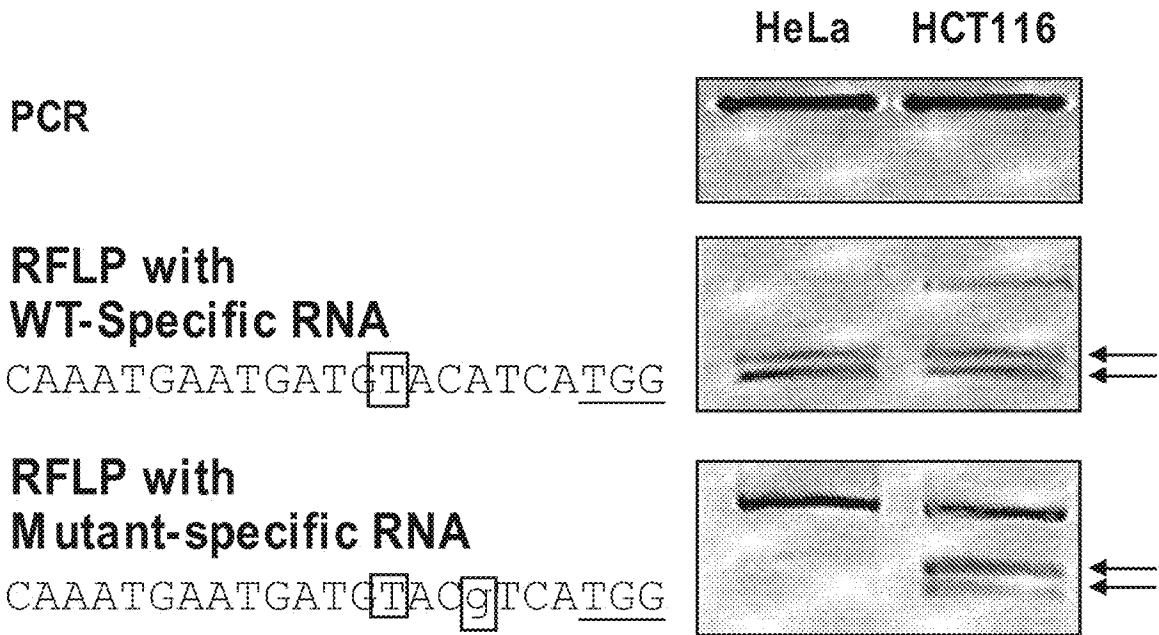


FIG. 33B

NRAS

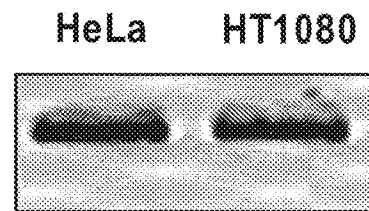
HeLa

CTGGACAAGAAGAGTACAGTGCC Wild-type

HT1080

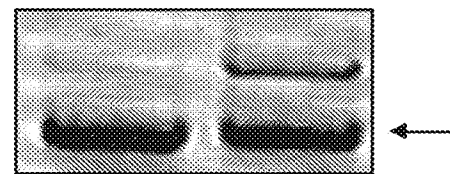
CTGGACAAGAAGAGTACAGTGCC Wild-type
CTGGAA**a**AAGAAGAGTACAGTGCC c.181C>A

PCR



**RFLP with
WT-Specific RNA**

CTGGACAAGAAGAGTACAGTGCC



**RFLP with
Mutant-specific RNA**

CTGGAA**a**AAGAAGAGTACAGTGCC

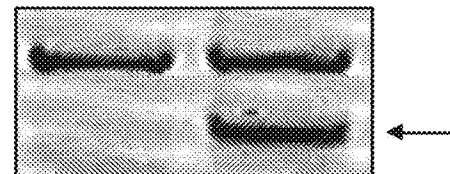


FIG. 33C

BRAF

HeLa

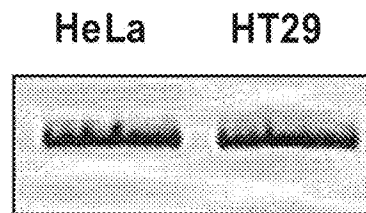
ACTCCATCGAGATTTCACTGTAG Wild-type

HT29

ACTCCATCGAGATTTCACTGTAG Wild-type

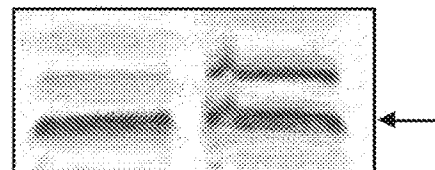
ACTCCATCGAGATTTCTCTGTAG (c.1799T>A)

PCR



**RFLP with
WT-Specific RNA**

ACTCCATCGAGATTTCACTGTAG



**RFLP with
Mutant-specific RNA**

ACTCCATCGAGATTTCTCTGTAG

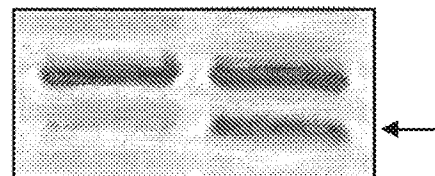


FIG. 33D

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**COMPOSITIONS FOR INDUCING
MODIFICATIONS OF TARGET
ENDOGENOUS NUCLEIC ACID SEQUENCES
IN NUCLEUSES OF EUKARYOTIC CELLS**

CROSS REFERENCE TO RELATED
APPLICATIONS

The present application is a continuation application of U.S. application Ser. No. 17/004, 338 filed Aug. 27, 2020, which is a continuation application of U.S. application Ser. No. 14/685, 568 filed Apr. 13, 2015, which is a continuation of PCT/KR2013/009488 filed Oct. 23, 2013, which claims priority to U.S. Provisional Application No. 61/837,481 filed on Jun. 20, 2013, U.S. Provisional Application No. 61/803, 599 filed Mar. 20, 2013, and U.S. Provisional Application No. 61/717, 324 filed Oct. 23, 2012, the entire contents of each aforementioned application are incorporated herein by reference.

SEQUENCE LISTING

The instant application contains a Sequence Listing which has been submitted electronically in XML file format and is hereby incorporated by reference in its entirety. Said XML copy, created on Oct. 29, 2024, is named 00138_SL.xml and is 495,164 bytes in size.

TECHNICAL FIELD

The present invention relates to targeted genome editing in eukaryotic cells or organisms. More particularly, the present invention relates to a composition for cleaving a target DNA in eukaryotic cells or organisms comprising a guide RNA specific for the target DNA and Cas protein-encoding nucleic acid or Cas protein, and use thereof.

BACKGROUND ART

CRISPRs (Clustered Regularly Interspaced Short Palindromic Repeats) are loci containing multiple short direct repeats that are found in the genomes of approximately 40% of sequenced bacteria and 90% of sequenced archaea. CRISPR functions as a prokaryotic immune system, in that it confers resistance to exogenous genetic elements such as plasmids and phages. The CRISPR system provides a form of acquired immunity. Short segments of foreign DNA, called spacers, are incorporated into the genome between CRISPR repeats, and serve as a memory of past exposures. CRISPR spacers are then used to recognize and silence exogenous genetic elements in a manner analogous to RNAi in eukaryotic organisms.

Cas9, an essential protein component in the Type II CRISPR/Cas system, forms an active endonuclease when complexed with two RNAs termed CRISPR RNA (crRNA) and trans-activating crRNA (tracrRNA), thereby slicing foreign genetic elements in invading phages or plasmids to protect the host cells. crRNA is transcribed from the CRISPR element in the host genome, which was previously captured from such foreign invaders. Recently, Jinek et al. (1) demonstrated that a single-chain chimeric RNA produced by fusing an essential portion of crRNA and tracrRNA could replace the two RNAs in the Cas9/RNA complex to form a functional endonuclease.

CRISPR/Cas systems offer an advantage to zinc finger and transcription activator-like effector DNA-binding proteins, as the site specificity in nucleotide binding CRISPR-

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Cas proteins is governed by a RNA molecule instead of the DNA-binding protein, which can be more challenging to design and synthesize.

However, until now, a genome editing method using the RNA-guided endonuclease (RGEN) based on CRISPR/Cas system has not been developed.

Meanwhile, Restriction fragment length polymorphism (RFLP) is one of the oldest, most convenient, and least expensive methods of genotyping that is still used widely in molecular biology and genetics but is often limited by the lack of appropriate sites recognized by restriction endonucleases.

Engineered nuclease-induced mutations are detected by various methods, which include mismatch-sensitive T7 endonuclease I (T7E1) or Surveyor nuclease assays, RFLP, capillary electrophoresis of fluorescent PCR products, Dideoxy sequencing, and deep sequencing. The T7E1 and Surveyor assays are widely used but are cumbersome. Furthermore, these enzymes tend to underestimate mutation frequencies because mutant sequences can form homoduplexes with each other and cannot distinguish homozygous bi-allelic mutant clones from wildtype cells. RFLP is free of these limitations and therefore is a method of choice. Indeed, RFLP was one of the first methods to detect engineered nuclease-mediated mutations in cells and animals. Unfortunately, however, RFLP is limited by the availability of appropriate restriction sites. It is possible that no restriction sites are available at the target site of interest.

DISCLOSURE OF INVENTION

Technical Problem

Until now, a genome editing and genotyping method using the RNA-guided endonuclease (RGEN) based on CRISPR/Cas system has not been developed.

Under these circumstances, the present inventors have made many efforts to develop a genome editing method based on CRISPR/Cas system and finally established a programmable RNA-guided endonuclease that cleave DNA in a targeted manner in eukaryotic cells and organisms.

In addition, the present inventors have made many efforts to develop a novel method of using RNA-guided endonucleases (RGENs) in RFLP analysis. They have used RGENs to genotype recurrent mutations found in cancer and those induced in cells and organisms by engineered nucleases including RGENs themselves, thereby completing the present invention.

Solution to Problem

It is an object of the present invention to provide a composition for cleaving target DNA in eukaryotic cells or organisms comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is another object of the present invention to provide a composition for inducing targeted mutagenesis in eukaryotic cells or organisms, comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a kit for cleaving a target DNA in eukaryotic cells or organisms comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

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It is still another object of the present invention to provide a kit for inducing targeted mutagenesis in eukaryotic cells or organisms comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a method for preparing a eukaryotic cell or organism comprising Cas protein and a guide RNA comprising a step of co-transfecting or serial-transfecting the eukaryotic cell or organism with a Cas protein-encoding nucleic acid or Cas protein, and a guide RNA or DNA that encodes the guide RNA.

It is still another object of the present invention to provide a eukaryotic cell or organism comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a method for cleaving a target DNA in eukaryotic cells or organisms comprising a step of transfecting the eukaryotic cells or organisms comprising a target DNA with a composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a method for inducing targeted mutagenesis in a eukaryotic cell or organism comprising a step of treating a eukaryotic cell or organism with a composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide an embryo, a genome-modified animal, or genome-modified plant comprising a genome edited by a composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a method of preparing a genome-modified animal comprising a step of introducing the composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein into an embryo of an animal; and a step of transferring the embryo into a oviduct of pseudopregnant foster mother to produce a genome-modified animal.

It is still another object of the present invention to provide a composition for genotyping mutations or variations in an isolated biological sample, comprising a guide RNA specific for the target DNA sequence Cas protein.

It is still another object of the present invention to provide a method of using a RNA-guided endonuclease (RGEN) to genotype mutations induced by engineered nucleases in cells or naturally-occurring mutations or variations, wherein the RGEN comprises a guide RNA specific for target DNA and Cas protein.

It is still another object of the present invention to provide a kit for genotyping mutations induced by engineered nucleases in cells or naturally-occurring mutations or variations, comprising a RNA-guided endonuclease (RGEN), wherein the RGEN comprises a guide RNA specific for target DNA and Cas protein.

It is an object of the present invention to provide a composition for cleaving target DNA in eukaryotic cells or organisms comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is another object of the present invention to provide a composition for inducing targeted mutagenesis in eukaryotic cells or organisms, comprising a guide RNA specific for

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target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a kit for cleaving a target DNA in eukaryotic cells or organisms comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a kit for inducing targeted mutagenesis in eukaryotic cells or organisms comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a method for preparing a eukaryotic cell or organism comprising Cas protein and a guide RNA comprising a step of co-transfecting or serial-transfecting the eukaryotic cell or organism with a Cas protein-encoding nucleic acid or Cas protein, and a guide RNA or DNA that encodes the guide RNA.

It is still another object of the present invention to provide a eukaryotic cell or organism comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a method for cleaving a target DNA in eukaryotic cells or organisms comprising a step of transfecting the eukaryotic cells or organisms comprising a target DNA with a composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a method for inducing targeted mutagenesis in a eukaryotic cell or organism comprising a step of treating a eukaryotic cell or organism with a composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide an embryo, a genome-modified animal, or genome-modified plant comprising a genome edited by a composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

It is still another object of the present invention to provide a method of preparing a genome-modified animal comprising a step of introducing the composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein into an embryo of an animal; and a step of transferring the embryo into a oviduct of pseudopregnant foster mother to produce a genome-modified animal.

It is still another object of the present invention to provide a composition for genotyping mutations or variations in an isolated biological sample, comprising a guide RNA specific for the target DNA sequence Cas protein.

It is still another object of the present invention to provide a composition for genotyping nucleic acid sequences in pathogenic microorganisms in an isolated biological sample, comprising a guide RNA specific for the target DNA sequence and Cas protein.

It is still another object of the present invention to provide a kit for genotyping mutations or variations in an isolated biological sample, comprising the composition, specifically comprising a RNA-guided endonuclease (RGEN), wherein the RGEN comprises a guide RNA specific for target DNA and Cas protein.

It is still another object of the present invention to provide a method of genotyping mutations or variations in an

isolated biological sample, using the composition, specifically comprising a RNA-guided endonuclease (RGEN), wherein the RGEN comprises a guide RNA specific for target DNA and Cas protein.

Advantageous Effects of Invention

The present composition for cleaving a target DNA or inducing a targeted mutagenesis in eukaryotic cells or organisms, comprising a guide RNA specific for the target DNA and Cas protein-encoding nucleic acid or Cas protein, the kit comprising the composition, and the method for inducing targeted mutagenesis provide a new convenient genome editing tools. In addition, because custom RGENs can be designed to target any DNA sequence, almost any single nucleotide polymorphism or small insertion/deletion (indel) can be analyzed via RGEN-mediated RFLP, therefore, the composition and method of the present invention may be used in detection and cleaving naturally-occurring variations and mutations.

BRIEF DESCRIPTION OF DRAWINGS

FIGS. 1A and 1B show Cas9-catalyzed cleavage of plasmid DNA in vitro. FIG. 1A: Schematic representation of target DNA (SEQ ID NO: 112) and chimeric RNA sequences (SEQ ID NO: 113). Triangles indicate cleavage sites. The PAM sequence recognized by Cas9 is shown in bold. The sequences in the guide RNA (SEQ ID NO: 113) derived from crRNA and tracrRNA are shown in box and underlined, respectively. FIG. 1B: In vitro cleavage of plasmid DNA by Cas9. An intact circular plasmid or ApaLI-digested plasmid was incubated with Cas9 and guide RNA.

FIGS. 2A and 2B show Cas9-induced mutagenesis at an episomal target site. FIG. 2A: Schematic overview of cell-based assays using a RFP-GFP reporter. GFP is not expressed from this reporter because the GFP sequence is fused to the RFP sequence out-of-frame. The RFP-GFP fusion protein is expressed only when the target site between the two sequences is cleaved by a site-specific nuclease. FIG. 2B: Flow cytometry of cells transfected with Cas9. The percentage of cells that express the RFP-GFP fusion protein is indicated.

FIGS. 3A and 3B show RGEN-driven mutations at endogenous chromosomal sites. FIG. 3A: CCR5 locus. FIG. 3B: C4BPB locus. (Top) The T7E1 assay was used to detect RGEN-driven mutations. Arrows indicate the expected position of DNA bands cleaved by T7E1. Mutation frequencies (Indels (%)) were calculated by measuring the band intensities. (Bottom) DNA sequences of the wild-type (WT) CCR5 (SEQ ID NO: 114) and C4BPB (SEQ ID NO: 122) and mutant clones. DNA sequences of RGEN-induced mutations at the CCR5 locus: +1 (SEQ ID NO: 115), -13 (SEQ ID NO: 116), -14 (SEQ ID NO: 117), -18 (SEQ ID NO: 118), -19 (SEQ ID NO: 119), -24 (SEQ ID NO: 120), and -30 (SEQ ID NO: 121). DNA sequences of RGEN-induced mutations at the C4BPB locus: +1 (SEQ ID NO: 122), +2 (SEQ ID NO: 123), -30 (SEQ ID NO: 125), and -180 (SEQ ID NO: 126). The region of the target sequence complementary to the guide RNA is shown in box. The PAM sequence is shown in bold. Triangles indicate the cleavage site. Bases corresponding to microhomologies are underlined. The column on the right indicates the number of inserted or deleted bases.

FIGS. 4A, 4B, and 4C show that RGEN-driven off-target mutations are undetectable. FIG. 4A: On-target and potential off-target sequences. The human genome was searched in

silico for potential off-target sites. Four sites were identified, ADCY5 (SEQ ID NO: 128), KCNJ6 (SEQ ID NO: 129), CNTNAP2 (SEQ ID NO: 130), and Chr. 5 N/A (SEQ ID NO: 131), each of which carries 3-base mismatches with the CCR5 on-target (SEQ ID NO: 127). Mismatched bases are underlined. FIG. 4B: The T7E1 assay was used to investigate whether these sites were mutated in cells transfected with the Cas9/RNA complex. No mutations were detected at these sites. N/A (not applicable), an intergenic site. FIG. 4C: Cas9 did not induce off-target-associated chromosomal deletions. The CCR5-specific RGEN and ZEN were expressed in human cells. PCR was used to detect the induction of the 15-kb chromosomal deletions in these cells.

FIGS. 5A, 5B, 5C, and 5D show RGEN-induced Foxn1 gene targeting in mice. FIG. 5A: A schematic diagram depicting target DNA (SEQ ID NO: 132) and a sgRNA specific to exon 2 of the mouse Foxn1 gene (SEQ ID NO: 133). PAM in exon 2 is shown in a box and the sequence in the sgRNA that is complementary to exon 2 is underlined. Triangles indicate cleavage sites. FIG. 5B: Representative T7E1 assays demonstrating gene-targeting efficiencies of Cas9 mRNA plus Foxn1-specific sgRNA that were delivered via intra-cytoplasmic injection into one-cell stage mouse embryos. Numbers indicate independent founder mice generated from the highest dose. Arrows indicate bands cleaved by T7E1. FIG. 5C: DNA sequences of wild-type (WT) Foxn1 (SEQ ID NO: 134) and mutant alleles (SEQ ID NOs. 135-141) observed in three Foxn1 mutant founders identified in FIG. 5B. DNA sequences of mutant alleles in founder #108: -44 (SEQ ID NO: 135), -23 (SEQ ID NO: 136), -17 (SEQ ID NO: 137), and +1 (SEQ ID NO: 138). DNA sequences of mutant alleles in founder #111: +1 (SEQ ID NO: 138) and -11 (SEQ ID NO: 139). DNA sequences of mutant alleles in founder #114: -6 (SEQ ID NO: 140), -17 (SEQ ID NO: 137), and -8 (SEQ ID NO: 141). The number of occurrences is shown in parentheses. FIG. 5D: PCR genotyping of F1 progenies derived from crossing Foxn1 founder #108 and wild-type FVB/NTac. Note the segregation of the mutant alleles found in Foxn1 founder #108 in the progenies.

FIGS. 6A, 6B, and 6C show Foxn1 gene targeting in mouse embryos by intra-cytoplasmic injection of Cas9 mRNA and Foxn1-sgRNA. FIG. 6A: A representative result of a T7E1 assay monitoring the mutation rate after injecting the highest dose. Arrows indicate bands cleaved by T7E1. FIG. 6B: A summary of T7E1 assay results. Mutant fractions among in vitro cultivated embryos obtained after intra-cytoplasmic injection of the indicated RGEN doses are indicated. FIG. 6C: DNA sequences of wild-type (WT) Foxn1 (SEQ ID NO: 143) and Foxn1 mutant alleles (SEQ ID Nos. 144-152) identified from a subset of T7E1-positive mutant embryos. The DNA sequences of the mutant alleles are: $\Delta 11$ (SEQ ID NO: 144), $\Delta 11+\Delta 17$ (SEQ ID NO: 145) $\Delta 57$ (SEQ ID NO: 146), $\Delta 17$ (SEQ ID NO: 147), +1 (SEQ ID NO: 148), $\Delta 12$ (SEQ ID NO: 149), $\Delta 72$ (SEQ ID NO: 150), $\Delta 25$ (SEQ ID NO: 151), $\Delta 24$ (SEQ ID NO: 152). The target sequence of the wild-type allele is denoted in box.

FIGS. 7A, 7B, and 7C show Foxn1 gene targeting in mouse embryos using the recombinant Cas9 protein: Foxn1-sgRNA complex. FIG. 7A and FIG. 7B are representative T7E1 assays results and their summaries. Embryos were cultivated in vitro after they underwent pronuclear (FIG. 7A) or intra-cytoplasmic injection (FIG. 7B). Underlined numbers indicate T7E1-positive mutant founder mice. FIG. 7C: DNA sequences of wild-type (WT) Foxn1 (SEQ ID NO: 153) and Foxn1 mutant alleles (SEQ ID NOs. 154-166) identified from the in vitro cultivated embryos that were

obtained by the pronucleus injection of recombinant Cas9 protein: Foxn1-sgRNA complex at the highest dose. The target sequence of the wild-type allele is denoted in box. The DNA sequences of the mutant alleles are: $\Delta 18$ (SEQ ID NO: 154), $\Delta 20$ (SEQ ID NO: 155), $\Delta 19$ (SEQ ID NO: 156), $\Delta 17$ (SEQ ID NO: 157), $\Delta 11$ (SEQ ID NO: 158), $\Delta 3+1$ (SEQ ID NO: 159), $\Delta 2$ (SEQ ID NO: 160), +1, Embryo 1 (SEQ ID NO: 161), +1, Embryo 10 (SEQ ID NO: 162), $\Delta 6$ (SEQ ID NO: 163), $\Delta 5$ (SEQ ID NO: 164), $\Delta 28$ (SEQ ID NO: 165), and $\Delta 126$ (SEQ ID NO: 166).

FIGS. 8A, 8B, and 8C show Germ-line transmission of the mutant alleles found in Foxn1 mutant founder #12. FIG. 8A: wild type *f*PCR analysis. FIG. 8B: Foxn1 mutant founder #12 *f*PCR analysis. FIG. 8C: PCR genotyping of wild-type FVB/NTac, the founder mouse, and their F1 progenies.

FIGS. 9A and 9B show Genotypes of embryos generated by crossing Prkdc mutant founders. Prkdc mutant founders $\delta 25$ and $\eta 15$ were crossed and E13.5 embryos were isolated. FIG. 9A: *f*PCR analysis of wild-type, founder $\delta 25$, and founder $\eta 15$. Note that, due to the technical limitations of *f*PCR analysis, these results showed small differences from the precise sequences of the mutant alleles; e.g., from the sequence analysis, $\Delta 269/\Delta 61/\text{WT}$ and $\Delta 5+1/+7/+12/\text{WT}$ were identified in founders $\delta 25$ and $\eta 15$, respectively. FIG. 9B: Genotypes of the generated embryos.

FIGS. 10A, 10B, 10C, 10D, and 10E show Cas9 protein/sgRNA complex induced targeted mutation at CCR5 gene (FIGS. 10A-10C) and ABCC11 gene (FIGS. 10D-10E). FIG. 10A: Results of a T7E1 assay monitoring the mutation rate at CCR5 locus after introducing Cas9 protein and sgRNA or Cas9 protein and crRNA+tracrRNA into K562 cells. FIG. 10B: Results of a T7E1 assay using $1/5$ scaled down doses of Cas9 protein and sgRNA. FIG. 10C: Wild-type (WT) CCR5 sequence (SEQ ID NO: 114) and Cas protein induced mutant sequences (SEQ ID NOs. 167-171 and 115) identified in CCR5 locus. The DNA sequences of the mutant sequences are: -4 (SEQ ID NO: 167), -4 (SEQ ID NO: 168), -7 (SEQ ID NO: 169), -1 (SEQ ID NO: 170), +1 (SEQ ID NO: 115), and -17, +1 (SEQ ID NO: 171). FIG. 10D: Results of a T7E1 assay monitoring the mutation rate at ABCC11 locus after introducing Cas9 protein and sgRNA into K562 cells. FIG. 10E: Wild-type (WT) ABCC11 sequence (SEQ ID NO: 172) and Cas9 protein induced mutant sequences (SEQ ID NOs. 173-176) identified in ABCC11 locus. The DNA sequences of the mutant sequences are: -6 (SEQ ID NO: 173), -3 (SEQ ID NO: 174), -29 (SEQ ID NO: 175), -20 (SEQ ID NO: 176), and -256 (TTCTC).

FIG. 11 shows recombinant Cas9 protein-induced mutations in Arabidopsis protoplasts.

FIG. 12 shows wild type BRIL sequence (SEQ ID NO: 177) and recombinant Cas9 protein-induced mutant sequences (SEQ ID NOs. 178-181) in the Arabidopsis BRIL gene. The DNA sequences of the mutant sequences are: -7 (SEQ ID NO: 178), -224 (SEQ ID NO: 179), -223 (SEQ ID NO: 180), and -223, +62 (SEQ ID NO: 181).

FIG. 13 shows T7E1 assay showing endogenous CCR5 gene disruption in 293 cells by treatment of Cas9-mal-9R4L and sgRNA/C9R4LC complex.

FIGS. 14A and 14B show mutation frequencies at on-target and off-target sites of RGENs reported in Fu et al. (2013). T7E1 assays analyzing genomic DNA from K562 cells (R) transfected serially with 20 μg of Cas9-encoding plasmid and with 60 μg and 120 μg of in vitro transcribed GX19 crRNA and tracrRNA, respectively (1×10^6 cells), or (D) co-transfected with 1 μg of Cas9-encoding plasmid and

1 μg of GX19 SgRNA expression plasmid (2×10^5 cells). FIG. 14A: VEGFA site 1 on target sequence (SEQ ID NO: 182) and off target sequences, OT1-3 (SEQ ID NO: 183) and OT1-11 (SEQ ID NO: 184). VEGFA site 2 on target sequence (SEQ ID NO: 185) and off target sequences OT2-1 (SEQ ID NO: 186), OT2-9 (SEQ ID NO: 187) and OT2-24 (SEQ ID NO: 188). FIG. 14B: VEGFA site 3 on target sequence (SEQ ID NO: 189) and off target sequence OT3-18 (SEQ ID NO: 190) and EMX1 on target sequence (SEQ ID NO: 191) and off target sequence OT4-1 (SEQ ID NO: 192).

FIGS. 15A and 15B show comparison of guide RNA structure. Mutation frequencies of the RGENs reported in Fu et al. (2013) were measured at on-target and off-target sites using the T7E1 assay. K562 cells were co-transfected with the Cas9-encoding plasmid and the plasmid encoding GX19 sgRNA or GGX20 sgRNA. Off-target sites (OT1-3 etc.) are labeled as in Fu et al. (2013). FIG. 15A: VEGFA site 1 on target sequence (SEQ ID NO: 182) and off target sequences OT1-3 (SEQ ID NO: 183 and OT1-11 (SEQ ID NO: 184). VEGFA site 2 on target sequence (SEQ ID NO: 185) and off target sequences OT2-1 (SEQ ID NO: 186), OT2-9 (SEQ ID NO: 187), and OT2-24 (SEQ ID NO: 188). FIG. 15B: VEGFA site 3 on target sequence (SEQ ID NO: 189) and off target sequence OT3-18 (SEQ ID NO: 190) and EMX1 on target sequence (SEQ ID NO: 191) and off target sequence OT4-1 (SEQ ID NO: 192).

FIGS. 16A, 16B, 16C, and 16D show that in vitro DNA cleavage by Cas9 nickases. FIG. 16A: Schematic overview of the Cas9 nuclease and the paired Cas9 nickase. The PAM sequences and cleavage sites are shown in box. FIG. 16B: Target sites in the human AAVS1 locus. The position of each target site is shown in triangle. FIG. 16C: Schematic overview of DNA cleavage reactions. FAM dyes (shown in box) were linked to both 5' ends of the DNA substrate. FIG. 16D: DSBs and SSBs analyzed using fluorescent capillary electrophoresis. Fluorescently-labeled DNA substrates were incubated with Cas9 nucleases or nickases before electrophoresis.

FIGS. 17A and 17B show comparison of Cas9 nuclease and nickase behavior. FIG. 17A: On-target mutation frequencies associated with Cas9 nucleases (WT), nickases (D10A), and paired nickases at the following target sequences of the AAVS1 locus: S1 (SEQ ID NO: 193), S2 (SEQ ID NO: 194), S3 (SEQ ID NO: 195), S4 (SEQ ID NO: 196), S5 (SEQ ID NO: 197), S6 (SEQ ID NO: 198), AS1 (SEQ ID NO: 199), AS2 (SEQ ID NO: 200), and AS3 (SEQ ID NO: 201). Paired nickases that would produce 5' overhangs or 3' overhangs are indicated. FIG. 17B: Analysis of off-target effects of Cas9 nucleases and paired nickases. A total of seven potential off-target sites (SEQ ID NOs. 202-208) for three sgRNAs were analyzed. The mutation frequency for the S2 on-target sequence (SEQ ID NO: 194) was compared to the off-target sequences, S2 Off-1 (SEQ ID NO: 202) and S2 Off-2 (SEQ ID NO: 203). The mutation frequency for the S3 on-target sequence (SEQ ID NO: 195) was compared to the off-target sequences, S3 Off-1 (SEQ ID NO: 204) and S3 Off-2 (SEQ ID NO: 205). The mutation frequency for the AS2 on-target sequence (SEQ ID NO: 198) was compared to the off-target sequences, AS2 Off-1 (SEQ ID NO: 206), AS2 Off-6 (SEQ ID NO: 207), and AS2 Off-9 (SEQ ID NO: 208).

FIGS. 18A, 18B, 18C, and 18D show paired Cas9 nickases tested at other endogenous human loci. The sgRNA target sites at the human CCR5 locus (FIG. 18A; SEQ ID NO: 209) and the BRCA2 locus (FIG. 18C; SEQ ID NO: 210). PAM sequences are indicated in a box. Genome editing activities at CCR5 (FIG. 18B) and BRCA2 (FIG.

18D) target sites were detected by the T7E1 assay. The repair of two nicks that would produce 5' overhangs led to the formation of indels much more frequently than did those producing 3' overhangs.

FIGS. 19A and 19B show that paired Cas9 nickases mediate homologous recombination. FIG. 19A: Strategy to detect homologous recombination. Donor DNA included an XbaI restriction enzyme site between two homology arms, whereas the endogenous target site lacked this site. A PCR assay was used to detect sequences that had undergone homologous recombination. To prevent amplification of contaminating donor DNA, primers specific to genomic DNA were used. FIG. 19B: Efficiency of homologous recombination. Only amplicons of a region in which homologous recombination had occurred could be digested with XbaI; the intensities of the cleavage bands were used to measure the efficiency of this method.

FIGS. 20A, 20B, 20C, and 20D show DNA splicing induced by paired Cas9 nickases. FIG. 20A: The target sites of paired nickases in the human AAVS1 locus. The distances between the AS2 site and each of the other sites are shown. Arrows indicate PCR primers. FIG. 20B: Genomic deletions detected using PCR. Asterisks indicate deletion-specific PCR products. FIG. 20C: DNA sequences of wild-type (WT) (SEQ ID NO: 211 and 332) and the following deletion-specific PCR products (SEQ ID Nos. 212-218) obtained using AS2 sgRNAs or deletion-specific PCR products (SEQ ID NOs. 219-224) using L1 sgRNAs. Target site PAM sequences are shown in box and sgRNA-matching sequences are shown in capital letters. Intact sgRNA-matching sequences are underlined. FIG. 20D: A schematic model of paired Cas9 nickase-mediated chromosomal deletions. Newly-synthesized DNA strands are shown in box.

FIGS. 21A, 21B, and 21C show that paired Cas9 nickases do not induce translocations. FIG. 21A: Schematic overview of chromosomal translocations between the on-target and off-target sites. FIG. 21B: PCR amplification to detect chromosomal translocations. FIG. 21C: Translocations induced by Cas9 nucleases but not by the nickase pair.

FIGS. 22A and 22B show a conceptual diagram of the T7E1 and RFLP assays. FIG. 22A: Comparison of assay cleavage reactions in four possible scenarios after engineered nuclease treatment in a diploid cell: (A) wild type, (B) a monoallelic mutation, (C) different biallelic mutations (hetero), and (D) identical biallelic mutations (homo). Black lines represent PCR products derived from each allele; dashed and dotted boxes indicate insertion/deletion mutations generated by NHEJ. FIG. 22B: Expected results of T7E1 and RGEN digestion resolved by electrophoresis.

FIG. 23 shows in vitro cleavage assay of a linearized plasmid containing the C4BPB target site bearing indels. DNA sequences of individual plasmid substrates (upper panel): WT (SEQ ID NO: 104), 11 (SEQ ID NO: 225), 12 (SEQ ID NO: 226), 13 (SEQ ID NO: 227), D1 (SEQ ID NO: 228), D2 (SEQ ID NO: 229), and D3 (SEQ ID NO: 230). The PAM sequence is underlined. Inserted bases are shown in box. Arrows (bottom panel) indicate expected positions of DNA bands cleaved by the wild-type-specific RGEN after electrophoresis.

FIGS. 24A and 24B show genotyping of mutations induced by engineered nucleases in cells via RGEN-mediated RFLP. FIG. 24A: Genotype of C4BPB wild type (SEQ ID NO: 231) and the following mutant K562 cell clones: +3 (SEQ ID NO: 232), -12 (SEQ ID NO: 233), -9 (SEQ ID NO: 234), -8 (SEQ ID NO: 235), -36 (SEQ ID NO: 236), +1 (SEQ ID NO: 237), +1 (SEQ ID NO: 238), +67 (SEQ ID NO: 239), -7, +1 (SEQ ID NO: 240), -94 (SEQ ID NO:

241). FIG. 24B: Comparison of the mismatch-sensitive T7E1 assay with RGEN-mediated RFLP analysis. Black arrows indicate the cleavage product by treatment of T7E1 enzyme or RGENS.

FIGS. 25A, 25B, and 25C show genotyping of RGEN-induced mutations via the RGEN-RFLP technique. FIG. 25A: Analysis of C4BPB-disrupted clones using RGEN-RFLP and T7E1 assays. Arrows indicate expected positions of DNA bands cleaved by RGEN or T7E1. FIG. 25B: Quantitative comparison of RGEN-RFLP analysis with T7E1 assays. Genomic DNA samples from wild-type and C4BPB-disrupted K562 cells were mixed in various ratios and subjected to PCR amplification. FIG. 25C: Genotyping of RGEN-induced mutations in the HLA-B gene in HeLa cells with RFLP and T7E1 analyses.

FIGS. 26A and 26B show genotyping of mutations induced by engineered nucleases in organisms via RGEN-mediated RFLP. FIG. 26A: Genotype of Pibf1 wild-type (WT) (SEQ ID NO: 242) and the following mutant founder mice: #1 (SEQ ID NO: 243 and SEQ ID NO: 244), #3 (SEQ ID NO: 245 and SEQ ID NO: 246), #4 (SEQ ID NO: 247 and SEQ ID NO: 242), #5 (SEQ ID NO: 246 and SEQ ID NO: 242), #6 (SEQ ID NO: 248 and SEQ ID NO: 249), #8 (SEQ ID NO: 250 and SEQ ID NO: 251), and #11 (SEQ ID NO: 252 and SEQ ID NO: 250). FIG. 26B: Comparison of the mismatch-sensitive T7E1 assay with RGEN-mediated RFLP analysis. Black arrows indicate the cleavage product by treatment of T7E1 enzyme or RGENS.

FIG. 27 shows RGEN-mediated genotyping of ZFN-induced mutations at a wild-type CCR5 sequence (SEQ ID NO: 253). The ZEN target site is shown in box. Black arrows indicate DNA bands cleaved by T7E1.

FIG. 28 shows polymorphic sites in a region of the human HLA-B gene (SEQ ID NO: 254). The sequence, which surrounds the RGEN target site, is that of a PCR amplicon from HeLa cells. Polymorphic positions are shown in box. The RGEN target site and the PAM sequence are shown in dashed and bolded box, respectively. Primer sequences are underlined.

FIGS. 29A and 29B show genotyping of oncogenic mutations via RGEN-RFLP analysis. FIG. 29A: A recurrent mutation (c.133-135 deletion of TCT; SEQ ID NO: 256) in the human CTNNB1 gene in HCT116 cells was detected by RGENs. The wild-type CTNNB1 sequence is represented by SEQ ID NO: 255. HeLa cells were used as a negative control. FIG. 29B: Genotyping of the KRAS substitution mutation (c.34 G>A) in the A549 cancer cell line with RGENs that contain mismatched guide RNA that are WT-specific (SEQ ID NO: 257) or mutant-specific (SEQ ID NO: 258). Mismatched nucleotides are shown in box. HeLa cells were used as a negative control. Arrows indicate DNA bands cleaved by RGENs. DNA sequences confirmed by Sanger sequencing are shown: wild-type (SEQ ID NO: 259) and c.34G>A (SEQ ID NO: 260).

FIGS. 30A, 30B, 30C, and 30D show genotyping of the CCR5 delta32 allele in HEK293T cells via RGEN-RFLP analysis. FIG. 30A: RGEN-RFLP assays of cell lines. DNA sequences of the wild-type CCR5 locus (SEQ ID NO: 262) and delta 32 mutation (SEQ ID NO: 261) are shown. K562, SKBR3, and HeLa cells were used as wild-type controls. Arrows indicate DNA bands cleaved by RGENs. FIG. 30B: DNA sequence of wild-type (SEQ ID NO: 263) and delta32 CCR5 alleles (SEQ ID NO: 264). Both on-target and off-target sites of RGENs used in RFLP analysis are underlined. A single-nucleotide mismatch between the two sites is shown in box. The PAM sequence is underlined. FIG. 30C: In vitro cleavage of plasmids harboring WT or del32 CCR5

alleles using the wild-type-specific RGEN. FIG. 30D Confirming the presence of an off-target site of the CCR5-delta32-specific RGEN at the CCR5 locus. In vitro cleavage assays of plasmids harboring either on-target (SEQ ID NO: 265) or off-target sequences (SEQ ID NO: 266) using various amounts of the del32-specific RGEN.

FIGS. 31A and 31B show genotyping of a KRAS point mutation (c.34 G>A). FIG. 31A: RGEN-RFLP analysis of the KRAS mutation (c.34 G>A) in cancer cell lines. PCR products from Hela cells (used as a wild-type control) or A549 cells, which are homozygous for the point mutation, were digested with RGENs with perfectly matched crRNA specific to the wild-type sequence (SEQ ID NO: 259) or the mutant sequence (SEQ ID NO: 260). KRAS genotypes in these cells were confirmed by Sanger sequencing. FIG. 31B: Plasmids harboring either the wild-type (SEQ ID NO: 259) or mutant KRAS sequences (SEQ ID NO: 260) were digested using RGENs with perfectly matched crRNAs or attenuated, one-base mismatched crRNAs: m7 (SEQ ID NO: 267), m6 (SEQ ID NO: 257), m5 (SEQ ID NO: 268), m4 (SEQ ID NO: 269), m8 (SEQ ID NO: 260), m7, 8 (SEQ ID NO: 270), m6, 8 (SEQ ID NO: 258), m5, 8 (SEQ ID NO: 271), and m4, 8 (SEQ ID NO: 272). Attenuated crRNAs that were chosen for genotyping are labeled in box above the gels.

FIGS. 32A and 32B show genotyping of a PIK3CA point mutation (c.3140 A>G). FIG. 32A: RGEN-RFLP analysis of the PIK3CA mutation (c.3140 A>G) in cancer cell lines. PCR products from Hela cells (used as a wild-type control) or HCT116 cells that are heterozygous for the point mutation were digested with RGENs with perfectly matched crRNA specific to the wild-type sequence (SEQ ID NO: 273) or the mutant sequence (SEQ ID NO: 274). PIK3CA genotypes in these cells were confirmed by Sanger sequencing. FIG. 32B: Plasmids harboring either the wild-type PIK3CA sequence (SEQ ID NO: 273) or mutant PIK3CA sequence (SEQ ID NO: 274) were digested using RGENs with perfectly matched crRNAs or attenuated, one-base mismatched crRNAs: m5 (SEQ ID NO: 275), m6 (SEQ ID NO: 276), m7 (SEQ ID NO: 277), m10 (SEQ ID NO: 278), m13 (SEQ ID NO: 279), m16 (SEQ ID NO: 280), m19 (SEQ ID NO: 281), m4 (SEQ ID NO: 274), m4, 5 (SEQ ID NO: 282), m4, 6 (SEQ ID NO: 283), m4, 7 (SEQ ID NO: 284), m4,10 (SEQ ID NO: 285), m4,13 (SEQ ID NO: 286), m4, 16 (SEQ ID NO: 287), and m4, 19 (SEQ ID NO: 288). Attenuated crRNAs that were chosen for genotyping are labeled in box above the gels.

FIGS. 33A, 33B, 33C, and 33D show genotyping of recurrent point mutations in cancer cell lines. FIG. 33A: RGEN-RFLP assays to distinguish between a wild-type IDH gene sequence (SEQ ID NO: 289) and a recurrent oncogenic point mutation sequence in the IDH gene (c.394c>T; SEQ ID NO: 290). RGENs with attenuated, one-base mismatched crRNAs, SEQ ID NO: 291 (WT-Specific RNA) and SEQ ID NO: 292 (Mutant-Specific RNA), distinguished the wild type and mutant IDH sequences. FIG. 33B: RGEN-RFLP assays to distinguish between a wild-type PIK3CA gene sequence (SEQ ID NO: 271) and a recurrent oncogenic point mutation sequence in the PIK3CA gene (c.3140A>G; SEQ ID NO: 273). RGENs with attenuated, one-base mismatched crRNAs, SEQ ID NO: 275 (WT-Specific RNA) and SEQ ID NO: 284 (Mutant-Specific RNA), distinguished the wild type and mutant PIK3CA sequences. FIG. 33C: RGEN-RFLP assays to distinguish between a wild-type NRAS gene sequence (SEQ ID NO: 293) and a recurrent oncogenic point mutation sequence in the NRAS gene (c.181C>A; SEQ ID NO: 294). RGENs with perfectly matched crRNAs, SEQ ID

NO: 293 (WT-Specific RNA) and SEQ ID NO: 294 (Mutant-Specific RNA), distinguished the wild type and mutant NRAS sequences. FIG. 33D: RGEN-RFLP assays to distinguish between a wild-type BRAF gene sequence (SEQ ID NO: 295) and a recurrent oncogenic point mutation sequence in the BRAF gene (c.1799T>A; SEQ ID NO: 296). RGENs with perfectly matched crRNAs, SEQ ID NO: 295 (WT-Specific RNA) and SEQ ID NO: 296 (Mutant-Specific RNA), distinguished the wild type and mutant BRAF sequences. Genotypes of each cell line confirmed by Sanger sequencing are shown. Mismatched nucleotides are shown in box. Black arrows indicate DNA bands cleaved by RGENs.

BEST MODE FOR CARRYING OUT THE INVENTION

In accordance with one aspect of the invention, the present invention provides a composition for cleaving target DNA in eukaryotic cells or organisms comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein. In addition, the present invention provides a use of the composition for cleaving target DNA in eukaryotic cells or organisms comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

In the present invention, the composition is also referred to as a RNA-guided endonuclease (RGEN) composition.

ZFNs and TALENs enable targeted mutagenesis in mammalian cells, model organisms, plants, and livestock, but the mutation frequencies obtained with individual nucleases are widely different from each other. Furthermore, some ZFNs and TALENs fail to show any genome editing activities. DNA methylation may limit the binding of these engineered nucleases to target sites. In addition, it is technically challenging and time-consuming to make customized nucleases.

The present inventors have developed a new RNA-guided endonuclease composition based on Cas protein to overcome the disadvantages of ZFNs and TALENs.

Prior to the present invention, an endonuclease activity of Cas proteins has been known. However, it has not been known whether the endonuclease activity of Cas protein would function in an eukaryotic cell because of the complexity of the eukaryotic genome. Further, until now, a composition comprising Cas protein or Cas protein-encoding nucleic acid and a guide RNA specific for the target DNA to cleave a target DNA in eukaryotic cells or organisms has not been developed.

Compared to ZFNs and TALENs, the present RGEN composition based on Cas protein can be more readily customized because only the synthetic guide RNA component is replaced to make a new genome-editing nuclease. No sub-cloning steps are involved to make customized RNA guided endonucleases. Furthermore, the relatively small size of the Cas gene (for example, 4.2 kbp for Cas9) as compared to a pair of TALEN genes (~6 kbp) provides an advantage for this RNA-guided endonuclease composition in some applications such as virus-mediated gene delivery. Further, this RNA-guided endonuclease does not have off-target effects and thus does not induce unwanted mutations, deletion, inversions, and duplications. These features make the present RNA-guided endonuclease composition a scalable, versatile, and convenient tool for genome engineering in eukaryotic cells and organisms. In addition, RGEN can be designed to target any DNA sequence, almost any single nucleotide polymorphism or small insertion/deletion (indel)

can be analyzed via RGEN-mediated RFLP. The specificity of RGENS is determined by the RNA component that hybridizes with a target DNA sequence of up to 20 base pairs (bp) in length and by the Cas9 protein that recognizes the protospacer-adjacent motif (PAM). RGENS are readily reprogrammed by replacing the RNA component. Therefore, RGENs provide a platform to use simple and robust ROOFLP analysis for various sequence variations.

The target DNA may be an endogenous DNA, or artificial DNA, preferably, endogenous DNA.

As used herein, the term “Cas protein” refers to an essential protein component in the CRISPR/Cas system, forms an active endonuclease or nickase when complexed with two RNAs termed CRISPR RNA (crRNA) and transactivating crRNA (tracrRNA).

The information on the gene and protein of Cas are available from GenBank of National Center for Biotechnology Information (NCBI), without limitation.

The CRISPR-associated (cas) genes encoding Cas proteins are often associated with CRISPR repeat-spacer arrays. More than forty different Cas protein families have been described. Of these protein families, Cas1 appears to be ubiquitous among different CRISPR/Cas systems. There are three types of CRISPR-Cas system. Among them, Type II CRISPR/Cas system involving Cas9 protein and crRNA and tracrRNA is representative and is well known. Particular combinations of cas genes and repeat structures have been used to define 8 CRISPR subtypes (*Ecoli*, *Ypest*, *Nmeni*, *Dvulg*, *Tneap*, *Hmari*, *Apern*, and *Mtube*).

The Cas protein may be linked to a protein transduction domain. The protein transduction domain may be poly-arginine or a TAT protein derived from HIV, but it is not limited thereto.

The present composition may comprise Cas component in the form of a protein or in the form of a nucleic acid encoding Cas protein.

In the present invention, Cas protein may be any Cas protein provided that it has an endonuclease or nickase activity when complexed with a guide RNA.

Preferably, Cas protein is Cas9 protein or variants thereof.

The variant of the Cas9 protein may be a mutant form of Cas9 in which the catalytic aspartate residue is changed to any other amino acid. Preferably, the other amino acid may be an alanine, but it is not limited thereto.

Further, Cas protein may be the one isolated from an organism such as *Streptococcus* sp., preferably *Streptococcus pyogenes* or a recombinant protein, but it is not limited thereto.

The Cas protein derived from *Streptococcus pyogenes* may recognize NGG trinucleotide. The Cas protein may comprise an amino acid sequence of SEQ ID NO: 109, but it is not limited thereto.

The term “recombinant” when used with reference, e.g., to a cell, nucleic acid, protein, or vector, indicates that the cell, nucleic acid, protein or vector, has been modified by the introduction of a heterologous nucleic acid or protein or the alteration of a native nucleic acid or protein, or that the cell is derived from a cell so modified. Thus, for example, a recombinant Cas protein may be generated by reconstituting Cas protein-encoding sequence using the human codon table.

As for the present invention, Cas protein-encoding nucleic acid may be a form of vector, such as plasmid comprising Cas-encoding sequence under a promoter such as CMV or CAG. When Cas protein is Cas9, Cas9 encoding sequence may be derived from *Streptococcus* sp., and preferably derived from *Streptococcus pyogenes*. For example,

Cas9 encoding nucleic acid may comprise the nucleotide sequence of SEQ ID. NO: 1. Moreover, Cas9 encoding nucleic acid may comprise the nucleotide sequence having homology of at least 50% to the sequence of SEQ ID NO: 1, preferably at least 60, 70, 80, 90, 95, 97, 98, or 99% to the SEQ ID NO:1, but it is not limited thereto. Cas9 encoding nucleic acid may comprise the nucleotide sequence of SEQ ID NOs.108, 110, 106, or 107.

As used herein, the term “guide RNA” refers to a RNA which is specific for the target DNA and can form a complex with Cas protein and bring Cas protein to the target DNA.

In the present invention, the guide RNA may consist of two RNA, i.e., CRISPR RNA (crRNA) and transactivating crRNA (tracrRNA) or be a single-chain RNA (sgRNA) produced by fusion of an essential portion of crRNA and tracrRNA.

The guide RNA may be a dualRNA comprising a crRNA and a tracrRNA.

If the guide RNA comprises the essential portion of crRNA and tracrRNA and a portion complementary to a target, any guide RNA may be used in the present invention.

The crRNA may hybridize with a target DNA.

The RGEN may consist of Cas protein, and dualRNA (invariable tracrRNA and target-specific crRNA), or Cas protein and sgRNA (fusion of an essential portion of invariable tracrRNA and target-specific crRNA), and may be readily reprogrammed by replacing crRNA.

The guide RNA further comprises one or more additional nucleotides at the 5' end of the single-chain guide RNA or the crRNA of the dualRNA.

Preferably, the guide RNA further comprises 2-additional guanine nucleotides at the 5' end of the single-chain guide RNA or the crRNA of the dualRNA.

The guide RNA may be transferred into a cell or an organism in the form of RNA or DNA that encodes the guide RNA.

The guide RNA may be in the form of an isolated RNA, RNA incorporated into a viral vector, or is encoded in a vector. Preferably, the vector may be a viral vector, plasmid vector, or agrobacterium vector, but it is not limited thereto.

A DNA that encodes the guide RNA may be a vector comprising a sequence coding for the guide RNA. For example, the guide RNA may be transferred into a cell or organism by transfecting the cell or organism with the isolated guide RNA or plasmid DNA comprising a sequence coding for the guide RNA and a promoter.

Alternatively, the guide RNA may be transferred into a cell or organism using virus-mediated gene delivery.

When the guide RNA is transfected in the form of an isolated RNA into a cell or organism, the guide RNA may be prepared by in vitro transcription using any in vitro transcription system known in the art. The guide RNA is preferably transferred to a cell in the form of isolated RNA rather than in the form of plasmid comprising encoding sequence for a guide RNA. As used herein, the term “isolated RNA” may be interchangeable to “naked RNA”. This is cost—and time-saving because it does not require a step of cloning. However, the use of plasmid DNA or virus-mediated gene delivery for transfection of the guide RNA is not excluded.

The present RGEN composition comprising Cas protein or Cas protein-encoding nucleic acid and a guide RNA can specifically cleave a target DNA due to a specificity of the guide RNA for a target and an endonuclease or nickase activity of Cas protein.

As used herein, the term “cleavage” refers to the breakage of the covalent backbone of a nucleotide molecule.

In the present invention, a guide RNA may be prepared to be specific for any target which is to be cleaved. Therefore, the present RGEN composition can cleave any target DNA by manipulating or genotyping the target-specific portion of the guide RNA.

The guide RNA and the Cas protein may function as a pair. As used herein, the term "paired Cas nickase" may refer to the guide RNA and the Cas protein functioning as a pair. The pair comprises two guide RNAs. The guide RNA and Cas protein may function as a pair, and induce two nicks on different DNA strand. The two nicks may be separated by at least 100 bps, but are not limited thereto.

In the Example, the present inventors confirmed that paired Cas nickase allow targeted mutagenesis and large deletions of up to 1-kbp chromosomal segments in human cells. Importantly, paired nickases did not induce indels at off-target sites at which their corresponding nucleases induce mutations. Furthermore, unlike nucleases, paired nickases did not promote unwanted translocations associated with off-target DNA cleavages. In principle, paired nickases double the specificity of Cas9-mediated mutagenesis and will broaden the utility of RNA-guided enzymes in applications that require precise genome editing such as gene and cell therapy.

In the present invention, the composition may be used in the genotyping of a genome in the eukaryotic cells or organisms *in vitro*.

In one specific embodiment, the guide RNA may comprise the nucleotide sequence of Seq ID. No. 1, wherein the portion of nucleotide position 3~22 is a target-specific portion and thus, the sequence of this portion may be changed depending on a target.

As used herein, a eukaryotic cell or organism may be yeast, fungus, protozoa, plant, higher plant, and insect, or amphibian cells, or mammalian cells such as CHO, HeLa, HEK293, and COS-1, for example, cultured cells (*in vitro*), graft cells and primary cell culture (*in vitro* and *ex vivo*), and *in vivo* cells, and also mammalian cells including human, which are commonly used in the art, without limitation.

In one specific embodiment, it was found that Cas9 protein/single-chain guide RNA could generate site-specific DNA double-strand breaks *in vitro* and in mammalian cells, whose spontaneous repair induced targeted genome mutations at high frequencies.

Moreover, it was found that gene-knockout mice could be induced by the injection of Cas9 protein/guide RNA complexes or Cas9 mRNA/guide RNA into one-cell stage embryo and germ-line transmittable mutations could be generated by Cas9/guide RNA system.

Using Cas protein rather than a nucleic acid encoding Cas protein to induce a targeted mutagenesis is advantageous because exogenous DNA is not introduced into an organism. Thus, the composition comprising Cas protein and a guide RNA may be used to develop therapeutics or value-added crops, livestock, poultry, fish, pets, etc.

In accordance with another aspect of the invention, the present invention provides a composition for inducing targeted mutagenesis in eukaryotic cells or organisms, comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein. In addition, the present invention provides a use of the composition for inducing targeted mutagenesis in eukaryotic cells or organisms, comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

A guide RNA, Cas protein-encoding nucleic acid or Cas protein are as described in the above.

In accordance with another aspect of the invention, the present invention provides a kit for cleaving a target DNA or inducing targeted mutagenesis in eukaryotic cells or organisms comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

A guide RNA, Cas protein-encoding nucleic acid or Cas protein are as described in the above.

The kit may comprise a guide RNA and Cas protein-encoding nucleic acid or Cas protein as separate components or as one composition.

The present kit may comprise some additional components necessary for transferring the guide RNA and Cas component to a cell or an organism. For example, the kit may comprise an injection buffer such as DEPC-treated injection buffer, and materials necessary for analysis of mutation of a target DNA, but are not limited thereto.

In accordance with another aspect, the present invention provides a method for preparing a eukaryotic cell or organism comprising Cas protein and a guide RNA comprising a step of co-transfecting or serial-transfecting the eukaryotic cell or organism with a Cas protein-encoding nucleic acid or Cas protein, and a guide RNA or DNA that encodes the guide RNA.

A guide RNA, Cas protein-encoding nucleic acid or Cas protein are as described in the above.

In the present invention, a Cas protein-encoding nucleic acid or Cas protein and a guide RNA or DNA that encodes the guide RNA may be transferred into a cell by various methods known in the art, such as microinjection, electroporation, DEAE-dextran treatment, lipofection, nanoparticle-mediated transfection, protein transduction domain mediated transduction, virus-mediated gene delivery, and PEG-mediated transfection in protoplast, and so on, but are not limited thereto. Also, a Cas protein encoding nucleic acid or Cas protein and a guide RNA may be transferred into an organism by various method known in the art to administer a gene or a protein such as injection. A Cas protein-encoding nucleic acid or Cas protein may be transferred into a cell in the form of complex with a guide RNA, or separately. Cas protein fused to a protein transduction domain such as Tat can also be delivered efficiently into cells.

Preferably, the eukaryotic cell or organism is co-transfected or serial-transfected with a Cas9 protein and a guide RNA.

The serial-transfection may be performed by transfection with Cas protein-encoding nucleic acid first, followed by second transfection with naked guide RNA. Preferably, the second transfection is after 3, 6, 12, 18, 24 hours, but it is not limited thereto.

In accordance with another aspect, the present invention provides a eukaryotic cell or organism comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

The eukaryotic cells or organisms may be prepared by transferring the composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein into the cell or organism.

The eukaryotic cell may be yeast, fungus, protozoa, higher plant, and insect, or amphibian cells, or mammalian cells such as CHO, HeLa, HEK293, and COS-1, for example, cultured cells (*in vitro*), graft cells and primary cell culture (*in vitro* and *ex vivo*), and *in vivo* cells, and also mammalian cells including human, which are commonly

used in the art, without limitation. Further the organism may be yeast, fungus, protozoa, plant, higher plant, insect, amphibian, or mammal.

In accordance with another aspect of the invention, the present invention provides a method for cleaving a target DNA or inducing targeted mutagenesis in eukaryotic cells or organisms, comprising a step of treating a cell or organism comprising a target DNA with a composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

The step of treating a cell or organism with the composition may be performed by transferring the present composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein into the cell or organism.

As described in the above, such transfer may be performed by microinjection, transfection, electroporation, and so on.

In accordance with another aspect of the invention, the present invention provides an embryo comprising a genome edited by the present RGEN composition comprising a guide RNA specific for target DNA or DNA that encodes the guide RNA, and Cas protein-encoding nucleic acid or Cas protein.

Any embryo can be used in the present invention, and for the present invention, the embryo may be an embryo of a mouse. The embryo may be produced by injecting PMSG (Pregnant Mare Serum Gonadotropin) and hCG (human Chorionic Gonadotropin) into a female mouse of 4 to 7 weeks and the super-ovulated female mouse may be mated to males, and the fertilized embryos may be collected from oviducts.

The present RGEN composition introduced into an embryo can cleave a target DNA complementary to the guide RNA by the action of Cas protein and cause a mutation in the target DNA. Thus, the embryo into which the present RGEN composition has been introduced has an edited genome.

In one specific embodiment, it was found that the present RGEN composition could cause a mutation in a mouse embryo and the mutation could be transmitted to offspring.

A method for introducing the RGEN composition into the embryo may be any method known in the art, such as microinjection, stem cell insertion, retrovirus insertion, and so on. Preferably, a microinjection technique can be used.

In accordance with another aspect, the present invention provides a genome-modified animal obtained by transferring the embryo comprising a genome edited by the present RGEN composition into the oviducts of an animal.

In the present invention, the term "genome-modified animal" refers to an animal of which genome has been modified in the stage of embryo by the present RGEN composition and the type of the animal is not limited.

The genome-modified animal has mutations caused by a targeted mutagenesis based on the present RGEN composition. The mutations may be any one of deletion, insertion, translocation, inversion. The site of mutation depends on the sequence of guide RNA of the RGEN composition.

The genome-modified animal having a mutation of a gene may be used to determine the function of the gene.

In accordance with another aspect of the invention, the present invention provides a method of preparing a genome-modified animal comprising a step of introducing the present RGEN composition comprising a guide RNA specific for the target DNA or DNA that encodes the guide RNA and Cas protein-encoding nucleic acid or Cas protein into an embryo

of an animal; and a step of transferring the embryo into a oviduct of pseudopregnant foster mother to produce a genome-modified animal.

The step of introducing the present RGEN composition may be accomplished by any method known in the art such as microinjection, stem cell insertion, retroviral insertion, and so on.

In accordance with another aspect of the invention, the present invention provides a plant regenerated from the genome-modified protoplasts prepared by the method for eukaryotic cells comprising the RGEN composition.

In accordance with another aspect of the invention, the present invention provides a composition for genotyping mutations or variations in an isolated biological sample, comprising a guide RNA specific for the target DNA sequence Cas protein. In addition, the present invention provides a composition for genotyping nucleic acid sequences in pathogenic microorganisms in an isolated biological sample, comprising a guide RNA specific for the target DNA sequence and Cas protein.

A guide RNA, Cas protein-encoding nucleic acid or Cas protein are as described in the above.

As used herein the term "genotyping" refers to the "Restriction fragment length polymorphism (RFLP) assay".

RFLP may be used in 1) the detection of indel in cells or organisms induced by the engineered nucleases, 2) the genotyping naturally-occurring mutations or variations in cells or organisms, or 3) the genotyping the DNA of infected pathogenic microorganisms including virus or bacteria, etc.

The mutations or variation may be induced by engineered nucleases in cells.

The engineered nuclease may be a Zinc Finger Nuclease (ZFNs), Transcription Activator-Like Effector Nucleases (TALENs), or RGENs, but it is not limited thereto.

As used herein the term "biological sample" includes samples for analysis, such as tissues, cells, whole blood, serum, plasma, saliva, sputum, cerebrospinal fluid or urine, but is not limited thereto.

The mutations or variation may be a naturally-occurring mutations or variations.

The mutations or variations are induced by the pathogenic microorganisms. Namely, the mutations or variations occur due to the infection of pathogenic microorganisms, when the pathogenic microorganisms are detected, the biological sample is identified as infected.

The pathogenic microorganisms may be virus or bacteria, but are not limited thereto.

Engineered nuclease-induced mutations are detected by various methods, which include mismatch-sensitive Surveyor or T7 endonuclease I (T7E1) assays, RFLP analysis, fluorescent PCR, DNA melting analysis, and Sanger and deep sequencing. The T7E1 and Surveyor assays are widely used but often underestimate mutation frequencies because the assays detect heteroduplexes (formed by the hybridization of mutant and wild-type sequences or two different mutant sequences); they fail to detect homoduplexes formed by the hybridization of two identical mutant sequences. Thus, these assays cannot distinguish homozygous biallelic mutant clones from wild-type cells nor heterozygous biallelic mutants from heterozygous monoallelic mutants (FIG. 22). In addition, sequence polymorphisms near the nuclease target site can produce confounding results because the enzymes can cleave heteroduplexes formed by hybridization of these different wild-type alleles. RFLP analysis is free of these limitations and therefore is a method of choice. Indeed, RFLP analysis was one of the first methods used to detect

engineered nuclease-mediated mutations. Unfortunately, however, it is limited by the availability of appropriate restriction sites.

In accordance with another aspect of the invention, the present invention provides a kit for genotyping mutations or variations in an isolated biological sample, comprising the composition for genotyping mutations or variations in an isolated biological sample. In addition, the present invention provides a kit for genotyping nucleic acid sequences in pathogenic microorganisms in an isolated biological sample, comprising a guide RNA specific for the target DNA sequence and Cas protein.

A guide RNA, Cas protein-encoding nucleic acid or Cas protein are as described in the above.

In accordance with another aspect of the invention, the present invention provides a method of genotyping mutations or variations in an isolated biological sample, using the composition for genotyping mutations or variations in an isolated biological sample. In addition, the present invention provides a method of genotyping nucleic acid sequences in pathogenic microorganisms in an isolated biological sample, comprising a guide RNA specific for the target DNA sequence and Cas protein.

A guide RNA, Cas protein-encoding nucleic acid or Cas protein are as described in the above.

Mode for the Invention

Hereinafter, the present invention will be described in more detail with reference to Examples. However, these Examples are for illustrative purposes only, and the invention is not intended to be limited by these Examples.

EXAMPLE 1: GENOME EDITING ASSAY

1-1. DNA Cleavage Activity of Cas9 Protein

Firstly, the DNA cleavage activity of Cas9 derived from *Streptococcus pyogenes* in the presence or absence of a chimeric guide RNA in vitro was tested.

To this end, recombinant Cas9 protein that was expressed in and purified from *E. coli* was used to cleave a predigested or circular plasmid DNA that contained the 23-base pair (bp) human CCR5 target sequence. A Cas9 target sequence consists of a 20-bp DNA sequence complementary to crRNA or a chimeric guide RNA and the trinucleotide (5'-NGG-3') protospacer adjacent motif (PAM) recognized by Cas9 itself (FIG. 1A). Specifically, the Cas9-coding sequence (4,104 bp), derived from *Streptococcus pyogenes* strain MI GAS (NC_002737.1), was reconstituted using the human codon usage table and synthesized using oligonucleotides. First, 1-kb DNA segments were assembled using overlapping ~35-mer oligonucleotides and Phusion™ polymerase (New England Biolabs) and cloned into T-vector (SolGent). A full-length Cas9 sequence was assembled using four 1-kbp DNA segments by overlap PCR. The Cas9-encoding DNA segment was subcloned into p3s, which was derived from pcDNA3.1 (Invitrogen). In this vector, a peptide tag (NH₂-GGSGPPKKRKRKVPYDVPDYA-COOH, SEQ ID NO: 2) containing the HA epitope and a nuclear localization signal (NLS) was added to the C-terminus of Cas9. Expression and nuclear localization of the Cas9 protein in HEK 293T cells were confirmed by western blotting using anti-HA antibody (Santa Cruz).

Then, the Cas9 cassette was subcloned into pET28-b(+) and transformed into BL21 (DE3). The expression of Cas9 was induced using 0.5 mM IPTG for 4 h at 25° C. The Cas9

protein containing the His6-tag at the C terminus was purified using Ni-NTA agarose resin (Qiagen) and dialyzed against 20 mM HEPES (pH 7.5), 150 mM KCl, 1 mM DTT, and 10% glycerol (1). Purified Cas9 (50 nM) was incubated with super-coiled or pre-digested plasmid DNA (300 ng) and chimeric RNA (50 nM) in a reaction volume of 20 μl in NEB buffer 3 for 1 h at 37° C. Digested DNA was analyzed by electrophoresis using 0.8% agarose gels.

Cas9 cleaved the plasmid DNA efficiently at the expected position only in the presence of the synthetic RNA and did not cleave a control plasmid that lacked the target sequence (FIG. 1B).

1-2. DNA Cleavage by Cas9/Guide RNA Complex in Human Cells

A RFP-GFP reporter was used to investigate whether the Cas9/guide RNA complex can cleave the target sequence incorporated between the RFP and GFP sequences in mammalian cells.

In this reporter, the GFP sequence is fused to the RFP sequence out-of-frame (2). The active GFP is expressed only when the target sequence is cleaved by site-specific nucleases, which causes frameshifting small insertions or deletions (indels) around the target sequence via error-prone non-homologous end-joining (NHEJ) repair of the double-strand break (DSB) (FIG. 2).

The RFP-GFP reporter plasmids used in this study were constructed as described previously (2). Oligonucleotides corresponding to target sites (Table 1) were synthesized (Macrogen) and annealed. The annealed oligonucleotides were ligated into a reporter vector digested with EcoRI and BamHI.

HEK 293T cells were co-transfected with Cas9-encoding plasmid (0.8 μg) and the RFP-GFP reporter plasmid (0.2 μg) in a 24-well plate using Lipofectamine™ 2000 (Invitrogen).

Meanwhile, the in vitro transcribed chimeric RNA had been prepared as follows. RNA was in vitro transcribed through run-off reactions using the MEGAscript™ T7 kit (Ambion) according to the manufacturer's manual. Templates for RNA in vitro transcription were generated by annealing two complementary single strand DNAs or by PCR amplification (Table 1). Transcribed RNA was resolved on a 8% denaturing urea-PAGE gel. The gel slice containing RNA was cut out and transferred to probe elution buffer. RNA was recovered in nuclease-free water followed by phenol: chloroform extraction, chloroform extraction, and ethanol precipitation. Purified RNAs were quantified by spectrometry.

At 12 h post transfection, chimeric RNA (1 μg) prepared by in vitro transcription was transfected using Lipofectamine 2000.

At 3 d post-transfection, transfected cells were subjected to flow cytometry and cells expressing both RFP and GFP were counted.

It was found that GFP-expressing cells were obtained only when the cells were transfected first with the Cas9 plasmid and then with the guide RNA 12 h later (FIG. 2), demonstrating that RGENs could recognize and cleave the target DNA sequence in cultured human cells. Thus GFP-expressing cells were obtained by serial-transfection of the Cas9 plasmid and the guide RNA rather than co-transfection.

TABLE 1

Gene		sequence (5' to 3')	SEQ ID NO.
Oligonucleotides used for the construction of the reporter plasmid			
CCR5	F	AATTCATGACATCAATTATTATACATCGGAGGAG	3
	R	GATCCTCCTCCGATGTATAATAATTGATGTCATG	4
Primers used in the T7E1 assay			
CCR5	F1	CTCCATGGTGTATAGAGCA	5
	F2	GAGCCAAGCTCTCCATCTAGT	6
	R	GCCCTGTCAAGAGTTGACAC	7
C4BPB	F1	TATTGGCTGGTTGAAAGGG	8
	R1	AAAGTCATGAAATAACACACCCCA	9
	F2	CTGCATTGATATGGTAGTACCATG	10
	R2	GCTGTTTCATTGCAATGGAATG	11
Primers used for the amplification of off-target sites			
ADCY5	F1	GCTCCCACCTTAGTGCTCTG	12
	R1	GGTGGCAGGAACCTGTATGT	13
	F2	GTCAATTGGCCAGAGATGTGGA	14
	R2	GTCCCATGACAGGCGGTGTAT	15
	F	GCCTGGCCAAGTTTCAGTTA	16
KCNJ6	R1	TGGAGCCATTGGTTTGCATC	17
	R2	CCAGAACTAAGCCGTTTCTGAC	18
	F1	ATCACCGACAACCAAGTTTCC	19
CNTNAP2	F2	TGCAGTGCAGACTCTTCCA	20
	R	AAGGACACAGGGCAACTGAA	21
N/A Chr. 5	F1	TGTGGAACGAGTGGTGACAG	22
	R1	GCTGGATTAGGAGGCAGGATTC	23
	F2	GTGCTGAGAACGCTTCATAGAG	24
	R2	GGACCAAACACATTCTTCTCAC	25
Primers used for the detection of chromosomal deletions			
Deletion	F	CCACATCTCGTTCTCGGTTT	26
	R	TCACAAGCCCACAGATATTT	27

1-3. Targeted Disruption of Endogenous Genes in Mammalian Cells by RGEN

To test whether RGENs could be used for targeted disruption of endogenous genes in mammalian cells, genomic DNA isolated from transfected cells using T7 endonuclease I (T7E1), a mismatch-sensitive endonuclease that specifically recognizes and cleaves heteroduplexes formed by the hybridization of wild-type and mutant DNA sequences was analyzed (3).

To introduce DSBs in mammalian cells using RGENs, 2×10^6 K562 cells were transfected with 20 μ g of Cas9-encoding plasmid using the 4D-Nucleofector™, SF Cell Line 4D-Nucleofector X Kit, Program FF-120 (Lonza) according to the manufacturer's protocol. For this experiment, K562 (ATCC, CCL-243) cells were grown in RPMI-1640 with 10% FBS and the penicillin/streptomycin mix (100 U/ml and 100 μ g/ml, respectively).

After 24 h, 10-40 μ g of in vitro transcribed chimeric RNA was nucleofected into 1×10^6 K562 cells. The in vitro transcribed chimeric RNA had been prepared as described in the Example 1-2.

Cells were collected two days after RNA transfection and genomic DNA was isolated. The region including the target site was PCR-amplified using the primers described in Table 1. The amplicons were subjected to the T7E1 assay as described previously (3). For sequencing analysis, PCR products corresponding to genomic modifications were purified and cloned into the T-Blunt vector using the T-Blunt PCR Cloning Kit (SolGent). Cloned products were sequenced using the M13 primer.

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It was found that mutations were induced only when the cells were transfected serially with Cas9-encoding plasmid and then with guide RNA (FIG. 3). Mutation frequencies (Indels (%) in FIG. 3A) estimated from the relative DNA band intensities were RNA-dosage dependent, ranging from 1.3% to 5.1%. DNA sequencing analysis of the PCR amplicons corroborated the induction of RGEN-mediated mutations at the endogenous sites. Indels and microhomologies, characteristic of error-prone NHEJ, were observed at the target site. The mutation frequency measured by direct sequencing was 7.3% (=7 mutant clones/96 clones), on par with those obtained with zinc finger nucleases (ZFNs) or transcription-activator-like effector nucleases (TALENs).

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Serial-transfection of Cas9 plasmid and guide RNA was required to induce mutations in cells. But when plasmids that encode guide RNA, serial transfection was unnecessary and cells were co-transfected with Cas9 plasmid and guide RNA-encoding plasmid.

In the meantime, both ZFNs and TALENs have been successfully developed to disrupt the human CCR5 gene (3-6), which encodes a G-protein-coupled chemokine receptor, an essential co-receptor of HIV infection. A CCR5-specific ZEN is now under clinical investigation in the US for the treatment of AIDS (7). These ZFNs and TALENs, however, have off-target effects, inducing both local mutations at sites whose sequences are homologous to the on-target sequence (6, 8-10) and genome rearrangements that arise from the repair of two concurrent DSBs induced at on-target and off-target sites (11-12). The most striking off-target sites associated with these CCR5-specific engineered nucleases reside in the CCR2 locus, a close homolog of CCR5, located 15-kbp upstream of CCR5. To avoid

off-target mutations in the CCR2 gene and unwanted deletions, inversions, and duplications of the 15-kbp chromosomal segment between the CCR5 on-target and CCR2 off-target sites, the present inventors intentionally chose the target site of our CCR5-specific RGEN to recognize a region within the CCR5 sequence that has no apparent homology with the CCR2 sequence.

The present inventors investigated whether the CCR5-specific RGEN had off-target effects. To this end, we searched for potential off-target sites in the human genome by identifying sites that are most homologous to the intended 23-bp target sequence. As expected, no such sites were found in the CCR2 gene. Instead, four sites, each of which carries 3-base mismatches with the on-target site, were found (FIG. 4A). The T7E1 assays showed that mutations were not detected at these sites (assay sensitivity, ~0.5%), demonstrating exquisite specificities of RGENS (FIG. 4B). Furthermore, PCR was used to detect the induction of chromosomal deletions in cells separately transfected with plasmids encoding the ZEN and RGEN specific to CCR5. Whereas the ZFN induced deletions, the RGEN did not (FIG. 4C).

Next, RGENs was reprogrammed by replacing the CCR5-specific guide RNA with a newly-synthesized RNA designed to target the human C4BPB gene, which encodes the beta chain of C4b-binding protein, a transcription factor. This RGEN induced mutations at the chromosomal target site in K562 cells at high frequencies (FIG. 3B). Mutation frequencies measured by the T7E1 assay and by direct sequencing were 14% and 8.3% (=4 mutant clones/48 clones), respectively. Out of four mutant sequences, two clones contained a single-base or two-base insertion precisely at the cleavage site, a pattern that was also observed at the CCR5 target site. These results indicate that RGENs cleave chromosomal target DNA at expected positions in cells.

EXAMPLE 2: PROTEINACEOUS RGEN-MEDIATED GENOME EDITING

RGENs can be delivered into cells in many different forms. RGENs consist of Cas9 protein, crRNA, and tracrRNA. The two RNAs can be fused to form a single-chain guide RNA (sgRNA). A plasmid that encodes Cas9 under a promoter such as CMV or CAG can be transfected into cells. crRNA, tracrRNA, or sgRNA can also be expressed in cells using plasmids that encode these RNAs. Use of plasmids, however, often results in integration of the whole or part of the plasmids in the host genome. The

bacterial sequences incorporated in plasmid DNA can cause unwanted immune response in vivo. Cells transfected with plasmid for cell therapy or animals and plants derived from DNA-transfected cells must go through a costly and lengthy regulation procedure before market approval in most developed countries. Furthermore, plasmid DNA can persist in cells for several days post-transfection, aggravating off-target effects of RGENS.

Here, we used recombinant Cas9 protein complexed with in vitro transcribed guide RNA to induce targeted disruption of endogenous genes in human cells. Recombinant Cas9 protein fused with the hexa-histidine tag was expressed in and purified from *E. coli* using standard Ni ion affinity chromatography and gel filtration. Purified recombinant Cas9 protein was concentrated in storage buffer (20 mM HEPES pH 7.5, 150 mM KCl, 1 mM DTT, and 10% glycerol). Cas9 protein/sgRNA complex was introduced directly into K562 cells by nucleofection: 1×10^6 K562 cells were transfected with 22.5-225 (1.4-14 μ M) of Cas9 protein mixed with 100 μ g (29 μ M) of in vitro transcribed sgRNA (or crRNA 40 μ g and tracrRNA 80 μ g) in 100 μ l solution using the 4D-Nucleofector, SF Cell Line 4D-Nucleofector X Kit, Program FF-120 (Lonza) according to the manufacturer's protocol. After nucleofection, cells were placed in growth media in 6-well plates and incubated for 48 hr. When 2×10^5 K562 cells were transfected with $\frac{1}{5}$ scale-downed protocol, 4.5-45 μ g of Cas9 protein mixed with 6-60 μ g of in vitro transcribed sgRNA (or crRNA 8 μ g and tracrRNA 16 μ g) were used and nucleofected in 20 μ l solution. Nucleofected cells were then placed in growth media in 48-well plates. After 48 hr, cells were collected and genomic DNA was isolated. The genomic DNA region spanning the target site was PCR-amplified and subjected to the T7E1 assay.

As shown in FIG. 10, Cas9 protein/sgRNA complex induced targeted mutation at the CCR5 locus at frequencies that ranged from 4.8 to 38% in a sgRNA or Cas9 protein dose-dependent manner, on par with the frequency obtained with Cas9 plasmid transfection (45%). Cas9 protein/crRNA/tracrRNA complex was able to induce mutations at a frequency of 9.4%. Cas9 protein alone failed to induce mutations. When 2×10^5 cells were transfected with $\frac{1}{5}$ scale-downed doses of Cas9 protein and sgRNA, mutation frequencies at the CCR5 locus ranged from 2.7 to 57% in a dose-dependent manner, greater than that obtained with co-transfection of Cas9 plasmid and sgRNA plasmid (32%).

We also tested Cas9 protein/sgRNA complex that targets the ABCC11 gene and found that this complex induced indels at a frequency of 35%, demonstrating general utility of this method.

TABLE 2

Sequences of guide RNA				
Target RNA	type	RNA sequence (5' to 3')	Length	SEQ ID NO
CCR5	sgRNA	GGUGACAUCAAUUUUUAUCAUGUUUUAGAGCUAG AAAUAGCAAGUUAAAAUAAGGCCUAGUCCGUUAUCA ACUUGAAAAAGUGGCACCGAGUCGGUGUCUUUUUUU	104 bp	28
	crRNA	GGUGACAUCAAUUUUUAUCAUGUUUUAGAGCUAU GCUGUUUUUG	44 bp	29
	tracrRNA	GGAACCAUUCAAACAGCAUAGCAAGUUAAAAUAA GGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCG AGUCGGUGUCUUUUUUU	86 bp	30

EXAMPLE 3: RNA-GUIDED GENOME EDITING IN MICE

To examine the gene-targeting potential of RGENs in pronuclear (PN)-stage mouse embryos, the forkhead box N1 (Foxn1) gene, which is important for thymus development and keratinocyte differentiation (Nehls et al., 1996), and the protein kinase, DNA activated, catalytic polypeptide (Prkdc) gene, which encodes an enzyme critical for DNA DSB repair and recombination (Taccioli et al., 1998) were used.

To evaluate the genome-editing activity of the Foxn1-RGEN, we injected Cas9 mRNA (10-ng/ μ l solution) with various doses of the sgRNA (FIG. 5a) into the cytoplasm of PN-stage mouse embryos, and conducted T7 endonuclease I (T7E1) assays (Kim et al. 2009) using genomic DNAs obtained from in vitro cultivated embryos (FIG. 6a).

Alternatively, we directly injected the RGEN in the form of recombinant Cas9 protein (0.3 to 30 ng/ μ l) complexed with the two-fold molar excess of Foxn1-specific sgRNA (0.14 to 14 ng/ μ l) into the cytoplasm or pronucleus of one-cell mouse embryos, and analyzed mutations in the Foxn1 gene using in vitro cultivated embryos (FIG. 7).

Specifically, Cas9 mRNA and sgRNAs were synthesized in vitro from linear DNA templates using the mMACHINE mMACHINE T7 Ultra kit (Ambion) and MEGAShortscript T7 kit (Ambion), respectively, according to the manufacturers' instructions, and were diluted with appropriate amounts of diethyl pyrocarbonate (DEPC, Sigma)-treated injection buffer (0.25 mM EDTA, 10 mM Tris, pH 7.4). Templates for sgRNA synthesis were generated using oligonucleotides listed in Table 3. Recombinant Cas9 protein was obtained from ToolGen, Inc.

TABLE 3

RNA Name	Direction	Sequence (5' to 3')	SEQ ID NO
Foxn1 #1 sgRNA	F	<u>GAAATTAATACGACTCACTATAGGCAGTCTGACG</u> <u>TCACACTTCGGTTTTAGAGCTAGAAATAGCAAGT</u> TAAAATAAGGCTAGTCCG	31
Foxn1 #2 sgRNA	F	<u>GAAATTAATACGACTCACTATAGGACTTCCAGGC</u> <u>TCCACCCGACGTTTTAGAGCTAGAAATAGCAAGT</u> TAAAATAAGGCTAGTCCG	32
Foxn1 #3 sgRNA	F	<u>GAAATTAATACGACTCACTATAGGCCAGGCTCCA</u> <u>CCCGACTGGAGTTTTAGAGCTAGAAATAGCAAGT</u> TAAAATAAGGCTAGTCCG	33
Foxn1 #4 sgRNA	F	<u>GAAATTAATACGACTCACTATAGGACTGGAGGGC</u> <u>GAAACCCCAAGGTTTTAGAGCTAGAAATAGCAAGT</u> TAAAATAAGGCTAGTCCG	34
Foxn1 #5 sgRNA	F	<u>GAAATTAATACGACTCACTATAGGACCCCAAGGG</u> <u>GACCTCATCGTTTTAGAGCTAGAAATAGCAAGT</u> TAAAATAAGGCTAGTCCG	35
Prkdc #1 sgRNA	F	<u>GAAATTAATACGACTCACTATAGGTTAGTTTTTT</u> <u>CCAGAGACTTGTGTTTTAGAGCTAGAAATAGCAAGT</u> TAAAATAAGGCTAGTCCG	36
Prkdc #2 sgRNA	F	<u>GAAATTAATACGACTCACTATAGGTTGGTTTGCT</u> <u>TGTGTTATCGTTTTAGAGCTAGAAATAGCAAGT</u> TAAAATAAGGCTAGTCCG	37
Prkdc #3 sgRNA	F	<u>GAAATTAATACGACTCACTATAGGCACAAGCAAA</u> <u>CCAAAGTCTCGTTTTAGAGCTAGAAATAGCAAGT</u> TAAAATAAGGCTAGTCCG	38
Prkdc #4 sgRNA	F	<u>GAAATTAATACGACTCACTATAGGCCTCAATGCT</u> <u>AAGCGACTTCGTTTTAGAGCTAGAAATAGCAAGT</u> TAAAATAAGGCTAGTCCG	39

All animal experiments were performed in accordance with the Korean Food and Drug Administration (KFDA) guidelines. Protocols were reviewed and approved by the Institutional Animal Care and Use Committees (IACUC) of the Laboratory Animal Research Center at Yonsei University (Permit Number: 2013-0099). All mice were maintained in the specific pathogen-free facility of the Yonsei Laboratory Animal Research Center. FVB/NTac (Taconic) and ICR mouse strains were used as embryo donors and foster mothers, respectively. Female FVB/NTac mice (7-8 weeks old) were super-ovulated by intra-peritoneal injections of 5 IU pregnant mare serum gonadotropin (PMSG, Sigma) and 5 IU human chorionic gonadotropin (hCG, Sigma) at 48-hour intervals. The super-ovulated female mice were mated to FVB/NTac stud males, and fertilized embryos were collected from oviducts.

Cas9 mRNA and sgRNAs in M2 medium (Sigma) were injected into the cytoplasm of fertilized eggs with well-recognized pronuclei using a Piezo-driven micromanipulator (Prime Tech).

In the case of injection of recombinant Cas9 protein, the recombinant Cas9 protein: Foxn1-sgRNA complex was diluted with DEPC-treated injection buffer (0.25 mM EDTA, 10 mM Tris, pH 7.4) and injected into male pronuclei using a TransferMan NK2 micromanipulator and a Femto-Jet® microinjector (Eppendorf).

The manipulated embryos were transferred into the oviducts of pseudopregnant foster mothers to produce live animals, or were cultivated in vitro for further analyses.

To screen F0 mice and in vitro cultivated mouse embryos with RGEN-induced mutations, T7E1 assays were performed as previously described using genomic DNA samples from tail biopsies and lysates of whole embryos (Cho et al., 2013).

Briefly, the genomic region encompassing the RGEN target site was PCR-amplified, melted, and re-annealed to form heteroduplex DNA, which was treated with T7 endonuclease 1 (New England Biolabs), and then analyzed by agarose gel electrophoresis. Potential off-target sites were identified by searching with bowtie 0.12.9 and were also similarly monitored by T7E1 assays. The primer pairs used in these assays were listed in Tables 4 and 5.

TABLE 4

Primers used in the T7E1 assay			
Gene	Direction	Sequence (5' to 3')	SEQ ID NO
Foxn1F1		GTCTGTCTATCATCTCTTCCCTTCTCTCC	40
	F2	TCCTAATCCGATGGCTAGCTCCAG	41
	R1	ACGAGCAGCTGAAGTTAGCATGC	42
	R2	CTACTCAATGCTCTTAGAGCTACCAGGC TTGC	43
PrkdcF		GACTGTTGTGGGGAGGGCCG	44
	F2	GGGAGGGCCGAAAGCTTATTTTG	45
	R1	CCTGAAGACTGAAGTTGGCAGAAGTGAG	46
	R2	CTTTAGGGCTTCTTCTACAATCACG	47

TABLE 5

Primers used for amplification of off-target sites				
Gene	Notation	Direction	Sequence (5' to 3')	SEQ ID NO
Foxn1	off 1	F	CTCGGTGTGTAGCCCTGAC	48
		R	AGACTGGCCTGGAACCTCACAG	49
	off 2	F	CACTAAAGCCTGTGAGGAAGCCG	50
		R	CTGTGGAGAGCACACAGCAGC	51
	off 3	F	GCTGCGACCTGAGACCATG	52
		R	CTTCAATGGCTTCTGCTTAGGCTAC	53
	off 4	F	GGTTCAGATGAGGCCATCCTTTC	54
		R	CCTGATCTGCAGGCTTAACCCCTTG	55
Prkdc	off 1	F	CTCACCTGCACATCACATGTGG	56
		R	GGCATCCACCCTATGGGGTC	57
	off 2	F	GCCTTGACCTAGAGCTTAAAGAGCC	58
		R	GGTCTTGTGTAGCAGGAAGGACACTG	59
	off 3	F	AAAACCTCTGCTTGATGGGATATGTGGG	60
		R	CTCTCACTGGTTATCTGTGCTCCTTC	61
	off 4	F	GGATCAATAGGTGGTGGGGATG	62
		R	GTGAATGACACAATGTGACAGCTTCAG	63
	off 5	F	CACAAGACAGACCTCTCAACATTCAGTC	64
		R	GTGCATGCATATAATCCATTCTGATTGCTCTC	65
	off 6	F1	GGGAGGCAGAGGCAGGT	66
		F2	GGATCTCTGTGAGTTTGGAGGCCA	67
R1		GCTCCAGAACTCACTCTTAGGCTC	68	

Mutant founders identified by the T7E1 assay were further analyzed by fPCR. Appropriate regions of genomic DNA were sequenced as described previously (Sung et al., 2013). For routine PCR genotyping of F1 progenies, the following primer pairs were used for both wild-type and

mutant alleles: 5'-CTACTCCCTCCGCGAGTCTGA-3' (SEQ ID NO: 69) and 5'-CCAGGCCTAGGTTCCAGGTA-3' (SEQ ID NO: 70) for the Foxn1 gene, 5'-CCCCAGCAT-TGCAGATTTCC-3' (SEQ ID NO: 71) and 5'-AGGGCTTCTTCTCTACAATCACG-3' (SEQ ID NO: 72) for Prkdc gene.

In the case of injection of Cas9 mRNA, mutant fractions (the number of mutant embryos/the number of total embryos) were dose-dependent, ranging from 33% (1 ng/μl sgRNA) to 91% (100 ng/μl) (FIG. 6b). Sequence analysis confirmed mutations in the Foxn1 gene; most mutations were small deletions (FIG. 6c), reminiscent of those induced by ZFNs and TALENS (Kim et al., 2013).

In the case of injection of Cas9 protein, these injection doses and methods minimally affected the survival and development of mouse embryos in vitro: over 70% of RGEN-injected embryos hatched out normally in both experiments. Again, mutant fractions obtained with Cas9 protein injection were dose-dependent, and reached up to 88% at the highest dose via pronucleus injection and to 71% via intra-cytoplasmic injection (FIGS. 7a and 7b). Similar to the mutation patterns induced by Cas9 mRNA plus sgRNA (FIG. 6c), those induced by the Cas9 protein-sgRNA complex were mostly small deletions (FIG. 7c). These results clearly demonstrate that RGENs have high gene-targeting activity in mouse embryos.

Encouraged by the high mutant frequencies and low cytotoxicity induced by RGENs, we produced live animals

by transferring the mouse embryos into the oviducts of pseudo-pregnant foster mothers.

Notably, the birth rates were very high, ranging from 58% to 73%, and were not affected by the increasing doses of Foxn1-sgRNA (Table 6).

TABLE 6

RGEN-mediated gene-targeting in FVB/NTac mice						
Target Gene	Cas9 mRNA + sgRNA (ng/μl)	Injected embryos	Transferred embryos (%)	Total newborns (%)	Live newborns * (%)	Founders † (%)
Foxn1	10 + 1	76	62 (82)	45 (73)	31 (50)	12 (39)
	10 + 10	104	90 (87)	52 (58)	58 (64)	33 (57)
	10 + 100	100	90 (90)	62 (69)	58 (64)	54 (93)
Prkdc	Total	280	242 (86)	159 (66)	147 (61)	99 (67)
	50 + 50	73	58 (79)	35 (60)	33 (57)	11 (33)
	50 + 100	79	59 (75)	22 (37)	21 (36)	7 (33)
	50 + 250	94	73 (78)	37 (51)	37 (51)	21 (57)
	Total	246	190 (77)	94 (49)	91 (48)	39 (43)

Out of 147 newborns, we obtained 99 mutant founder mice. Consistent with the results observed in cultivated embryos (FIG. 6b), mutant fractions

were proportional to the doses of Foxn1-sgRNA, and reached up to 938 (100 ng/μl Foxn1-sgRNA) (Tables 6 and 7, FIG. 5b).

TABLE 7

DNA sequences of Foxn1 mutant alleles identified from a subset of T7E1-positive mutant founders			
ACTTCCAGGCTCCACCCGACTGGAGGGCGAACCCCAAGGGGACCTCATGCAGG (SEQ ID NO: 134)	del + ins	#	Founder mice
ACTTCCAGGC-----AACCCCAAGGGGACCTCATGCAGG Δ19 (SEQ ID NO: 297)		1	20
ACTTCCAGGC-----GAACCCCAAGGGGACCTCATGCAGG Δ18 (SEQ ID NO: 298)		1	115
ACTTCCAGGCTCC-----Δ60 (SEQ ID NO: 299)		1	19
ACTTCCAGGCTCC-----Δ44 (SEQ ID NO: 300)		1	108
ACTTCCAGGCTCC-----CAAGGGGACCTCATGCAGG Δ21 (SEQ ID NO: 3001)		1	64
ACTTCCAGGCTCC-----TTAGGAGCGAACCCCAAGGGGACCTCA Δ12 + 6 (SEQ ID NO: 302)		1	126
ACTTCCAGGCTCCACC-----TCATGCAGG Δ28 (SEQ ID NO: 303)		1	5
ACTTCCAGGCTCCACCC-----CCAAGGGACCTCATG Δ21 + 4 (SEQ ID NO: 304)		1	61
ACTTCCAGGCTCCACCC-----AAGGGGACCTCATGCAGG Δ18 (SEQ ID NO: 305)		2	95, 29
ACTTCCAGGCTCCACCC-----CAAGGGGACCTCATGCAGG Δ17 (SEQ ID NO: 306)		7	12, 14, 27, 66, 108, 114, 126
ACTTCCAGGCTCCACCC-----ACCCAAGGGGACCTCATGCAG Δ15 + 1 (SEQ ID NO: 307)		1	32
ACTTCCAGGCTCCACCC-----CACCCAAGGGGACCTCATGCA Δ15 + 2 (SEQ ID NO: 308)		1	124
ACTTCCAGGCTCCACCC-----ACCCCAAGGGGACCTCATGCAGG Δ13 (SEQ ID NO: 309)		1	32
ACTTCCAGGCTCCACCC-----GGCGAACCCCAAGGGGACCTCATGCAGG Δ8 (SEQ ID NO: 310)		1	110
ACTTCCAGGCTCCACCT-----GGGGACCTCATGCAGG Δ20 + 1 (SEQ ID NO: 311)		1	29

TABLE 7-continued

DNA sequences of Foxn1 mutant alleles identified from a subset of T7E1-positive mutant founders			
(SEQ ID NO: 134)	del + ins	#	Founder mice
ACTTCCAGGCTCCACCCGACTGGAGGGCGAACCCCAAGGGGACCTCATGCAGG (SEQ ID NO: 312)	-----AACCCCAAGGGGACCTCATGCAGGΔ11	1	111
ACTTCCAGGCTCCACCCGA-----ACCTCATGCAGGΔ22 (SEQ ID NO: 313)	-----ACCTCATGCAGGΔ22	1	79
ACTTCCAGGCTCCACCCGA-----GGGGACCTCATGCAGGΔ18 (SEQ ID NO: 314)	-----GGGGACCTCATGCAGGΔ18	2	13, 127
ACTTCCAGGCTCCACCCCA-----AGGGGACCTCATGCAGGΔ17 (SEQ ID NO: 315)	-----AGGGGACCTCATGCAGGΔ17	1	24
ACTTCCAGGCTCCACCCGA-----ACCCCAAGGGGACCTCATGCAGGΔ11 (SEQ ID NO: 316)	-----ACCCCAAGGGGACCTCATGCAGGΔ11	5	14, 53, 58, 69, 124
ACTTCCAGGCTCCACCCGA-----GACCCCAAGGGGACCTCATGCAGGΔ10 (SEQ ID NO: 317)	-----GACCCCAAGGGGACCTCATGCAGGΔ10	1	14
ACTTCCAGGCTCCACCCGA----GGGCGAACCCCAAGGGGACCTCATGCAGGΔ5 (SEQ ID NO: 318)	----GGGCGAACCCCAAGGGGACCTCATGCAGGΔ5	3	53, 79, 115
ACTTCCAGGCTCCACCCGAC-----CTCATGCAGGΔ23 (SEQ ID NO: 319)	-----CTCATGCAGGΔ23	1	108
ACTTCCAGGCTCCACCCGAC-----CCCAAGGGGACCTCATGCAGGΔ11 (SEQ ID NO: 320)	-----CCCAAGGGGACCTCATGCAGGΔ11	1	3
ACTTCCAGGCTCCACCCGAC-----GAAGGGCCCCAAGGGGACCTCAΔ11 + 6 (SEQ ID NO: 321)	-----GAAGGGCCCCAAGGGGACCTCAΔ11 + 6	1	66
ACTTCCAGGCTCCACCCGAC-----GAACCCCAAGGGGACCTCATGCAGGΔ8 (SEQ ID NO: 322)	-----GAACCCCAAGGGGACCTCATGCAGGΔ8	2	3, 66
ACTTCCAGGCTCCACCCGAC----GGCGAACCCCAAGGGGACCTCATGCAGGΔ5 (SEQ ID NO: 323)	----GGCGAACCCCAAGGGGACCTCATGCAGGΔ5	1	27
ACTTCCAGGCTCCACCCGAC--GTGCTTGAGGGCGAACCCCAAGGGGACCTCAΔ2 + 6 (SEQ ID NO: 324)	--GTGCTTGAGGGCGAACCCCAAGGGGACCTCAΔ2 + 6	2	5
ACTTCCAGGCTCCACCCGACT-----CACTATCTTCTGGGCTCCTCCATGTCΔ6 + 25 (SEQ ID NO: 325)	-----CACTATCTTCTGGGCTCCTCCATGTCΔ6 + 25	2	21, 114
ACTTCCAGGCTCCACCCGACT---TGGCGAACCCCAAGGGGACCTCATGCAGΔ4 + 1 (SEQ ID NO: 326)	---TGGCGAACCCCAAGGGGACCTCATGCAGΔ4 + 1	1	53
ACTTCCAGGCTCCACCCGACT--TGCAGGGCGAACCCCAAGGGGACCTCATGCΔ2 + 3 (SEQ ID NO: 327)	--TGCAGGGCGAACCCCAAGGGGACCTCATGCΔ2 + 3	1	126
ACTTCCAGGCTCCACCCGACTTGGAGGGCGAACCCCAAGGGGACCTCATGCAG + 1 (SEQ ID NO: 328)	TGGAGGGCGAACCCCAAGGGGACCTCATGCAG + 1	15	3, 5, 12, 19, 29, 55, 56, 61, 66, 68, 81, 108, 111, 124, 127
ACTTCCAGGCTCCACCCGACTTGGAGGGCGAACCCCAAGGGGACCTCATGCA + 2 (SEQ ID NO: 329)	TGGAGGGCGAACCCCAAGGGGACCTCATGCA + 2	2	79, 120
ACTTCCAGGCTCCACCCGACTGTTGGAGGGCGAACCCCAAGGGGACCTCATGC + 3 (SEQ ID NO: 330)	GTTGGAGGGCGAACCCCAAGGGGACCTCATGC + 3	1	55
ACTTCCAGGCTCCACCCGACTGGAG (+455) GGCGAACCCCAAGGGGACCTCC + 455 (SEQ ID NO: 331)	GGAG (+455) GGCGAACCCCAAGGGGACCTCC + 455	1	13

To generate Prkdc-targeted mice, we applied a 5-fold higher concentration of Cas9 mRNA (50 ng/μl) with increasing doses of Prkdc-sgRNA (50, 100, and 250 ng/μl). Again, the birth rates were very high, ranging from 51% to 60%, enough to produce a sufficient number of newborns for the

analysis (Table 6). The mutant fraction was 57% (21 mutant founders among 37 newborns) at the maximum dose of Prkdc-sgRNA. These birth rates obtained with RGENs were approximately 2-to 10-fold higher than those with TALENs reported in our previous study (Sung et al., 2013). These

results demonstrate that RGENs are potent gene-targeting reagents with minimal toxicity.

To test the germ-line transmission of the mutant alleles, we crossed the *Foxn1* mutant founder #108, a mosaic with four different alleles (FIG. 5c, and Table 8) with wild-type mice, and monitored the genotypes of F1 offspring.

TABLE 8

Genotypes of <i>Foxn1</i> mutant mice			
Founder NO.	sgRNA (ng/ml)	Genotyping Summary	Detected alleles
58*	1	not determined	$\Delta 11$
19	100	bi-allelic	$\Delta 60/+1$
20	100	bi-allelic	$\Delta 67/\Delta 19$
13	100	bi-allelic	$\Delta 18/+455$
32	10	bi-allelic (heterozygote)	$\Delta 13/\Delta 15+1$
115	10	bi-allelic (heterozygote)	$\Delta 18/\Delta 5$
111	10	bi-allelic (heterozygote)	$\Delta 11/+1$
110	10	bi-allelic (homozygote)	$\Delta 8/\Delta 8$
120	10	bi-allelic (homozygote)	$+2/+2$
81	100	heterozygote	$+1/WT$
69	100	homozygote	$\Delta 11/\Delta 11$
55	1	mosaic	$\Delta 18/\Delta 1/+1/+3$
56	1	mosaic	$\Delta 127/\Delta 41/\Delta 2/+1$
127	1	mosaic	$\Delta 18/+1/WT$
53	1	mosaic	$\Delta 11/\Delta 5/\Delta 4+1/WT$
27	10	mosaic	$\Delta 17/\Delta 5/WT$
29	10	mosaic	$\Delta 18/\Delta 20+1/+1$
95	10	mosaic	$\Delta 18/\Delta 14/\Delta 8/\Delta 4$
108	10	mosaic	$+1/\Delta 17/\Delta 23/\Delta 44$
114	10	mosaic	$\Delta 17/\Delta 8/\Delta 6+25$
124	10	mosaic	$\Delta 11/\Delta 15+2+1$
126	10	mosaic	$\Delta 17/\Delta 2+3/\Delta 12+6$
12	100	mosaic	$\Delta 30/\Delta 28/\Delta 17/+1$
5	100	mosaic	$\Delta 28/\Delta 11/\Delta 2+6/+1$
14	100	mosaic	$\Delta 17/\Delta 11/\Delta 10$
21	100	mosaic	$\Delta 127/\Delta 41/\Delta 2/\Delta 6+25$
24	100	mosaic	$\Delta 17/+1/WT$
64	100	mosaic	$\Delta 31/\Delta 21/+1/WT$
68	100	mosaic	$\Delta 17/\Delta 11/+1/WT$
79	100	mosaic	$\Delta 22/\Delta 5/+2/WT$
61	100	mosaic	$\Delta 21+4/\Delta 6/+1/+9$

TABLE 8-continued

Genotypes of <i>Foxn1</i> mutant mice			
Founder NO.	sgRNA (ng/ml)	Genotyping Summary	Detected alleles
66**	100	mosaic	$\Delta 17/\Delta 8/\Delta 11+6/+1/WT$
3	100	mosaic	$\Delta 11/\Delta 8/+1$

Underlined alleles were sequenced.

Alleles in red, detected by sequencing, but not by fPCR.

*only one clone sequenced.

**Not determined by fPCR.

As expected, all the progenies were heterozygous mutants possessing the wild-type allele and one of the mutant alleles (FIG. 5d). We also confirmed the germ-line transmission in independent founder mice of *Foxn1* (FIG. 8) and *Prkdc* (FIG. 9). To the best of our knowledge, these results provide the first evidence that RGEN-induced mutant alleles are stably transmitted to F1 progenies in animals.

EXAMPLE 4: RNA-GUIDED GENOME EDITING IN PLANTS

4-1. Production of Cas9 Protein

The Cas9 coding sequence (4104 bps), derived from *Streptococcus pyogenes* strain M1 GAS (NC_002737.1), was cloned to pET28-b(+) plasmid. A nuclear targeting sequence (NLS) was included at the protein N terminus to ensure the localization of the protein to the nucleus. pET28-b(+) plasmid containing Cas9 ORF was transformed into BL21 (DE3). Cas9 was then induced using 0.2 mM IPTG for 16 hrs at 18° C. and purified using Ni-NTA agarose beads (Qiagen) following the manufacturer's instructions. Purified Cas9 protein was concentrated using Ultracel-100K (Millipore).

4-2. Production of Guide RNA

The genomic sequence of the Arabidopsis gene encoding the BR11 was screened for the presence of a NGG motif, the so called protospacer adjacent motif (PAM), in an exon which is required for Cas9 targeting. To disrupt the BR11 gene in Arabidopsis, we identified two RGEN target sites in an exon that contain the NGG motif. sgRNAs were produced in vitro using template DNA. Each template DNA was generated by extension with two partially overlapped oligonucleotides (Macrogen, Table X1) and Phusion polymerase (Thermo Scientific) using the following conditions: -98° C. 30 sec {98° C. 10 sec, 54° C. 20 sec, 72° C. 2 min}×20, 72° C. 5 min.

TABLE 9

Oligonucleotides for the production of the template DNA for in vitro transcription		
Oligonucleotides	Sequence (5'-3')	SEQ ID NO
BRI1 target 1 (Forward)	GAAATTAATACGACTCACTATAGGTTTGAAGATGG AAGCGCGGGTTT TAGAGCTAGAAATAGCAAGTTAAA ATAAGGCTAGTCCG	73
BRI1 target 2 (Forward)	GAAATTAATACGACTCACTATAGGTGAACTAACT GGTCCACAGTTT TAGAGCTAGAAATAGCAAGTTAAA ATAAGGCTAGTCCG	74
Universal (Reverse)	AAAAAAGCACCGACTCGGTGCCACTTTTTCAAGTTG ATAACGGACTAGCCTTATTTAACTTGC	75

The extended DNA was purified and used as a template for the in vitro production of the guide RNA's using the MEGAshortscript T7 kit (Life Technologies). Guide RNA were then purified by Phenol/Chloroform extraction and ethanol precipitation. To prepare Cas9/sgRNA complexes, 10 μ l of purified Cas9 protein (12 μ g/ μ l) and 4 μ l each of two sgRNAs (11 μ g/ μ l) were mixed in 20 μ l NEB3 buffer (New England Biolabs) and incubated for 10 min at 37° C.

4-3. Transfection of Cas9/sgRNA Complex to Protoplast

The leaves of 4-week-old Arabidopsis seedlings grown aseptically in petri dishes were digested in enzyme solution (1% cellulose R10, 0.5% macerozyme R10, 450 mM mannitol, 20 mM MES pH 5.7 and CPW salt) for 8-16 hrs at 25° C. with 40 rpm shaking in the dark. Enzyme/protoplast solutions were filtered and centrifuged at 100 \times g for 3~5 min. Protoplasts were re-suspended in CPW solution after counting cells under the microscope (\times 100) using a hemacytometer. Finally, protoplasts were re-suspended at 1 \times 10⁶/ml in MMG solution (4 mM HEPES pH 5.7, 400 mM mannitol and 15 mM MgCl₂). To transfect the protoplasts with Cas9/sgRNA complex, 200 μ l (200,000 protoplasts) of the protoplast suspension were gently mixed with 3.3 or 10 μ l of Cas9/sgRNA complex [Cas9 protein (6 μ g/ μ l) and two sgRNAs (2.2 μ g/ μ l each)] and 200 μ l of 40% polyethylene glycol transfection buffer (40% PEG4000, 200 mM mannitol and 100 mM CaCl₂) in 2 ml tubes. After 5~20 min incubation at room temperature, transfection was stopped by adding wash buffer with W5 solution (2 mM MES pH 5.7, 154 mM NaCl, 125 mM CaCl₂ and 5 mM KCl). Protoplasts were then collected by centrifugation for 5 min at 100 \times g, washed with 1 ml of W5 solution, centrifuged for another 5 min at 100 \times g. The density of protoplasts was adjusted to 1 \times 10⁵/ml and they were cultured in modified KM 8p liquid medium with 400 mM glucose.

4-4. Detection of Mutations in Arabidopsis Protoplasts and Plants

After 24 hr or 72 hr post-transfection, protoplasts were collected and genomic DNA was isolated. The genomic DNA region spanning the two target sites was PCR-amplified and subjected to the T7E1 assay. As shown in FIG. 11, indels were induced by RGENs at high frequencies that ranged from 50% to 70%. Surprisingly, mutations were induced at 24 hr post-transfection. Apparently, Cas9 protein functions immediately after transfection. PCR products were purified and cloned into T-Blunt PCR Cloning Kit (Solgent). Plasmids were purified and subjected to Sanger sequencing with M13F primer. One mutant sequence had a 7-bp deletion at one site (FIG. 12). The other three mutant sequences had deletions of ~220-bp DNA segments between the two RGEN site.

EXAMPLE 5: CAS9 PROTEIN TRANSDUCTION USING A CELL-PENETRATING PEPTIDE OR PROTEIN TRANSDUCTION DOMAIN

5-1. Construction of His-Cas9-Encoding Plasmid

Cas9 with a cysteine at the C-terminal was prepared by PCR amplification using the previously described Cas9 plasmid {Cho, 2013 #166} as the template and cloned into

pET28-(a) vector (Novagen, Merck Millipore, Germany) containing His-tag at the N-terminus.

5-2. Cell Culture

293T (Human embryonic kidney cell line), and HeLa (human ovarian cancer cell line) were grown in DMEM (GIBCO-BRL Rockville) supplemented with 10% FBS and 1% penicillin and streptomycin.

5-3. EXPRESSION AND PURIFICATION OF CAS9 PROTEIN

To express the Cas9 protein, *E. coli* BL21 cells were transformed with the pET28-(a) vector encoding Cas9 and plated onto Luria-Bertani (LB) agar medium containing 50 μ g/mL kanamycin (Amresco, Solon, OH). Next day, a single colony was picked and cultured in LB broth containing 50 μ g/mL kanamycin at 37° C. overnight. Following day, this starter culture at 0.1 OD₆₀₀ was inoculated into Luria broth containing 50 μ g/mL kanamycin and incubated for 2 hrs at 37° C. until OD₆₀₀ reached to 0.6-0.8. To induce Cas9 protein expression, the cells were cultured at 30° C. overnight after addition of isopropyl- β -D-thiogalactopyranoside (IPTG) (Promega, Madison, WI) to the final concentration of 0.5 mM.

The cells were collected by centrifugation at 4000 rpm for 15-20 mins, resuspended in a lysis buffer (20 mM Tris-Cl pH8.0, 300 mM NaCl, 20 mM imidazole, 1 \times protease inhibitor cocktail, 1 mg/ml lysozyme), and lysed by sonication (40% duty, 10 sec pulse, 30 sec rest, for 10 mins on ice). The soluble fraction was separated as the supernatant after centrifugation at 15,000 rpm for 20 mins at 4° C. Cas9 protein was purified at 4° C. using a column containing Ni-NTA agarose resin (QIAGEN) and AKTA prime instrument (AKTA prime, GE Healthcare, UK). During this chromatography step, soluble protein fractions were loaded onto Ni-NTA agarose resin column (GE Healthcare, UK) at the flow rate of 1 mL/min. The column was washed with a washing buffer (20 mM Tris-Cl pH8.0, 300 mM NaCl, 20 mM imidazole, 1 \times protease inhibitor cocktail) and the bound protein was eluted at the flow rate of 0.5 ml/min with an elution buffer (20 mM Tris-Cl pH8.0, 300 mM NaCl, 250 mM imidazole, 1 \times protease inhibitor cocktail). The pooled eluted fraction was concentrated and dialyzed against storage buffer (50 mM Tris-HCl, pH8.0, 200 mM KCl, 0.1 mM EDTA, 1 mM DTT, 0.5 mM PMSF, 20% Glycerol). Protein concentration was quantitated by Bradford assay (Biorad, Hercules, CA) and purity was analyzed by SDS-PAGE using bovine serum albumin as the control.

5-4. Conjugation of Cas9 to 9R4L

1 mg Cas9 protein diluted in PBS at the concentration of 1 mg/mL and 50 μ g of maleimide-9R4L peptide in 25 μ l DW (Pepton, Korea) were gently mixed using a rotor at room temperature for 2 hrs and at 4° C. overnight. To remove unconjugated maleimide-9R4L, the samples were dialyzed using 50kDa molecular weight cutoff membrane against of DPBS (pH 7.4) at 4° C. for 24 hrs. Cas9-9R4L protein was collected from the dialysis membrane and the protein amount was determined using Bradford assay.

5-5. Preparation of sgRNA-9R4L

sgRNA (1 μ g) was gently added to various amounts of C9R4LC peptide (ranging from 1 to 40 weight ratio) in 100

µl of DPBS (pH 7.4). This mixture was incubated at room temperature for 30 mins and diluted to 10 folds using RNase-free deionized water. The hydrodynamic diameter and z-potential of the formed nanoparticles were measured using dynamic light scattering (Zetasizer-nano analyzer ZS; Malvern instruments, Worcestershire, UK).

5-6. Cas9 Protein and sgRNA Treatments

Cas9-9R4L and sgRNA-C9R4LC were treated to the cells as follows: 1 µg of sgRNA and 15 µg of C9R4LC peptide were added to 250 mL of OPTIMEM medium and incubated at room temperature for 30 mins. At 24 hrs after seeding, cells were washed with OPTIMEM medium and treated with sgRNA-C9R4LC complex for 4 hrs at 37° C. Cells were washed again with OPTIMEM medium and treated with Cas9-9R4L for 2 hrs at 37° C. After treatment, culture media was replaced with serum-containing complete medium and incubated at 37° C. for 24 hrs before the next treatment. Same procedure was followed for multiple treatments of Cas9 and sgRNA for three consecutive days.

5-7. Cas9-9R4L and sgRNA-9R4L Can Edit Endogenous Genes in Cultured Mammalian Cells Without the Use of Additional Delivery Tools

To determine whether Cas9-9R4L and sgRNA-9R4L can edit endogenous genes in cultured mammalian cells without the use of additional delivery tools, we treated 293 cells with Cas9-9R4L and sgRNA-9R4L targeting the CCR5 gene and analyzed the genomic DNA. T7E1 assay showed that 9% of CCR5 gene was disrupted in cells treated with both Cas9-9R4L and sgRNA-9R4L and that the CCR5 gene disruption was not observed in control cells including those untreated, treated with either Cas9-9R or sgRNA-9R4L, or treated with both unmodified Cas-9 and sgRNA (FIG. 13), suggesting that the treatment with Cas9-9R4L protein and sgRNA conjugated with 9R4L, but not unmodified Cas9 and sgRNA, can lead to efficient genome editing in mammalian cells.

EXAMPLE 6: CONTROL OF OFF-TARGET MUTATION ACCORDING TO GUIDE RNA STRUCTURE

Recently, three groups reported that RGENs had off-target effects in human cells. To our surprise, RGENs induced mutations efficiently at off-target sites that differ by 3 to 5 nucleotides from on-target sites. We noticed, however, that there were several differences between our RGENs and those used by others. First, we used dualRNA, which is crRNA plus tracrRNA, rather than single-guide RNA (sgRNA) that is composed of essential portions of crRNA and tracrRNA. Second, we transfected K562 cells (but not HeLa cells) with synthetic crRNA rather than plasmids encoding crRNA. HeLa cells were transfected with crRNA-encoding plasmids. Other groups used sgRNA-encoding plasmids. Third, our guide RNA had two additional guanine nucleotides at the 5' end, which are required for efficient transcription by T7 polymerase in vitro. No such additional nucleotides were included in the sgRNA used by others. Thus, the RNA sequence of our guide RNA can be shown as 5'-GGX₂₀, whereas 5'-GX₁₉, in which X₂₀ or GX₁₉ corresponds to the 20-bp target sequence, represents the sequence used by others. The first guanine nucleotide is required for transcription by RNA polymerase in cells. To test whether off-target RGEN effects can be attributed to these differ-

ences, we chose four RGENs that induced off-target mutations in human cells at high frequencies (13). First, we compared our method of using in vitro transcribed dualRNA with the method of transfecting sgRNA-encoding plasmids in K562 cells and measured mutation frequencies at the on-target and off-target sites via the T7E1 assay. Three RGENs showed comparable mutation frequencies at on-target and off-target sites regardless of the composition of guide RNA. Interestingly, one RGEN (VEFGA site 1) did not induce indels at one validated off-target site, which differs by three nucleotides from the on-target site (termed OT1-11, FIG. 14), when synthetic dualRNA was used. But the synthetic dualRNA did not discriminate the other validated off-target site (OT1-3), which differs by two nucleotides from the on-target site.

Next, we tested whether the addition of two guanine nucleotides at the 5' end of sgRNA could make RGENs more specific by comparing 5'-GGX₂₀ (or 5'-GGGX₁₉) sgRNA with 5'-GX₁₉ sgRNA. Four GX₁₉ sgRNAs complexed with Cas9 induced indels equally efficiently at on-target and off-target sites, tolerating up to four nucleotide mismatches. In sharp contrast, GGX₂₀ sgRNAs discriminated off-target sites effectively. In fact, the T7E1 assay barely detected RGEN-induced indels at six out of the seven validated off-target sites when we used the four GGX₂₀ sgRNAs (FIG. 15). We noticed, however, that two GGX₂₀ sgRNAs (VEFGA sites 1 and 3) were less active at on-target sites than were the corresponding GX₁₉ sgRNAs. These results show that the extra nucleotides at the 5' end can affect mutation frequencies at on-target and off-target sites, perhaps by altering guide RNA stability, concentration, or secondary structure.

These results suggest that three factors—the use of synthetic guide RNA rather than guide RNA-encoding plasmids, dualRNA rather than sgRNA, and GGX₂₀ sgRNA rather than GX₁₉ sgRNA—have cumulative effects on the discrimination of off-target sites.

EXAMPLE 7: PAIRED CAS9 NICKASES

In principle, single-strand breaks (SSBs) cannot be repaired by error-prone NHEJ but still trigger high fidelity homology-directed repair (HDR) or base excision repair. But nickase-induced targeted mutagenesis via HDR is much less efficient than is nuclease-induced mutagenesis. We reasoned that paired Cas9 nickases would produce composite DSBs, which trigger DNA repair via NHEJ or HDR, leading to efficient mutagenesis (FIG. 16A). Furthermore, paired nickases would double the specificity of Cas9-based genome editing.

We first tested several Cas9 nucleases and nickases designed to target sites in the AAVSI locus (FIG. 16B) in vitro via fluorescent capillary electrophoresis. Unlike Cas9 nucleases that cleaved both strands of DNA substrates, Cas9 nickases composed of guide RNA and a mutant form of Cas9 in which a catalytic aspartate residue is changed to an alanine (D10A Cas9) cleaved only one strand, producing site-specific nicks (FIG. 16C, D). Interestingly, however, some nickases (AS1, AS2, AS3, and S6 in FIG. 17A) induced indels at target sites in human cells, suggesting that nicks can be converted to DSBs, albeit inefficiently, in vivo. Paired Cas9 nickases producing two adjacent nicks on opposite DNA strands yielded indels at frequencies that ranged from 14% to 91%, comparable to the effects of paired nucleases (FIG. 17A). The repair of two nicks that would produce 5' overhangs led to the formation of indels much more frequently than those producing 3' overhangs at three

genomic loci (FIG. 17A and FIG. 18). In addition, paired nickases enabled targeted genome editing via homology-directed repair more efficiently than did single nickases (FIG. 19).

We next measured mutation frequencies of paired nickases and nucleases at off-target sites using deep sequencing. Cas9 nucleases complexed with three sgRNAs induced off-target mutations at six sites that differ by one or two nucleotides from their corresponding on-target sites with frequencies that ranged from 0.5% to 10% (FIG. 17B). In contrast, paired Cas9 nickases did not produce indels above the detection limit of 0.1% at any of the six off-target sites. The S2 Off-1 site that differs by a single nucleotide at the first position in the PAM (i.e., N in NGG) from its on-target site can be considered as another on-target site. As expected, the Cas9 nuclease complexed with the S2 sgRNA was equally efficient at this site and the on-target site. In sharp contrast, D10A Cas9 complexed with the S2 and AS2 sgRNAs discriminated this site from the on-target site by a factor of 270 fold. This paired nickase also discriminated the AS2 off-target sites (Off-1 and Off-9 in FIG. 17B) from the on-target site by factors of 160 fold and 990 fold, respectively.

EXAMPLE 8: CHROMOSOMAL DNA SPLICING INDUCED BY PAIRED CAS9 NICKASES

Two concurrent DSBs produced by engineered nucleases such as ZENs and TALENs can promote large deletions of the intervening chromosomal segments has been reported. We tested whether two SSBs induced by paired Cas9 nickases can also produce deletions in human cells. We used PCR to detect deletion events and found that seven paired nickases induced deletions of up to 1.1-kbp chromosomal segments as efficiently as paired Cas9 nucleases did (FIG. 20A, B). DNA sequences of the PCR products confirmed the deletion events (FIG. 20C). Interestingly, the sgRNA-matching sequence remained intact in two out of seven deletion-specific PCR amplicons (underlined in FIG. 20C). In contrast, Cas9 nuclease pairs did not produce sequences that contained intact target sites. This finding suggests that two distant nicks were not converted to two separate DSBs to promote deletions of the intervening chromosomal segment. In addition, it is unlikely that two nicks separated by more than a 100 bp can produce a composite DSB with large overhangs under physiological conditions because the melting temperature is very high.

We propose that two distant nicks are repaired by strand displacement in a head-to-head direction, resulting in the formation of a DSB in the middle, whose repair via NHEJ causes small deletions (FIG. 20D). Because the two target sites remain intact during this process, nickases can induce SSBs again, triggering the cycle repeatedly until the target sites are deleted. This mechanism explains why two offset nicks producing 5' overhangs but not those producing 3' overhangs induced indels efficiently at three loci.

We then investigated whether Cas9 nucleases and nickases can induce unwanted chromosomal translocations that result from NHEJ repair of on-target and off-target DNA cleavages (FIG. 21A). We were able to detect translocations induced by Cas9 nucleases using PCR (FIG. 21B, C). No such PCR products were amplified using genomic DNA isolated from cells transfected with the plasmids encoding the AS2+S3 Cas9 nickase pair. This result is in line with the fact that both AS2 and S3 nickases, unlike their corresponding nucleases, did not produce indels at off-target sites (FIG. 17B).

These results suggest that paired Cas9 nickases allow targeted mutagenesis and large deletions of up to 1-kbp chromosomal segments in human cells. Importantly, paired nickases did not induce indels at off-target sites at which their corresponding nucleases induce mutations. Furthermore, unlike nucleases, paired nickases did not promote unwanted translocations associated with off-target DNA cleavages. In principle, paired nickases double the specificity of Cas9-mediated mutagenesis and will broaden the utility of RNA-guided enzymes in applications that require precise genome editing such as gene and cell therapy. One caveat to this approach is that two highly active sgRNAs are needed to make an efficient nickase pair, limiting targetable sites. As shown in this and other studies, not all sgRNAs are equally active. When single clones rather than populations of cells are used for further studies or applications, the choice of guide RNAs that represent unique sequences in the genome and the use of optimized guide RNAs would suffice to avoid off-target mutations associated with Cas9 nucleases. We propose that both Cas9 nucleases and paired nickases are powerful options that will facilitate precision genome editing in cells and organisms.

EXAMPLE 9: GENOTYPING WITH CRISPR/CAS-DERIVED RNA-GUIDED ENDONUCLEASES

Next, we reasoned that RGENs can be used in Restriction fragment length polymorphism (RFLP) analysis, replacing conventional restriction enzymes. Engineered nucleases including RGENs induce indels at target sites, when the DSBs caused by the nucleases are repaired by the error-prone non-homologous end-joining (NHEJ) system. RGENs that are designed to recognize the target sequences cannot cleave mutant sequences with indels but will cleave wild-type target sequences efficiently.

9-1. RGEN Components

crRNA and tracrRNA were prepared by in vitro transcription using MEGAscript T7 kit (Ambion) according to the manufacturer's instruction. Transcribed RNAs were resolved on a 8% denaturing urea-PAGE gel. The gel slice containing RNA was cut out and transferred to elution buffer. RNA was recovered in nuclease-free water followed by phenol: chloroform extraction, chloroform extraction, and ethanol precipitation. Purified RNA was quantified by spectrometry. Templates for crRNA were prepared by annealing an oligonucleotide whose sequence is shown as 5'-GAAATTAATACGACTCACTATAGGX₂₀GTTTTAGAGCTATGCTGTTTTG-3' (SEQ ID NO: 76), in which X₂₀ is the target sequence, and its complementary oligonucleotide. The template for tracrRNA was synthesized by extension of forward and reverse oligonucleotides (5'-GAAATTAATACGACTCACTATAGGAACCAATCAAACAGCATAGCAAGTTAAAATAAGGCTA GTCCG-3' (SEQ ID NO: 77) and 5'-AAAAAAGCACCGACTCGGTGC-CACTTTTCAAGTTGATAACGGACTAGCCTATT-TAACT TGCTATG-3' (SEQ ID NO: 78)) using Phusion polymerase (New England Biolabs).

9-2. Recombinant Cas9 Protein Purification

The Cas9 DNA construct used in our previous Example, which encodes Cas9 fused to the His6-tag at the C terminus, was inserted in the pET-28a expression vector. The recombinant Cas9 protein was expressed in *E. coli* strain BL21

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(DE3) cultured in LB medium at 25° C. for 4 hours after induction with 1 mM IPTG. Cells were harvested and resuspended in buffer containing 20 mM Tris pH 8.0, 500 mM NaCl, 5 mM imidazole, and 1 mM PMSF. Cells were frozen in liquid nitrogen, thawed at 4° C., and sonicated. After centrifugation, the Cas9 protein in the lysate was bound to Ni-NTA agarose resin (Qiagen), washed with buffer containing 20 mM Tris pH 8.0, 500 mM NaCl, and 20 mM imidazole, and eluted with buffer containing 20 mM Tris pH 8.0, 500 mM NaCl, and 250 mM imidazole. Purified Cas9 protein was dialyzed against 20 mM HEPES (pH 7.5), 150 mM KCl, 1 mM DTT, and 10% glycerol and analyzed by SDS-PAGE.

9-3. T7 Endonuclease I Assay

The T7E1 assay was performed as following. In brief, PCR products amplified using genomic DNA were denatured at 95° C., reannealed at 16° C., and incubated with 5 units of T7 Endonuclease I (New England BioLabs) for 20 min at 37° C. The reaction products were resolved using 2 to 2.5% agarose gel electrophoresis.

9-4. RGEN-RFLP Assay

PCR products (100-150 ng) were incubated for 60 min at 37° C. with optimized concentrations (Table 10) of Cas9 protein, tracrRNA, crRNA in 10 µl NEB buffer 3 (1×). After the cleavage reaction, RNase A (4 µg) was added, and the reaction mixture was incubated for 30 min at 37° C. to remove RNA. Reactions were stopped with 6× stop solution

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buffer containing 30% glycerol, 1.2% SDS, and 100 mM EDTA. Products were resolved with 1-2.5% agarose gel electrophoresis and visualized with EtBr staining.

TABLE 10

Concentration of RGEN components in RFLP assays			
Target Name	Cas9 (ng/µl)	crRNA (ng/µl)	tracrRNA (ng/µl)
C4BPB	100	25	60
PIBF-NGG-RGEN	100	25	60
HLA-B	1.2	0.3	0.7
CCR5-ZFN	100	25	60
CTNNB1 Wild type specific	30	10	20
CTNNB1 mutant specific	30	10	20
CCR5 WT-specific	100	25	60
CCR5 Δ32-specific	10	2.5	6
KRAS WT specific(wt)	30	10	20
KRAS mutant specific(m8)	30	10	20
KRAS WT specific (m6)	30	10	20
KRAS mutant specific (m6, 8)	30	10	20
PIK3CA WT specific (wt)	100	25	60
PIK3CA mutant specific(m4)	30	10	20
PIK3CA WT specific (m7)	100	25	60
PIK3CA mutant specific(m4, 7)	30	10	20
BRAF WT-specific	30	10	20
BRAF mutant-specific	100	25	60
NRAS WT-specific	100	25	60
NRAS mutant-specific	30	10	20
IDH WT-specific	30	10	20
IDH mutant-specific	30	10	20
PIBF-NAG-RGEN	30	10	60

TABLE 11

Primers			
Gene(site)	Direction	Sequence(5' to 3')	SEQ ID NO
CCR5 (RGEN)	F1	CTCCATGGTGCTATAGAGCA	79
	F2	GAGCCAAGCTCTCCATCTAGT	80
	R	GCCCTGTCAAGAGTTGACAC	81
CCR5 (ZFN)	F	GCACAGGGTGAACAAGATGGA	82
	R	GCCAGGTACCTATCGATTGTCAGG	83
CCR5 (del32)	F	GAGCCAAGCTCTCCATCTAGT	84
	R	ACTCTGACTG GGTACCAGC	85
C4BPB	F1	TATTTGGCTGGTTGAAAGGG	86
	R1	AAAGTCATGAAATAAACACACCCCA	87
	F2	CTGCATTGATATGGTAGTACCATG	88
	R2	GCTGTTCAATTGCAATGGAATG	89
CTNNB1	F	ATGGAGTTGGACATGGCCATGG	90
	R	ACTCACTATCCACAGTTCAGCATTTACC	91
KRAS	F	TGGAGATAGCTGTCAGCAACTTT	92
	R	CAACAA AGCAAAGGTAAAGTTGGTAATAG	93
PIK3CA	F	GGTTTCAGGAGATGTGTTACAAGGC	94
	R	GATTGTGCAATTCCATGCAATCGGTC	95
NRAS	F	CACTGGGTACTTAATCTGTAGCCCTC	96
	R	GGTTCCAAGTCATTCCAGTAGC	97
IDH1	F	CATCACTGCAGTTGTAGGTTATAACTATCC	98
	R	TTGAAAACCAACAGATCTGGTTGAACC	99
BRAF	F	GGAGTGCCAAGAGAATATCTGG	100
	R	CTGAAACTGGTTTCAAATAATTCGTTTAAAGG	101
PIBF	F	GCTCTGTATGCCCTGTAGTAGG	102
	R	TTTGCATCTGACCTTACCTTTG	103

9-5. Plasmid Cleavage Assay

Restriction enzyme-treated linearized plasmid (100 ng) was incubated for 60 min at 37° C. with Cas9 protein (0.1 µg), trackRNA (60 ng), and crRNA (25 ng) in 10 µl NEB 3 buffer (1×). Reactions were stopped with 6× stop solution containing 30% glycerol, 1.2% SDS, and 100 mM EDTA. Products were resolved with 1% agarose gel electrophoresis and visualized with EtBr staining.

9-6. Strategy of RFLP

New RGENs with desired DNA specificities can be readily created by replacing crRNA; no de novo purification of custom proteins is required once recombinant Cas9 protein is available. Engineered nucleases, including RGENs, induce small insertions or deletions (indels) at target sites when the DSBs caused by the nucleases are repaired by error-prone non-homologous end-joining (NHEJ). RGENs that are designed to recognize the target sequences cleave wild-type sequences efficiently but cannot cleave mutant sequences with indels (FIG. 22).

We first tested whether RGENs can differentially cleave plasmids that contain wild-type or modified C4BPB target sequences that harbor 1- to 3-base indels at the cleavage site. None of the six plasmids with these indels were cleaved by a C4BPB-specific RGEN5 composed of target-specific crRNA, tracrRNA, and recombinant Cas9 protein (FIG. 23). In contrast, the plasmid with the intact target sequence was cleaved efficiently by this RGEN.

9-7. Detection of Mutations Induced by the Same RGENs Using RGEN-Mediated RFLP

Next, to test the feasibility of RGEN-mediated RFLP for detection of mutations induced by the same RGENs, we utilized gene-modified K562 human cancer cell clones established using an RGEN targeting C4BPB gene (Table 12).

TABLE 12

Target sequence of RGENs used in this study		
Gene	Target sequence	SEQ ID NO
human C4BPB	<u>AATGACCACTACATCCTCAAGGG</u>	104
mouse Pibf1	<u>AGATGATGTCATCATCAGAGG</u>	105

C4BPB mutant clones used in this study have various mutations ranging from 94 bp deletion to 67 bp insertion (FIG. 24A). Importantly, all mutations occurred in mutant clones resulted in the loss of RGEN target site. Among 6 C4BPB clones analyzed, 4 clones have both wildtype and mutant alleles (+/-) and 2 clones have only mutant alleles (-/-).

The PCR products spanning the RGEN target site amplified from wildtype K562 genomic DNA were digested completely by the RGEN composed of target-specific crRNA, tracrRNA, and recombinant Cas9 protein expressed in and purified from *E. coli* (FIG. 24B/Lane 1). When the C4BPB mutant clones were subjected to RFLP analysis using the RGEN, PCR amplicons of +/- clones that contained both wildtype and mutant alleles were partially digested, and those of -/- clones that did not contain the wildtype allele were not digested at all, yielding no cleavage products corresponding to the wildtype sequence (FIG.

24B). Even a single-base insertion at the target site blocked the digestion (#12 and #28 clones) of amplified mutant alleles by the C4BPB RGEN, showing the high specificity of RGEN-mediated RFLP. We subjected the PCR amplicons to the mismatch-sensitive T7E1 assay in parallel (FIG. 24B). Notably, the T7E1 assay was not able to distinguish -/- clones from +/- clones. To make it matters worse, the T7E1 assay cannot distinguish homozygous mutant clones that contain the same mutant sequence from wildtype clones, because annealing of the same mutant sequence will form a homoduplex. Thus, RGEN-mediated RFLP has a critical advantage over the conventional mismatch-sensitive nucle- ase assay in the analysis of mutant clones induced by engineered nucleases including ZENs, TALENs and RGENs.

9-8. Quantitative Assay for RGEN-RFLP Analysis

We also investigated whether RGEN-RFLP analysis is a quantitative method. Genomic DNA samples isolated from the C4BPB null clone and the wild-type cells were mixed at various ratios and used for PCR amplifications. The PCR products were subjected to RGEN genotyping and the T7E1 assay in parallel (FIG. 25b). As expected, DNA cleavage by the RGEN was proportional to the wild type to mutant ratio. In contrast, results of the T7E1 assay correlated poorly with mutation frequencies inferred from the ratios and were inaccurate, especially at high mutant %, a situation in which complementary mutant sequences can hybridize with each other to form homoduplexes.

9-9. Analysis of Mutant Mouse Founders Using a RGEN-Mediated RFLP Genotyping

We also applied RGEN-mediated RFLP genotyping (RGEN genotyping in short) to the analysis of mutant mouse founders that had been established by injection of TALENs into mouse one-cell embryos (FIG. 26A). We designed and used an RGEN that recognized the TALEN target site in the Pibf1 gene (Table 10). Genomic DNA was isolated from a wildtype mouse and mutant mice and subjected to RGEN genotyping after PCR amplification. RGEN genotyping successfully detected various mutations, which ranged from one to 27-bp deletions (FIG. 26B). Unlike the T7E1 assay, RGEN genotyping enabled differential detection of +/- and -/- founder.

9-10. Detection of Mutations Induced in Human Cells by a CCR5-Specific ZEN using RGENs

In addition, we used RGENs to detect mutations induced in human cells by a CCR5-specific ZFN, representing yet another class of engineered nucleases (FIG. 27). These results show that RGENs can detect mutations induced by nucleases other than RGENs themselves. In fact, we expect that RGENs can be designed to detect mutations induced by most, if not all, engineered nucleases. The only limitation in the design of an RGEN genotyping assay is the requirement for the GG or AG (CC or CT on the complementary strand) dinucleotide in the PAM sequence recognized by the Cas9 protein, which occurs once per 4 bp on average. Indels induced anywhere within the seed region of several bases in crRNA and the PAM nucleotides are expected to disrupt RGEN-catalyzed DNA cleavage. Indeed, we identified at least one RGEN site in most (98%) of the ZEN and TALEN sites.

9-11. Detection of Polymorphisms or Variations Using RGEN

Next, we designed and tested a new RGEN that targets a highly polymorphic locus, HLA-B, that encodes Human Leukocyte Antigen B (a.k.a. MHC class I protein) (FIG. 28). HeLa cells were transfected with RGEN plasmids, and the genomic DNA was subjected to T7E1 and RGEN-RFLP analyses in parallel. T7E1 produced false positive bands that resulted from sequence polymorphisms near the target site (FIG. 25c). As expected, however, the same RGEN used for gene disruption cleaved PCR products from wild-type cells completely but those from RGEN-transfected cells partially, indicating the presence of RGEN-induced indels at the target site. This result shows that RGEN-RFLP analysis has a clear advantage over the T7E1 assay, especially when it is not known whether target genes have polymorphisms or variations in cells of interest.

9-12. Detection of Recurrent Mutations Found in Cancer and Naturally-Occurring Polymorphisms through RGEN-RFLP Analysis

RGEN-RFLP analysis has applications beyond genotyping of engineered nuclease-induced mutations. We sought to use RGEN genotyping to detect recurrent mutations found in cancer and naturally-occurring polymorphisms. We chose the human colorectal cancer cell line, HCT116, which carries a gain-of-function 3-bp deletion in the oncogenic CTNNB1 gene encoding beta-catenin. PCR products amplified from HCT116 genomic DNA were cleaved partially by both wild-type-specific and mutant-specific RGENs, in line with the heterozygous genotype in HCT116 cells (FIG. 29a). In sharp contrast, PCR products amplified from DNA from HeLa cells harboring only wild-type alleles were digested completely by the wild-type-specific RGEN and were not cleaved at all by the mutation-specific RGEN.

We also noted that HEK293 cells harbor the 32-bp deletion (del32) in the CCR5 gene, which encodes an essential co-receptor of HIV infection: Homozygous del32 CCR5 carriers are immune to HIV infection. We designed one RGEN specific to the del32 allele and the other to the wild-type allele. As expected, the wild-type-specific RGEN cleaved the PCR products obtained from K562, SKBR3, or HeLa cells (used as wild-type controls) completely but those from HEK293 cells partially (FIG. 30a), confirming the presence of the uncleavable del32 allele in HEK293 cells. Unexpectedly, however, the del32-specific RGEN cleaved the PCR products from wild-type cells as efficiently as those from HEK293 cells. Interestingly, this RGEN had an off-target site with a single-base mismatch immediately downstream of the on-target site (FIG. 30). These results suggest that RGENs can be used to detect naturally-occurring indels but cannot distinguish sequences with single nucleotide polymorphisms or point mutations due to their off-target effects.

To genotype oncogenic single-nucleotide variations using RGENs, we attenuated RGEN activity by employing a single-base mismatched guide RNA instead of a perfectly-matched RNA. RGENs that contained the perfectly-matched guide RNA specific to the wild-type sequence or mutant sequence cleaved both sequences (FIGS. 31a and 32a). In contrast, RGENs that contained a single-base mismatched guide RNA distinguished the two sequences, enabling genotyping of three recurrent oncogenic point mutations in the

KRAS, PIK3CA, and IDH1 genes in human cancer cell lines (FIG. 29b and FIGS. 33a, b). In addition, we were able to detect point mutations in the BRAF and NRAS genes using RGENs that recognize the NAG PAM sequence (FIGS. 33c, d). We believe that we can use RGEN-RFLP to genotype almost any, if not all, mutations or polymorphisms in the human and other genomes.

The above data proposes RGENs as providing a platform to use simple and robust RFLP analysis for various sequence variations. With high flexibility in reprogramming target sequence, RGENs can be used to detect various genetic variations (single nucleotide variations, small insertion/deletions, structural variations) such as disease-related recurring mutations, genotypes related to drug-response by a patient and also mutations induced by engineered nucleases in cells. Here, we used RGEN genotyping to detect mutations induced by engineered nucleases in cells and animals. In principle, one could also use RGENs that will specifically detect and cleave naturally-occurring variations and mutations.

Based on the above description, it should be understood by those skilled in the art that various alternatives to the embodiments of the invention described herein may be employed in practicing the invention without departing from the technical idea or essential features of the invention as defined in the following claims. In this regard, the above-described examples are for illustrative purposes only, and the invention is not intended to be limited by these examples. The scope of the present invention should be understood to include all of the modifications or modified form derived from the meaning and scope of the following claims or its equivalent concepts.

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SEQUENCE LISTING

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ccccccgct tcaagtactt cgacaccacc atcgaccgca agcgctacac cagcaccaag 4020
gaggtgctgg acgccaccct gatccaccag agcatcaccg gtctgtacga gaccgcacatc 4080
gacctgagcc agctggggcg cgactaa 4107

SEQ ID NO: 2          moltype = AA  length = 21
FEATURE              Location/Qualifiers
REGION              1..21
                    note = Description of Artificial Sequence: Synthetic peptide
REGION              1..21
                    note = Peptide tag
source              1..21
                    mol_type = protein
                    organism = synthetic construct

SEQUENCE: 2
GGSGPPKKKR KVYPDVPDY A 21

SEQ ID NO: 3          moltype = DNA  length = 34
FEATURE              Location/Qualifiers
misc_feature         1..34
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
misc_feature         1..34
                    note = F primer for CCR5
source              1..34
                    mol_type = other DNA
                    organism = synthetic construct

SEQUENCE: 3
aattcatgac atcaattatt atacatcgga ggag 34

SEQ ID NO: 4          moltype = DNA  length = 34
FEATURE              Location/Qualifiers
misc_feature         1..34
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
misc_feature         1..34
                    note = R primer for CCR5
source              1..34
                    mol_type = other DNA
                    organism = synthetic construct

SEQUENCE: 4
gatcctcctc cgatgtataa taattgatgt catg 34

SEQ ID NO: 5          moltype = DNA  length = 20
FEATURE              Location/Qualifiers
misc_feature         1..20
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
misc_feature         1..20
                    note = F1 primer for CCR5
source              1..20
                    mol_type = other DNA
                    organism = synthetic construct

SEQUENCE: 5
ctccatggtg ctatagagca 20

SEQ ID NO: 6          moltype = DNA  length = 21
FEATURE              Location/Qualifiers
misc_feature         1..21
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
misc_feature         1..21
                    note = F2 primer for CCR5
source              1..21
                    mol_type = other DNA
                    organism = synthetic construct

SEQUENCE: 6
gagccaagct ctccatctag t 21

SEQ ID NO: 7          moltype = DNA  length = 20
FEATURE              Location/Qualifiers
misc_feature         1..20
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
misc_feature         1..20
                    note = R primer for CCR5
source              1..20
                    mol_type = other DNA
                    organism = synthetic construct

SEQUENCE: 7

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gccctgtcaa gagttgacac 20

SEQ ID NO: 8 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..20
 note = F1 primer for C4BPB
 source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 8
 tatttggctg gttgaaagg 20

SEQ ID NO: 9 moltype = DNA length = 24
 FEATURE Location/Qualifiers
 misc_feature 1..24
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..24
 note = R1 primer for C4BPB
 source 1..24
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 9
 aaagtcatga aataaacaca ccca 24

SEQ ID NO: 10 moltype = DNA length = 24
 FEATURE Location/Qualifiers
 misc_feature 1..24
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..24
 note = F2 primer for C4BPB
 source 1..24
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 10
 ctgcattgat atggtagtag catg 24

SEQ ID NO: 11 moltype = DNA length = 21
 FEATURE Location/Qualifiers
 misc_feature 1..21
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..21
 note = R2 primer for C4BPB
 source 1..21
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 11
 gctgttcatt gcaatggaat g 21

SEQ ID NO: 12 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..20
 note = F1 primer for ADCY5
 source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 12
 gctcccacct tagtgetctg 20

SEQ ID NO: 13 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..20
 note = R1 primer for ADCY5
 source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 13
 ggtggcagga acctgtatgt 20

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FEATURE                               Location/Qualifiers
misc_feature                           1..20
                                         note = Description of Artificial Sequence: Synthetic
                                         oligonucleotide
misc_feature                           1..20
                                         note = F primer for deletion
source                                 1..20
                                         mol_type = other DNA
                                         organism = synthetic construct

SEQUENCE: 26
ccacatctcg ttctcggtt                                     20

SEQ ID NO: 27                               moltype = DNA length = 20
FEATURE                               Location/Qualifiers
misc_feature                           1..20
                                         note = Description of Artificial Sequence: Synthetic
                                         oligonucleotide
misc_feature                           1..20
                                         note = R primer for deletion
source                                 1..20
                                         mol_type = other DNA
                                         organism = synthetic construct

SEQUENCE: 27
tcacaagccc acagatattt                                   20

SEQ ID NO: 28                               moltype = RNA length = 105
FEATURE                               Location/Qualifiers
misc_feature                           1..105
                                         note = Description of Artificial Sequence: Synthetic
                                         polynucleotide
misc_feature                           1..105
                                         note = sgRNA for CCR5
source                                 1..105
                                         mol_type = other RNA
                                         organism = synthetic construct

SEQUENCE: 28
ggtgacatca attattatac atgttttaga gctagaaata gcaagttaaa ataaggctag 60
tccgttatca acttgaaaaa gtcggcaccga gtcggtgctt ttttt          105

SEQ ID NO: 29                               moltype = RNA length = 44
FEATURE                               Location/Qualifiers
misc_feature                           1..44
                                         note = Description of Artificial Sequence: Synthetic
                                         oligonucleotide
misc_feature                           1..44
                                         note = crRNA for CCR5
source                                 1..44
                                         mol_type = other RNA
                                         organism = synthetic construct

SEQUENCE: 29
ggtgacatca attattatac atgttttaga gctatgctgt tttg          44

SEQ ID NO: 30                               moltype = RNA length = 86
FEATURE                               Location/Qualifiers
misc_feature                           1..86
                                         note = Description of Artificial Sequence: Synthetic
                                         oligonucleotide
misc_feature                           1..86
                                         note = tracrRNA for CCR5
source                                 1..86
                                         mol_type = other RNA
                                         organism = synthetic construct

SEQUENCE: 30
ggaaccattc aaaacagcat agcaagttaa aataaggcta gtccgttatc aacttgaaaa 60
agtggcaccg agtcggtgct tttttt          86

SEQ ID NO: 31                               moltype = DNA length = 86
FEATURE                               Location/Qualifiers
misc_feature                           1..86
                                         note = Description of Artificial Sequence: Synthetic
                                         oligonucleotide
misc_feature                           1..86
                                         note = Foxn1 #1 sgRNA
source                                 1..86
                                         mol_type = other DNA
                                         organism = synthetic construct

SEQUENCE: 31
gaaattaata cgactcacta taggcagtct gacgtcacac ttccgtttta gagctagaaa 60
tagcaagtta aaataaggct agtccc          86

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SEQ ID NO: 32 moltype = DNA length = 86
 FEATURE Location/Qualifiers
 misc_feature 1..86
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..86
 note = Foxn1 #2 sgRNA
 source 1..86
 mol_type = other DNA
 organism = synthetic construct

 SEQUENCE: 32
 gaaattaata cgactcacta taggacttcc aggctccacc cgacgtttta gagctagaaa 60
 tagcaagtta aaataaggct agtccg 86

SEQ ID NO: 33 moltype = DNA length = 86
 FEATURE Location/Qualifiers
 misc_feature 1..86
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..86
 note = Foxn1 #3 sgRNA
 source 1..86
 mol_type = other DNA
 organism = synthetic construct

 SEQUENCE: 33
 gaaattaata cgactcacta taggccaggc tccaccgcac tggagtttta gagctagaaa 60
 tagcaagtta aaataaggct agtccg 86

SEQ ID NO: 34 moltype = DNA length = 86
 FEATURE Location/Qualifiers
 misc_feature 1..86
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..86
 note = Foxn1 #4 sgRNA
 source 1..86
 mol_type = other DNA
 organism = synthetic construct

 SEQUENCE: 34
 gaaattaata cgactcacta taggactgga gggcgaaacc caaggtttta gagctagaaa 60
 tagcaagtta aaataaggct agtccg 86

SEQ ID NO: 35 moltype = DNA length = 86
 FEATURE Location/Qualifiers
 misc_feature 1..86
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..86
 note = Foxn1 #5 sgRNA
 source 1..86
 mol_type = other DNA
 organism = synthetic construct

 SEQUENCE: 35
 gaaattaata cgactcacta taggacccca aggggacctc atgcgtttta gagctagaaa 60
 tagcaagtta aaataaggct agtccg 86

SEQ ID NO: 36 moltype = DNA length = 86
 FEATURE Location/Qualifiers
 misc_feature 1..86
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..86
 note = Prkdc #1 sgRNA
 source 1..86
 mol_type = other DNA
 organism = synthetic construct

 SEQUENCE: 36
 gaaattaata cgactcacta taggttagtt tttccagag acttgtttta gagctagaaa 60
 tagcaagtta aaataaggct agtccg 86

SEQ ID NO: 37 moltype = DNA length = 86
 FEATURE Location/Qualifiers
 misc_feature 1..86
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..86
 note = Prkdc #2 sgRNA
 source 1..86

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mol_type = other DNA
organism = synthetic construct

SEQUENCE: 37
gaaattaata cgactcacta taggttggtt tgcttggtt tatcgttta gagctagaaa 60
tagcaagtta aaataaggct agtccg 86

SEQ ID NO: 38      moltype = DNA length = 86
FEATURE          Location/Qualifiers
misc_feature     1..86
                 note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
misc_feature     1..86
                 note = Prkdc #3 sgRNA
source          1..86
                 mol_type = other DNA
                 organism = synthetic construct

SEQUENCE: 38
gaaattaata cgactcacta taggcacaag caaaccaaag tctcgttta gagctagaaa 60
tagcaagtta aaataaggct agtccg 86

SEQ ID NO: 39      moltype = DNA length = 86
FEATURE          Location/Qualifiers
misc_feature     1..86
                 note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
misc_feature     1..86
                 note = Prkdc #4 sgRNA
source          1..86
                 mol_type = other DNA
                 organism = synthetic construct

SEQUENCE: 39
gaaattaata cgactcacta taggcctcaa tgctaagcga cttcgttta gagctagaaa 60
tagcaagtta aaataaggct agtccg 86

SEQ ID NO: 40      moltype = DNA length = 29
FEATURE          Location/Qualifiers
misc_feature     1..29
                 note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
misc_feature     1..29
                 note = F1 primer for Foxn1
source          1..29
                 mol_type = other DNA
                 organism = synthetic construct

SEQUENCE: 40
gtctgtctat catctcttcc cttctctcc 29

SEQ ID NO: 41      moltype = DNA length = 25
FEATURE          Location/Qualifiers
misc_feature     1..25
                 note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
misc_feature     1..25
                 note = F2 primer for Foxn1
source          1..25
                 mol_type = other DNA
                 organism = synthetic construct

SEQUENCE: 41
tccctaattcc gatggctagc tccag 25

SEQ ID NO: 42      moltype = DNA length = 23
FEATURE          Location/Qualifiers
misc_feature     1..23
                 note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
misc_feature     1..23
                 note = R1 primer for Foxn1
source          1..23
                 mol_type = other DNA
                 organism = synthetic construct

SEQUENCE: 42
acgagcagct gaagtagca tgc 23

SEQ ID NO: 43      moltype = DNA length = 32
FEATURE          Location/Qualifiers
misc_feature     1..32
                 note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
misc_feature     1..32

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source note = R2 primer for Foxnl
 1..32
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 43
 ctactcaatg ctcttagagc taccaggctt gc 32

SEQ ID NO: 44 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

misc_feature 1..20
 note = F primer for Prkdc

source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 44
 gactgttgtg gggagggccg 20

SEQ ID NO: 45 moltype = DNA length = 24
 FEATURE Location/Qualifiers
 misc_feature 1..24
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

misc_feature 1..24
 note = F2 primer for Prkdc

source 1..24
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 45
 gggagggccg aaagtcttat tttg 24

SEQ ID NO: 46 moltype = DNA length = 28
 FEATURE Location/Qualifiers
 misc_feature 1..28
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

misc_feature 1..28
 note = R1 primer for Prkdc

source 1..28
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 46
 cctgaagact gaagttggca gaagtgag 28

SEQ ID NO: 47 moltype = DNA length = 27
 FEATURE Location/Qualifiers
 misc_feature 1..27
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

misc_feature 1..27
 note = R2 primer for Prkdc

source 1..27
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 47
 ctttaggct tcttctctac aatcacg 27

SEQ ID NO: 48 moltype = DNA length = 38
 FEATURE Location/Qualifiers
 misc_feature 1..38
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

misc_feature 1..38
 note = F primer for Foxnl

source 1..38
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 48
 ctcggtgtg agccctgacc tcgggtgta gccctgac 38

SEQ ID NO: 49 moltype = DNA length = 21
 FEATURE Location/Qualifiers
 misc_feature 1..21
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide

misc_feature 1..21
 note = R primer for Foxnl

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source 1..21
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 49
 agactggcct ggaactcaca g 21

SEQ ID NO: 50 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..23
 note = F primer for Foxn1
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 50
 cactaaagcc tgtcaggaag ccg 23

SEQ ID NO: 51 moltype = DNA length = 21
 FEATURE Location/Qualifiers
 misc_feature 1..21
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..21
 note = R primer for Foxn1
 source 1..21
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 51
 ctgtggagag cacacagcag c 21

SEQ ID NO: 52 moltype = DNA length = 19
 FEATURE Location/Qualifiers
 misc_feature 1..19
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..19
 note = F primer for Foxn1
 source 1..19
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 52
 gctgcgacct gagaccatg 19

SEQ ID NO: 53 moltype = DNA length = 26
 FEATURE Location/Qualifiers
 misc_feature 1..26
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..26
 note = R primer for Foxn1
 source 1..26
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 53
 ctccaatggc ttctgctta ggctac 26

SEQ ID NO: 54 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..23
 note = F primer for Foxn1
 source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 54
 ggttcagatg aggccatcct ttc 23

SEQ ID NO: 55 moltype = DNA length = 24
 FEATURE Location/Qualifiers
 misc_feature 1..24
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..24
 note = R primer for Foxn1
 source 1..24

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mol_type = other DNA
organism = synthetic construct
SEQUENCE: 55
cctgatctgc aggcttaacc cttg                24

SEQ ID NO: 56      moltype = DNA length = 22
FEATURE           Location/Qualifiers
misc_feature      1..22
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..22
                  note = F primer for Prkdc
source           1..22
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 56
ctcacctgca catcacatgt gg                    22

SEQ ID NO: 57      moltype = DNA length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..20
                  note = R primer for Prkdc
source           1..20
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 57
ggcatccacc ctatggggtc                       20

SEQ ID NO: 58      moltype = DNA length = 25
FEATURE           Location/Qualifiers
misc_feature      1..25
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..25
                  note = F primer for Prkdc
source           1..25
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 58
gccttgacct agagcttaaa gagcc                 25

SEQ ID NO: 59      moltype = DNA length = 25
FEATURE           Location/Qualifiers
misc_feature      1..25
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..25
                  note = R primer for Prkdc
source           1..25
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 59
ggtcttgta gcaggaagga cactg                  25

SEQ ID NO: 60      moltype = DNA length = 27
FEATURE           Location/Qualifiers
misc_feature      1..27
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..27
                  note = F primer for Prkdc
source           1..27
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 60
aaaactctgc ttgatgggat atgtggg              27

SEQ ID NO: 61      moltype = DNA length = 26
FEATURE           Location/Qualifiers
misc_feature      1..26
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..26
                  note = R primer for Prkdc
source           1..26
                  mol_type = other DNA

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organism = synthetic construct

SEQUENCE: 61
ctctcaactgg ttatctgtgc tccttc 26

SEQ ID NO: 62 moltype = DNA length = 23
FEATURE Location/Qualifiers
misc_feature 1..23
note = Description of Artificial Sequence: Synthetic
oligonucleotide
misc_feature 1..23
note = F primer for Prkdc
source 1..23
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 62
ggatcaatag gtggtggggg atg 23

SEQ ID NO: 63 moltype = DNA length = 27
FEATURE Location/Qualifiers
misc_feature 1..27
note = Description of Artificial Sequence: Synthetic
oligonucleotide
misc_feature 1..27
note = R primer for Prkdc
source 1..27
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 63
gtgaatgaca caatgtgaca gcttcag 27

SEQ ID NO: 64 moltype = DNA length = 28
FEATURE Location/Qualifiers
misc_feature 1..28
note = Description of Artificial Sequence: Synthetic
oligonucleotide
misc_feature 1..28
note = F primer for Prkdc
source 1..28
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 64
cacaagacag acctctcaac attcagtc 28

SEQ ID NO: 65 moltype = DNA length = 32
FEATURE Location/Qualifiers
misc_feature 1..32
note = Description of Artificial Sequence: Synthetic
oligonucleotide
misc_feature 1..32
note = R primer for Prkdc
source 1..32
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 65
gtgcatgcat ataatccatt ctgattgctc tc 32

SEQ ID NO: 66 moltype = DNA length = 17
FEATURE Location/Qualifiers
misc_feature 1..17
note = Description of Artificial Sequence: Synthetic
oligonucleotide
misc_feature 1..17
note = F1 primer for Prkdc
source 1..17
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 66
gggaggcaga ggcaggt 17

SEQ ID NO: 67 moltype = DNA length = 23
FEATURE Location/Qualifiers
misc_feature 1..23
note = Description of Artificial Sequence: Synthetic
oligonucleotide
misc_feature 1..23
note = F2 primer for Prkdc
source 1..23
mol_type = other DNA
organism = synthetic construct

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SEQUENCE: 67
ggatctctgt gagtttgagg cca 23

SEQ ID NO: 68 moltype = DNA length = 24
FEATURE Location/Qualifiers
misc_feature 1..24
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
misc_feature 1..24
 note = R1 primer for Prkdc
source 1..24
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 68
gctccagaac tcactcttag gctc 24

SEQ ID NO: 69 moltype = DNA length = 20
FEATURE Location/Qualifiers
misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
misc_feature 1..20
 note = Primer for Foxn1
source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 69
ctactccctc cgcagtctga 20

SEQ ID NO: 70 moltype = DNA length = 20
FEATURE Location/Qualifiers
misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
misc_feature 1..20
 note = Primer for Foxn1
source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 70
ccaggcctag gttccaggta 20

SEQ ID NO: 71 moltype = DNA length = 20
FEATURE Location/Qualifiers
misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
misc_feature 1..20
 note = Primer for Prkdc
source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 71
ccccagcatt gcagatttcc 20

SEQ ID NO: 72 moltype = DNA length = 23
FEATURE Location/Qualifiers
misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
misc_feature 1..23
 note = Primer for Prkdc
source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 72
agggttctt ctctacaatc acg 23

SEQ ID NO: 73 moltype = DNA length = 86
FEATURE Location/Qualifiers
misc_feature 1..86
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
misc_feature 1..86
 note = BR11 target 1
source 1..86
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 73

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gaaattaata cgactcacta taggtttgaa agatggaagc gcggtttta gagctagaaa 60
tagcaagtta aaataaggct agtccg 86

SEQ ID NO: 74      moltype = DNA length = 86
FEATURE          Location/Qualifiers
misc_feature     1..86
                 note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
misc_feature     1..86
                 note = BRI1 target 2
source          1..86
                 mol_type = other DNA
                 organism = synthetic construct

SEQUENCE: 74
gaaattaata cgactcacta taggtgaaac taaactggtc cacagtttta gagctagaaa 60
tagcaagtta aaataaggct agtccg 86

SEQ ID NO: 75      moltype = DNA length = 64
FEATURE          Location/Qualifiers
misc_feature     1..64
                 note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
misc_feature     1..64
                 note = Universal
source          1..64
                 mol_type = other DNA
                 organism = synthetic construct

SEQUENCE: 75
aaaaaagcac cgactcggcg ccactttttc aagttgataa cggactagcc ttattttaac 60
ttgc 64

SEQ ID NO: 76      moltype = DNA length = 65
FEATURE          Location/Qualifiers
misc_feature     1..65
                 note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
misc_feature     1..65
                 note = Templates for crRNA
misc_difference 25..44
                 note = modified_base - a, c, t, g, unknown or other
source          1..65
                 mol_type = other DNA
                 organism = synthetic construct

SEQUENCE: 76
gaaattaata cgactcacta taggnnnnnn nnnnnnnnnn nnnngtttta gagctatgct 60
gtttt 65

SEQ ID NO: 77      moltype = DNA length = 67
FEATURE          Location/Qualifiers
misc_feature     1..67
                 note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
misc_feature     1..67
                 note = tracrRNA
source          1..67
                 mol_type = other DNA
                 organism = synthetic construct

SEQUENCE: 77
gaaattaata cgactcacta taggaacct tcaaacagc atagcaagtt aaaataaggc 60
tagtccg 67

SEQ ID NO: 78      moltype = DNA length = 69
FEATURE          Location/Qualifiers
misc_feature     1..69
                 note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
misc_feature     1..69
                 note = tracrRNA
source          1..69
                 mol_type = other DNA
                 organism = synthetic construct

SEQUENCE: 78
aaaaaaagca cggactcggg gccacttttt caagttgata acggactagc cttattttaa 60
cttgctatg 69

SEQ ID NO: 79      moltype = DNA length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
                 note = Description of Artificial Sequence: Synthetic

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misc_feature      oligonucleotide
                  1..20
                  note = Primer
source            1..20
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 79
ctccatggtg ctatagagca                               20

SEQ ID NO: 80      moltype = DNA length = 21
FEATURE           Location/Qualifiers
misc_feature      1..21
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..21
                  note = Primer
source            1..21
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 80
gagccaagct ctccatctag t                             21

SEQ ID NO: 81      moltype = DNA length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..20
                  note = Primer
source            1..20
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 81
gccctgtcaa gagttgacac                               20

SEQ ID NO: 82      moltype = DNA length = 22
FEATURE           Location/Qualifiers
misc_feature      1..22
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..22
                  note = Primer
source            1..22
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 82
gcacagggtg gaacaagatg ga                            22

SEQ ID NO: 83      moltype = DNA length = 24
FEATURE           Location/Qualifiers
misc_feature      1..24
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..24
                  note = Primer
source            1..24
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 83
gccaggtacc tatcgattgt cagg                           24

SEQ ID NO: 84      moltype = DNA length = 21
FEATURE           Location/Qualifiers
misc_feature      1..21
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature      1..21
                  note = Primer
source            1..21
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 84
gagccaagct ctccatctag t                             21

SEQ ID NO: 85      moltype = DNA length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide

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misc_feature      1..20
                  note = Primer
source            1..20
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 85
actctgactg ggtcaccagc                               20

SEQ ID NO: 86    moltype = DNA length = 20
FEATURE         Location/Qualifiers
misc_feature    1..20
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature    1..20
                  note = Primer
source          1..20
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 86
tatttggctg gttgaaaggg                               20

SEQ ID NO: 87    moltype = DNA length = 24
FEATURE         Location/Qualifiers
misc_feature    1..24
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature    1..24
                  note = Primer
source          1..24
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 87
aaagtcatga aataaacaca ccca                           24

SEQ ID NO: 88    moltype = DNA length = 24
FEATURE         Location/Qualifiers
misc_feature    1..24
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature    1..24
                  note = Primer
source          1..24
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 88
ctgcattgat atggtagtac catg                            24

SEQ ID NO: 89    moltype = DNA length = 21
FEATURE         Location/Qualifiers
misc_feature    1..21
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature    1..21
                  note = Primer
source          1..21
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 89
gctgttcatt gcaatggaat g                               21

SEQ ID NO: 90    moltype = DNA length = 22
FEATURE         Location/Qualifiers
misc_feature    1..22
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature    1..22
                  note = Primer
source          1..22
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 90
atggagtggg acatggccat gg                               22

SEQ ID NO: 91    moltype = DNA length = 28
FEATURE         Location/Qualifiers
misc_feature    1..28
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
misc_feature    1..28

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source	note = Primer 1..28 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 91		
actcactatc cacagttcag catttacc		28
SEQ ID NO: 92	moltype = DNA length = 23 Location/Qualifiers	
FEATURE	1..23	
misc_feature	note = Description of Artificial Sequence: Synthetic oligonucleotide	
misc_feature	1..23	
source	note = Primer 1..23 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 92		
tggagatagc tgtcagcaac ttt		23
SEQ ID NO: 93	moltype = DNA length = 29 Location/Qualifiers	
FEATURE	1..29	
misc_feature	note = Description of Artificial Sequence: Synthetic oligonucleotide	
misc_feature	1..29	
source	note = Primer 1..29 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 93		
caacaaagca aaggtaaagt tggtaatag		29
SEQ ID NO: 94	moltype = DNA length = 25 Location/Qualifiers	
FEATURE	1..25	
misc_feature	note = Description of Artificial Sequence: Synthetic oligonucleotide	
misc_feature	1..25	
source	note = Primer 1..25 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 94		
ggtttcagga gatgtgttac aaggc		25
SEQ ID NO: 95	moltype = DNA length = 27 Location/Qualifiers	
FEATURE	1..27	
misc_feature	note = Description of Artificial Sequence: Synthetic oligonucleotide	
misc_feature	1..27	
source	note = Primer 1..27 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 95		
gattgtgcaa ttcctatgca atcggtc		27
SEQ ID NO: 96	moltype = DNA length = 25 Location/Qualifiers	
FEATURE	1..25	
misc_feature	note = Description of Artificial Sequence: Synthetic oligonucleotide	
misc_feature	1..25	
source	note = Primer 1..25 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 96		
cactgggtac ttaatctgta gcctc		25
SEQ ID NO: 97	moltype = DNA length = 23 Location/Qualifiers	
FEATURE	1..23	
misc_feature	note = Description of Artificial Sequence: Synthetic oligonucleotide	
misc_feature	1..23	
	note = Primer	

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source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 97
 ggttccaagt cattcccagt agc 23

SEQ ID NO: 98 moltype = DNA length = 30
 FEATURE Location/Qualifiers
 misc_feature 1..30
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..30
 note = Primer
 source 1..30
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 98
 catcactgca gttgtaggtt ataactatcc 30

SEQ ID NO: 99 moltype = DNA length = 26
 FEATURE Location/Qualifiers
 misc_feature 1..26
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..26
 note = Primer
 source 1..26
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 99
 ttgaaaacca cagatctggt tgaacc 26

SEQ ID NO: 100 moltype = DNA length = 22
 FEATURE Location/Qualifiers
 misc_feature 1..22
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..22
 note = Primer
 source 1..22
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 100
 ggagtgccaa gagaatatct gg 22

SEQ ID NO: 101 moltype = DNA length = 32
 FEATURE Location/Qualifiers
 misc_feature 1..32
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..32
 note = Primer
 source 1..32
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 101
 ctgaaactgg tttcaaaata ttcgttttaa gg 32

SEQ ID NO: 102 moltype = DNA length = 22
 FEATURE Location/Qualifiers
 misc_feature 1..22
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..22
 note = Primer
 source 1..22
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 102
 gctctgtatg ccctgtagta gg 22

SEQ ID NO: 103 moltype = DNA length = 22
 FEATURE Location/Qualifiers
 misc_feature 1..22
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 misc_feature 1..22
 note = Primer
 source 1..22

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mol_type = other DNA
organism = synthetic construct
SEQUENCE: 103
ttgcatctg accttacctt tg 22

SEQ ID NO: 104      moltype = DNA length = 23
FEATURE            Location/Qualifiers
misc_feature        1..23
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
misc_feature        1..23
                    note = Target sequence of RGEN
source              1..23
                    mol_type = other DNA
                    organism = synthetic construct

SEQUENCE: 104
aatgaccact acatcctcaa ggg 23

SEQ ID NO: 105      moltype = DNA length = 23
FEATURE            Location/Qualifiers
misc_feature        1..23
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
misc_feature        1..23
                    note = Target sequence of RGEN
source              1..23
                    mol_type = other DNA
                    organism = synthetic construct

SEQUENCE: 105
agatgatgtc tcatcatcag agg 23

SEQ ID NO: 106      moltype = DNA length = 4170
FEATURE            Location/Qualifiers
misc_feature        1..4170
                    note = Description of Artificial Sequence: Synthetic
                    polynucleotide
misc_feature        1..4170
                    note = Cas9-coding sequence in p3s-Cas9HC (humanized,
                    C-term tagging, human cell experiments)
source              1..4170
                    mol_type = other DNA
                    organism = synthetic construct

SEQUENCE: 106
atggacaaga agtacagcat cggcctggac atcgggtacca acagcgtggg ctgggcccgtg 60
atcaccgcag agtacaaggt gccccagcaag aagttcaagg tgctgggcaa caccgaccgc 120
cacagcatca agaagaacct gatcggcgcc ctgctgttcg acagcggcga gaccgcgcag 180
gccaccgcc tgaagcgcac cgcccgccgc cgctacacc gccgcaagaa ccgcatctgc 240
tacctgcagg agatcttcag caacgagatg gccaaaggtgg acgacagctt cttccaccgc 300
ctggaggaga gcttcctggg ggaggaggac aagaagcacg agcggcacc catcttcggc 360
aacatcgtgg acgaggtggc ctaccacgag aagtacccca ccatctacca cctgcgcaag 420
aagctggtgg acagcaccga caaggccgac ctgctgctga tctacctggc cctggcccac 480
atgatcaagt tccgcgcca cttcctgatc gagggcgacc tgaaccgca caacagcgac 540
gtggacaagc tgttcatcca gctggtgcag acctacaacc agctgttcga ggagaacccc 600
atcaacgcca gccgctgga cgccaaggcc atcctgagcg cccgcctgag caagagccgc 660
cgcctggaga acctcgatcg ccaagctgccc ggccgagaaga agaacggcct gttcggcaac 720
ctgatcgccc tgaacctggg cctgaccccc aactcaaga gcaacttga cctggccgag 780
gacgccaagc tgcagctgag caaggacacc tacgacgacg acctggacaa cctgctggcc 840
cagatcggcg accagtacgc cgacctgttc ctggccgcca agaacctgag cgacgccatc 900
ctgctgagcg acatcctgcg cgtgaacacc gagatcacca agggccccct gagcggcagc 960
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aagcagcgca ccttcgacaa cggcagcatc ccccaccaga tccacctggg cgagctgcac 1260
gccatcctgc gcccccagga ggacttctac cccttctga aggacaaccg cgagaagatc 1320
gagaagatcc tgaccttcgg catcccctac tacgtgggccc ccctggcccgc cggaacagc 1380
cgcttcgcct gtagtaccgc caagagcggg gagaccatca ccccctggaa cttcgaggag 1440
gtggtggaca agggcgccag cggccagagc ttcctcgagc gcatgaccaa cttcgacaag 1500
aacctgcccc acgagaaggt gctgcccagg cacagcctgc tgtacgagta cttcaccgtg 1560
tacaacgagc tgaccaaggt gaagtacgtg accgaggcca tgcgcaagcc cgccttctctg 1620
agcggcgagc agaagaaggg catcgtggac ctgctgttca agaccaaccg caaggtgacc 1680
gtgaagcagc tgaaggagga ctacttcaag aagatcgagt gcttcgacag cgtggagatc 1740
agcggcgctg aggaccgctt caacgcccgc ctgggcaact accacgacct gctgaagatc 1800
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ctgaccctga cccgtgttga ggaccgcgag atgatcgagg agcgcctgaa gacctacgcc 1920
cacctgttcg acgacaaggt gatgaagcag ctgaagcgcg gccgctacac cggctggggc 1980
cgctgagacc gcaagcttat caacggcatc cgcgacaagc agagcggcaa gacctcctg 2040
gacttctga agagcgacgc cttcgccaac cgcaacttca tgcagctgat ccacgacgac 2100
agcctgacct tcaaggagga catccagaag gcccaaggtga gcgccaggcg cgacagcctg 2160

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cacgagcaca tgcgcaacct ggcggcagc cccgccatca agaagggcat cctgcagacc 2220
gtgaaggtgg tggacagact ggtgaaggtg atgggcccgc acaagcccga gaacatcgtg 2280
atcgagatgg cccgagagaa ccagaccacc cagaagggcc agaagaacag ccgagagcgc 2340
atgaagcgca tgcaggaggg catcaaggag ctgggcagcc agatcctgaa ggagcacc 2400
gtggagaaca cccagctgca gaacgagaag ctgtacctgt actacctgca gaacggccgc 2460
gacatgtacg tggaccagga gctggacatc aaccgcctga gcgactacga cgtggaccac 2520
atcgtgcccc agagcttctt gaaggacgac agcatcgaca acaaggtgct gacccgcagc 2580
gacaagaacc gcggcaagag cgacaacgtg cccagcgagg aggtggtgaa gaagatgaa 2640
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taccccaagc tggagagcga gttcgtgtac ggcgactaca aggtgtacga cgtgcccag 3060
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gcccgaaga aggactggga cccaagaag tacggcggct tcgacagccc caccgtggcc 3420
tacagcgtgc tgggtggtgg caaggtggag aaggccaaga gcaagaagt gaagagcgtg 3480
aaggagctgc tgggcatcac catcatggag cgcagcagct tcgagaagaa ccccatgac 3540
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tacagcctgt tcgagctgga caacggccgc aagcgcctgc tggccagcgc cggcgagctg 3660
cagaagggca cagactgggc cctgccagc aagtacgtga acttctctga cctggcccagc 3720
cactacgaga agctgaaggg cagccccgag gacaacgagc agaagcagct gttcgtggag 3780
cagcacaagc actacctgga ggagatcatc gagcagatca gcgagttcac caagcgcgtg 3840
atcctggccg agcccaacct ggacaaggtg ctgagcgcct acaacaagca ccgagacaag 3900
cccatccgcg agcaggccga gaacatcatc cactctgtca ccctgaccaa cctggggcgc 3960
cccgcgcct tcaagtaact gacaccacc atcgaccgca agcgtacac cagcacaag 4020
gaggtgctgg gaccaccct gatccaccag agcatcacg gtctgtacga gaccgcctc 4080
gacctgagcc agctggggcg cgacggcggc tccggacctc caaagaaaa gagaaaagta 4140
taccctacg acgtgcccga ctacgcctaa 4170

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SEQ ID NO: 107      moltype = DNA length = 4194
FEATURE            Location/Qualifiers
misc_feature        1..4194
                    note = Description of Artificial Sequence: Synthetic
                    polynucleotide
misc_feature        1..4194
                    note = Cas9 coding sequence in p3s-Cas9HN (humanized codon,
                    N-term tagging (underlined), human cell experiments)
source              1..4194
                    mol_type = other DNA
                    organism = synthetic construct

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SEQUENCE: 107
atggtgtacc cctacgactg gcccgactac gccgaattgc tccaaaaaa gaagagaag 60
gtagggatcc gaattcccgg ggaaaaaccg gacaagaagt acagcatcgg cctggacatc 120
ggtaccaaca gctgtggcgt ggccgtgatc accgacagat acaaggtgcc cagcaagaag 180
ttcaaggtgc tgggcaacac cgaccgccac agcatcaaga agaacctgat cggcgcctcg 240
ctggtcgaca gcgcgagacc cccgagggcc accgcctga agcgcaccgc ccgcccgcgc 300
tacaccgcc ccaagaaccg catctgctac ctgcaggaga tcttcagcaa cgagatggcc 360
aaggtggacg acagcttctt ccaccgcctg gaggagagct tcctggtgga ggaggacaag 420
aagcacgagc gccaccacct ctccggcaac atcgtggacg aggtggccta ccacgagaag 480
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SEQ ID NO: 108          moltype = DNA length = 4107
FEATURE
misc_feature           1..4107
                        note = Description of Artificial Sequence: Synthetic
                        polynucleotide
misc_feature           1..4107
                        note = Cas9-coding sequence in Streptococcus pyogenes
source                 1..4107
                        mol_type = other DNA
                        organism = synthetic construct

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SEQ ID NO: 109 moltype = AA length = 1368
FEATURE Location/Qualifiers
REGION 1..1368
note = Description of Artificial Sequence: Synthetic
polypeptide
REGION 1..1368
note = Amino acid sequence of Cas9 from S.pyogenes
source 1..1368
mol_type = protein
organism = synthetic construct

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RLSRKLINGI RDKQSGKTIL DFLKSDGFAN RNFMQLIHDD SLTFKEDIQK AQVSGQGDLS 720
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SEQ ID NO: 110 moltype = DNA length = 4221
FEATURE Location/Qualifiers
misc_feature 1..4221

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note = Description of Artificial Sequence: Synthetic
      polynucleotide
misc_feature 1..4221
      note = Cas9-coding sequence in pET-Cas9N3T for the
            production of recombinant Cas9 protein in E. coli
            (humanized codon; hexa-His-tag and a nuclear localization
            signal at the N terminus)
source 1..4221
      mol_type = other DNA
      organism = synthetic construct

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SEQUENCE: 110

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ggcaacgagc tggccctgcc cgcgaagtac gtgaacttcc tgtacctggc cagccactac 3840
gagaagctga agggcagccc cgaggacaac gagcagaagc agctgttcgt ggagcagcac 3900
aagcactacc tggacgagat catcgagcag atcagcgagt tcagcaagcg cgtgatcctg 3960
gcccagccca acctggacaa ggtgctgagc gcctacaaca agcaccgcca caagcccatc 4020
cgcgagcagg ccgagaacat catccaactg ttcacctgca ccaacctggg cgcccccgcc 4080

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gccttcaagt acttcgacac caccatcgac cgcaagcgct acaccagcac caaggaggtg 4140
ctggacgccca ccctgatoca ccagagcadc accggtctgt acgagacccg catcgacctg 4200
agccagctgg gcgcgcgacta a 4221

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SEQ ID NO: 111      moltype = AA length = 1406
FEATURE           Location/Qualifiers
REGION           1..1406
                  note = Description of Artificial Sequence: Synthetic
                  polypeptide
REGION           1..1406
                  note = Amino acid sequence of Cas9 (pET-Cas9N3T)
source           1..1406
                  mol_type = protein
                  organism = synthetic construct

```

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SEQUENCE: 111
MGSSHHHHHH VPPYDVPDYA ELPPKKRKRK GIENLYFQGD KKYSIGLDIG TNSVGWAVIT 60
DEYKVPKSKF KVLGNTDRHS IKKNLIGALL FDSGETAEAT RLKRTARRRY TRRKNRICYL 120
QEIFSNEMAK VDDSFPHRLE ESFLVEEDKK HERHPIFGNI VDEVAYHEKY PTIYHLRKKL 180
VDSTDKADLR LIYLALAHMI KFRGHPLIEG DLNPDNSDWD KLFIQLVQTY NQLFEENPIN 240
ASGVDAKAIL SARLSKSRRL ENLIAQLPGE KKNGLFGNLI ALSLGLTPNF KSNFDLAEDA 300
KLQLSKDTYD DDLNLLAQI GDQYADLFLA AKNLSDAILL SDILRVNTEI TKAPLSASMI 360
KRYDEHHQDL TLLKALVRQQ LPEKYKEIFF DQSKNGYAGY IDGGASQEEF YKFIKPILEK 420
MDGTEELLVK LNRELLLRKQ RTFDNGSIPH QIHLGELHAI LRRQEDFYPP LKDNREKIEK 480
LLTFRIPYYV GPLARGNSRF AWMTRKSEET ITPWNFEEV DKGASAQFSI ERMNPFDKNL 540
PNEKVLPKHS LLYEYFTVYN ELTKVKYVTE GMRKPAFLSG EQKKAIVDLL FKTNRKVTVK 600
QLKEDYFKKI ECFDSVEISG VEDRFNASLG TYHDLLKIIK DKDFLDNEEN EDILEDIVLT 660
LTLFEDREMI EERLKYAHL FDDKVMKQLK RRRYTGWGR LSRKLINGIRD KQSGKTILDF 720
LKSDGFANRN FMQLIHDDSL TFKEDIQKAQ VSGQGDLSHE HIANLAGSPA IKKGILQTVK 780
VVDLVLKVMG RHKPENIVIE MARENQTTQK GQKNSRERMK RIEEGIKELG SQILKEHPVE 840
NTQLQNEKLY LYYLQNGRDM YVQQLDINR LSDYDVDHIV PQSFLKDDSI DNKVLTRSDK 900
NRGKSDNVPS EEVVKMKMNY WRQLLNAKLI TQRKFDNLTK AERGGLSELD KAGFIKRLV 960
ETRQITKHVA QILDSRMNTK YDENDKLIRE VKVITLKS KL VSDFRKDFQF YKVINNNYH 1020
HAHDAYLNAV VGTALIKKYP KLESEFVYGD YKVDVVRMI AKSQEIGKA TAKYFPYSNI 1080
MNFPKTEITL ANGEIRKRPL IETNGETGEI VWDKGRDPAT VRKVLSPQV NIVKKTVEVQT 1140
GGFSKESILP KRNSDKLIAR KKDWDPKKYG GFDSPTVAYS VLVVAKVEKG KSKKLKSVKE 1200
LLGITIMERS SFEKNPDPL EAKGYKEVKK DLI IKLPKYS LPELENGRKR MLASAGELQK 1260
GNELALPSKY VNFYLYLASHY EKLGSPEDN EQQLFVEQH KHYLDEIEQ ISEFSKRVL 1320
ADANLDKVL S AYNKHRDKPI REQAENIIHL FTLTNLGAPA AFKYFDTTID RKRYTSTKEV 1380
LDATLIHQSI TGLYETRIDL SQLGGD 1406

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SEQ ID NO: 112      moltype = DNA length = 34
FEATURE           Location/Qualifiers
source           1..34
                  mol_type = unassigned DNA
                  organism = Homo sapiens

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SEQUENCE: 112
caatctatga catcaattat tatacatcgg agcc 34

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```

SEQ ID NO: 113      moltype = RNA length = 64
FEATURE           Location/Qualifiers
misc_feature     1..64
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source           1..64
                  mol_type = other RNA
                  organism = synthetic construct

```

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SEQUENCE: 113
ggtgacatca attattatac atgttttaga gctagaataa gcaagttaaa ataaggctag 60
tccg 64

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SEQ ID NO: 114      moltype = DNA length = 49
FEATURE           Location/Qualifiers
misc_feature     1..49
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source           1..49
                  mol_type = other DNA
                  organism = synthetic construct

```

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SEQUENCE: 114
caatctatga catcaattat tatacatcgg agccctgccca aaaaatcaa 49

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```

SEQ ID NO: 115      moltype = DNA length = 50
FEATURE           Location/Qualifiers
misc_feature     1..50
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source           1..50
                  mol_type = other DNA
                  organism = synthetic construct

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SEQUENCE: 115
caatctatga catcaattat tataacatcg gagccctgcc aaaaaatcaa 50

SEQ ID NO: 116 moltype = DNA length = 36
FEATURE Location/Qualifiers
misc_feature 1..36
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..36
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 116
caatctatga catcaattat tatgccaaaa aatcaa 36

SEQ ID NO: 117 moltype = DNA length = 35
FEATURE Location/Qualifiers
misc_feature 1..35
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..35
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 117
caatctatga catcgagacc ctgccaaaa atcaa 35

SEQ ID NO: 118 moltype = DNA length = 31
FEATURE Location/Qualifiers
misc_feature 1..31
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..31
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 118
caatctatga catgccctgc caaaaaatca a 31

SEQ ID NO: 119 moltype = DNA length = 30
FEATURE Location/Qualifiers
misc_feature 1..30
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..30
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 119
caatctatga catcaattat tataaatcaa 30

SEQ ID NO: 120 moltype = DNA length = 25
FEATURE Location/Qualifiers
misc_feature 1..25
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..25
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 120
caatctatga catccaaaa atcaa 25

SEQ ID NO: 121 moltype = DNA length = 19
FEATURE Location/Qualifiers
misc_feature 1..19
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..19
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 121
caatctatga caaaatcaa 19

SEQ ID NO: 122 moltype = DNA length = 46
FEATURE Location/Qualifiers
source 1..46
mol_type = unassigned DNA
organism = Homo sapiens

SEQUENCE: 122
tatgtgcaat gaccactaca tcctcaaggg cagcaatcgg agccag 46

SEQ ID NO: 123 moltype = DNA length = 47
FEATURE Location/Qualifiers

-continued

misc_feature	1..47		
		note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..47		
		mol_type = other DNA	
		organism = synthetic construct	
SEQUENCE: 123			
tatgtgcaat gaccactaca tccttcaagg gcagcaatcg gagccag			47
SEQ ID NO: 124		moltype = DNA length = 48	
FEATURE		Location/Qualifiers	
misc_feature	1..48		
		note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..48		
		mol_type = other DNA	
		organism = synthetic construct	
SEQUENCE: 124			
tatgtgcaat gaccactaca tcctctcaag ggcagcaatc ggagccag			48
SEQ ID NO: 125		moltype = DNA length = 18	
FEATURE		Location/Qualifiers	
misc_feature	1..18		
		note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..18		
		mol_type = other DNA	
		organism = synthetic construct	
SEQUENCE: 125			
tatgtgcaat ggagccag			18
SEQ ID NO: 126		moltype = DNA length = 13	
FEATURE		Location/Qualifiers	
misc_feature	1..13		
		note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..13		
		mol_type = other DNA	
		organism = synthetic construct	
SEQUENCE: 126			
tatgtgcaat gac			13
SEQ ID NO: 127		moltype = DNA length = 23	
FEATURE		Location/Qualifiers	
source	1..23		
		mol_type = unassigned DNA	
		organism = Homo sapiens	
SEQUENCE: 127			
tgacatcaat tattatacat cgg			23
SEQ ID NO: 128		moltype = DNA length = 23	
FEATURE		Location/Qualifiers	
source	1..23		
		mol_type = unassigned DNA	
		organism = Homo sapiens	
SEQUENCE: 128			
tgacatcaat tattatagat gga			23
SEQ ID NO: 129		moltype = DNA length = 23	
FEATURE		Location/Qualifiers	
source	1..23		
		mol_type = unassigned DNA	
		organism = Homo sapiens	
SEQUENCE: 129			
tgacatcact tattatgcat ggg			23
SEQ ID NO: 130		moltype = DNA length = 23	
FEATURE		Location/Qualifiers	
source	1..23		
		mol_type = unassigned DNA	
		organism = Homo sapiens	
SEQUENCE: 130			
tgacataaat tattctacat ggg			23
SEQ ID NO: 131		moltype = DNA length = 23	
FEATURE		Location/Qualifiers	
source	1..23		
		mol_type = unassigned DNA	
		organism = Homo sapiens	

-continued

SEQUENCE: 131
 tgaaatcaat tatcatagat cgg 23

SEQ ID NO: 132 moltype = DNA length = 23
 FEATURE Location/Qualifiers
 source 1..23
 mol_type = unassigned DNA
 organism = Homo sapiens

SEQUENCE: 132
 ccaggctcca cccgactgga ggg 23

SEQ ID NO: 133 moltype = RNA length = 106
 FEATURE Location/Qualifiers
 misc_feature 1..106
 note = Description of Artificial Sequence: Synthetic
 polynucleotide
 source 1..106
 mol_type = other RNA
 organism = synthetic construct

SEQUENCE: 133
 ggccaggctc caccgactg gagttttaga gctagaaata gcaagttaaa ataaggctag 60
 tccgttatca acttgaaaaa gtggcaccga gtcggtgctt tttttt 106

SEQ ID NO: 134 moltype = DNA length = 53
 FEATURE Location/Qualifiers
 source 1..53
 mol_type = unassigned DNA
 organism = Homo sapiens

SEQUENCE: 134
 acttccaggc tccaccggac tggagggcga accccaaggg gacctcatgc agg 53

SEQ ID NO: 135 moltype = DNA length = 13
 FEATURE Location/Qualifiers
 misc_feature 1..13
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..13
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 135
 acttccaggc tcc 13

SEQ ID NO: 136 moltype = DNA length = 30
 FEATURE Location/Qualifiers
 misc_feature 1..30
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..30
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 136
 acttccaggc tccaccggac ctcatgcagg 30

SEQ ID NO: 137 moltype = DNA length = 36
 FEATURE Location/Qualifiers
 misc_feature 1..36
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..36
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 137
 acttccaggc tccaccctcaa ggggacctca tgcagg 36

SEQ ID NO: 138 moltype = DNA length = 54
 FEATURE Location/Qualifiers
 misc_feature 1..54
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..54
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 138
 acttccaggc tccaccggac ttggagggcg aaccctcagg ggacctcatg cagg 54

SEQ ID NO: 139 moltype = DNA length = 43
 FEATURE Location/Qualifiers
 misc_feature 1..43
 note = Description of Artificial Sequence: Synthetic

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source                oligonucleotide
                    1..43
                    mol_type = other DNA
                    organism = synthetic construct
SEQUENCE: 139
acttccaggc tccacccgaa cccaagggg acctcatgca ggg                43

SEQ ID NO: 140        moltype = DNA length = 47
FEATURE              Location/Qualifiers
misc_feature         1..47
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
source               1..47
                    mol_type = other DNA
                    organism = synthetic construct
SEQUENCE: 140
acttccaggc tccacccgac tcactatctt ctgggctcct ccatgtc                47

SEQ ID NO: 141        moltype = DNA length = 45
FEATURE              Location/Qualifiers
misc_feature         1..45
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
source               1..45
                    mol_type = other DNA
                    organism = synthetic construct
SEQUENCE: 141
acttccaggc tccacccgac gaacccaag gggacctcat gcagg                45

SEQ ID NO: 142        moltype = AA length = 18
FEATURE              Location/Qualifiers
source               1..18
                    mol_type = protein
                    organism = Homo sapiens
SEQUENCE: 142
LPGSTRLEGE PQGDLMQA                18

SEQ ID NO: 143        moltype = DNA length = 57
FEATURE              Location/Qualifiers
source               1..57
                    mol_type = unassigned DNA
                    organism = Homo sapiens
CDS                  2..55
SEQUENCE: 143
acttccaggc tccacccgac tggagggcga accccaaggg gacctcatgc aggctcc                57

SEQ ID NO: 144        moltype = DNA length = 46
FEATURE              Location/Qualifiers
misc_feature         1..46
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
source               1..46
                    mol_type = other DNA
                    organism = synthetic construct
SEQUENCE: 144
acttccaggc tccacccgaa cccaagggg acctcatgca ggctcc                46

SEQ ID NO: 145        moltype = DNA length = 43
FEATURE              Location/Qualifiers
misc_feature         1..43
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
source               1..43
                    mol_type = other DNA
                    organism = synthetic construct
SEQUENCE: 145
acttccaggc tccacccgaa cccaagggg acctcatgca ggc                43

SEQ ID NO: 146        moltype = DNA length = 20
FEATURE              Location/Qualifiers
misc_feature         1..20
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
source               1..20
                    mol_type = other DNA
                    organism = synthetic construct
SEQUENCE: 146
acttccaggc tccacccgac                20

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source	1..34	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 154		
acttccaggc gaacccaag gggacctcat gcag		34
SEQ ID NO: 155	moltype = DNA length = 32	
FEATURE	Location/Qualifiers	
misc_feature	1..32	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..32	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 155		
acttccaggc tccacaagg gacctcatgc ag		32
SEQ ID NO: 156	moltype = DNA length = 34	
FEATURE	Location/Qualifiers	
misc_feature	1..34	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..34	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 156		
acttccaggc tccaccaag gggacctcat gcc		34
SEQ ID NO: 157	moltype = DNA length = 35	
FEATURE	Location/Qualifiers	
misc_feature	1..35	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..35	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 157		
acttccaggc tccacccaa ggggacctca tgcag		35
SEQ ID NO: 158	moltype = DNA length = 41	
FEATURE	Location/Qualifiers	
misc_feature	1..41	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..41	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 158		
acttccaggc tccaccgaa cccaagggg acctcatgca g		41
SEQ ID NO: 159	moltype = DNA length = 50	
FEATURE	Location/Qualifiers	
misc_feature	1..50	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..50	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 159		
acttccaggc tccaccgaa ggaggcgaa cccaagggg acctcatgca		50
SEQ ID NO: 160	moltype = DNA length = 50	
FEATURE	Location/Qualifiers	
misc_feature	1..50	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..50	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 160		
acttccaggc tccaccgac tagggcgaa cccaagggga cctcatgcag		50
SEQ ID NO: 161	moltype = DNA length = 52	
FEATURE	Location/Qualifiers	
misc_feature	1..52	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..52	
	mol_type = other DNA	

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organism = synthetic construct
 SEQUENCE: 161
 acttcaggc tccaccgac tgggagggcg aacccaagg ggacctcatg ca 52

SEQ ID NO: 162 moltype = DNA length = 52
 FEATURE Location/Qualifiers
 misc_feature 1..52
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..52
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 162
 acttcaggc tccaccgac ttggagggcg aacccaagg ggacctcatg ca 52

SEQ ID NO: 163 moltype = DNA length = 46
 FEATURE Location/Qualifiers
 misc_feature 1..46
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..46
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 163
 acttcaggc tccaccgag gcgaaccca aggggacctc atgcag 46

SEQ ID NO: 164 moltype = DNA length = 47
 FEATURE Location/Qualifiers
 misc_feature 1..47
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..47
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 164
 acttcaggc tccaccgag gggaacccc aaggggacct catgcag 47

SEQ ID NO: 165 moltype = DNA length = 24
 FEATURE Location/Qualifiers
 misc_feature 1..24
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..24
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 165
 acttcaggc tccacctcat gcag 24

SEQ ID NO: 166 moltype = DNA length = 29
 FEATURE Location/Qualifiers
 misc_feature 1..29
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..29
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 166
 agggcgaacc ccaaggggac ctcatgcag 29

SEQ ID NO: 167 moltype = DNA length = 45
 FEATURE Location/Qualifiers
 misc_feature 1..45
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..45
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 167
 caatctatga catcaattat tatcgagacc ctgccaaaaa atcaa 45

SEQ ID NO: 168 moltype = DNA length = 45
 FEATURE Location/Qualifiers
 misc_feature 1..45
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..45
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 168

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caatctatga catcaattat catcggagcc ctgccaaaaa atcaa 45

SEQ ID NO: 169 moltype = DNA length = 42
 FEATURE Location/Qualifiers
 misc_feature 1..42
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..42
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 169
 caatctatga catcaattat cggagccctg ccaaaaaatc aa 42

SEQ ID NO: 170 moltype = DNA length = 48
 FEATURE Location/Qualifiers
 misc_feature 1..48
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..48
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 170
 caatctatga catcaattat tatcatcgga gccctgccaa aaaatcaa 48

SEQ ID NO: 171 moltype = DNA length = 33
 FEATURE Location/Qualifiers
 misc_feature 1..33
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..33
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 171
 caatctatga caagagccct gccaaaaaat caa 33

SEQ ID NO: 172 moltype = DNA length = 52
 FEATURE Location/Qualifiers
 source 1..52
 mol_type = unassigned DNA
 organism = Homo sapiens

SEQUENCE: 172
 ttctcaaggc agcatcatac ttcccccaag gtgggacagc tgcctccct gg 52

SEQ ID NO: 173 moltype = DNA length = 46
 FEATURE Location/Qualifiers
 misc_feature 1..46
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..46
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 173
 ttctcaaggc agcatcatac ttccctggga cagctgccct ccctgg 46

SEQ ID NO: 174 moltype = DNA length = 49
 FEATURE Location/Qualifiers
 misc_feature 1..49
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..49
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 174
 ttctcaaggc agcatcatac ttccacggtg ggacagctgc cctccctgg 49

SEQ ID NO: 175 moltype = DNA length = 25
 FEATURE Location/Qualifiers
 misc_feature 1..25
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..25
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 175
 ttctcaaggc agctgccctc cctgg 25

SEQ ID NO: 176 moltype = DNA length = 32
 FEATURE Location/Qualifiers
 misc_feature 1..32

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                note = Description of Artificial Sequence: Synthetic
                oligonucleotide
source          1..32
                mol_type = other DNA
                organism = synthetic construct

SEQUENCE: 176
ttctcaaggc agcatcatac ttccctccct gg                               32

SEQ ID NO: 177      moltype = DNA length = 264
FEATURE            Location/Qualifiers
misc_difference    38..227
                note = modified_base - a, c, t, g, unknown or other
source            1..264
                mol_type = unassigned DNA
                organism = Homo sapiens

SEQUENCE: 177
acaaagcgat tttgaaagat ggaagcgcgg tggctatnnn nnnnnnnnnn nnnnnnnnnn 60
nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 120
nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 180
nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnggg gtgaaactaa 240
actggtccac acggcggaag attg                                       264

SEQ ID NO: 178      moltype = DNA length = 257
FEATURE            Location/Qualifiers
misc_feature      1..257
                note = Description of Artificial Sequence: Synthetic
                polynucleotide
misc_difference    38..227
                note = modified_base - a, c, t, g, unknown or other
source            1..257
                mol_type = other DNA
                organism = synthetic construct

SEQUENCE: 178
acaaagcgat tttgaaagat ggaagcgcgg tggctatnnn nnnnnnnnnn nnnnnnnnnn 60
nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 120
nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 180
nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnggg gtgaaactaa 240
aacacggcgg aagattg                                               257

SEQ ID NO: 179      moltype = DNA length = 43
FEATURE            Location/Qualifiers
misc_feature      1..43
                note = Description of Artificial Sequence: Synthetic
                oligonucleotide
source            1..43
                mol_type = other DNA
                organism = synthetic construct

SEQUENCE: 179
acaaagcgat tttgaaagat ggaagcgaca cggcggaaga ttg                               43

SEQ ID NO: 180      moltype = DNA length = 44
FEATURE            Location/Qualifiers
misc_feature      1..44
                note = Description of Artificial Sequence: Synthetic
                oligonucleotide
source            1..44
                mol_type = other DNA
                organism = synthetic construct

SEQUENCE: 180
acaaagcgat tttgaaagat ggaagcgcac acggcggaag attg                               44

SEQ ID NO: 181      moltype = DNA length = 106
FEATURE            Location/Qualifiers
misc_feature      1..106
                note = Description of Artificial Sequence: Synthetic
                polynucleotide
source            1..106
                mol_type = other DNA
                organism = synthetic construct

SEQUENCE: 181
acaaagcgat tttgaaagat ggaagcgaaa tagcaagtta aaataaggct agtccgttat 60
caactgaaa aagtgccacc gagtcggtgc acacggcgga agattg                    106

SEQ ID NO: 182      moltype = DNA length = 23
FEATURE            Location/Qualifiers
misc_feature      1..23
                note = Description of Artificial Sequence: Synthetic
                oligonucleotide
source            1..23

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mol_type = other DNA
organism = synthetic construct
SEQUENCE: 182
gggtgggggg agtttgcctc tgg 23

SEQ ID NO: 183      moltype = DNA length = 23
FEATURE            Location/Qualifiers
misc_feature        1..23
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
source              1..23
                    mol_type = other DNA
                    organism = synthetic construct
SEQUENCE: 183
ggatggaggg agtttgcctc tgg 23

SEQ ID NO: 184      moltype = DNA length = 23
FEATURE            Location/Qualifiers
misc_feature        1..23
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
source              1..23
                    mol_type = other DNA
                    organism = synthetic construct
SEQUENCE: 184
ggggagggga agtttgcctc tgg 23

SEQ ID NO: 185      moltype = DNA length = 23
FEATURE            Location/Qualifiers
misc_feature        1..23
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
source              1..23
                    mol_type = other DNA
                    organism = synthetic construct
SEQUENCE: 185
gacccctcc accccgcctc cgg 23

SEQ ID NO: 186      moltype = DNA length = 23
FEATURE            Location/Qualifiers
misc_feature        1..23
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
source              1..23
                    mol_type = other DNA
                    organism = synthetic construct
SEQUENCE: 186
gaccccccc accccgcccc cgg 23

SEQ ID NO: 187      moltype = DNA length = 23
FEATURE            Location/Qualifiers
misc_feature        1..23
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
source              1..23
                    mol_type = other DNA
                    organism = synthetic construct
SEQUENCE: 187
gccccaccc accccgcctc tgg 23

SEQ ID NO: 188      moltype = DNA length = 23
FEATURE            Location/Qualifiers
misc_feature        1..23
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
source              1..23
                    mol_type = other DNA
                    organism = synthetic construct
SEQUENCE: 188
ctccccacc accccgcctc agg 23

SEQ ID NO: 189      moltype = DNA length = 23
FEATURE            Location/Qualifiers
misc_feature        1..23
                    note = Description of Artificial Sequence: Synthetic
                    oligonucleotide
source              1..23
                    mol_type = other DNA
                    organism = synthetic construct

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SEQUENCE: 189
ggtgagtgag tgtgtgcgtg tgg 23

SEQ ID NO: 190 moltype = DNA length = 23
FEATURE Location/Qualifiers
misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 190
tgtgggtgag tgtgtgcgtg agg 23

SEQ ID NO: 191 moltype = DNA length = 23
FEATURE Location/Qualifiers
misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 191
gagtccgagc agaagaagaa ggg 23

SEQ ID NO: 192 moltype = DNA length = 23
FEATURE Location/Qualifiers
misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 192
gagttagagc agaagaagaa agg 23

SEQ ID NO: 193 moltype = DNA length = 23
FEATURE Location/Qualifiers
misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 193
gaacctgagc tgctctgagc cgg 23

SEQ ID NO: 194 moltype = DNA length = 23
FEATURE Location/Qualifiers
misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 194
ttggcagggg gtgggagggg agg 23

SEQ ID NO: 195 moltype = DNA length = 23
FEATURE Location/Qualifiers
misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 195
gggagggaga gcttggcagg ggg 23

SEQ ID NO: 196 moltype = DNA length = 23
FEATURE Location/Qualifiers
misc_feature 1..23
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..23
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 196
gatggagcca gagaggatcc tgg 23

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FEATURE	Location/Qualifiers	
misc_feature	1..23	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..23	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 204		
gggaagggga gcttggcagg	tgg	23
SEQ ID NO: 205	moltype = DNA length = 23	
FEATURE	Location/Qualifiers	
misc_feature	1..23	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..23	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 205		
ggtagtgaga gcttggcagg	tgg	23
SEQ ID NO: 206	moltype = DNA length = 23	
FEATURE	Location/Qualifiers	
misc_feature	1..23	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..23	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 206		
ctccctccca ggatcctccc	agg	23
SEQ ID NO: 207	moltype = DNA length = 23	
FEATURE	Location/Qualifiers	
misc_feature	1..23	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..23	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 207		
gtccatggca ggatcctctc	agg	23
SEQ ID NO: 208	moltype = DNA length = 23	
FEATURE	Location/Qualifiers	
misc_feature	1..23	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..23	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 208		
ctcccccca gtatcctctc	agg	23
SEQ ID NO: 209	moltype = DNA length = 108	
FEATURE	Location/Qualifiers	
source	1..108	
	mol_type = unassigned DNA	
	organism = Homo sapiens	
SEQUENCE: 209		
ccaatctatg acatcaatta ttatacatg gagccctgcc aaaaaatcaa tgtgaagcaa	60	
atcgagccc gctcctgcc tccgtctac tcactggtgt tcattctt	108	
SEQ ID NO: 210	moltype = DNA length = 138	
FEATURE	Location/Qualifiers	
source	1..138	
	mol_type = unassigned DNA	
	organism = Homo sapiens	
SEQUENCE: 210		
ccaccctata attctgaacc tgcagaagaa tctgaacata aaaacaacaa ttacgaacca	60	
aacctattta aaactccaca aaggaaacca tcttataatc agctggcttc aactccaata	120	
atattcaaag agcaaggg	138	
SEQ ID NO: 211	moltype = DNA length = 49	
FEATURE	Location/Qualifiers	
source	1..49	
	mol_type = unassigned DNA	
	organism = Homo sapiens	
SEQUENCE: 211		

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ggccgggaat caagagtcac ccagagacag tgaccaacca tcctgttt      49

SEQ ID NO: 212      moltype = DNA length = 77
FEATURE            Location/Qualifiers
misc_feature       1..77
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..77
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 212
ggccgggaat caagagtcac ccagtgacca accatccctg taagcaaacc ttagaggttc 60
tggcaaggag agagatg      77

SEQ ID NO: 213      moltype = DNA length = 27
FEATURE            Location/Qualifiers
misc_feature       1..27
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..27
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 213
ggccgggaat caagagtcac ccaggaa      27

SEQ ID NO: 214      moltype = DNA length = 79
FEATURE            Location/Qualifiers
misc_feature       1..79
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..79
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 214
ggccgggaat caagagtcac ccagacctct ctggctccat cgtaagcaaa ccttagagggt 60
tctggcaagg agagagatg      79

SEQ ID NO: 215      moltype = DNA length = 28
FEATURE            Location/Qualifiers
misc_feature       1..28
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..28
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 215
ggccgggaat caagagtcac cctaacag      28

SEQ ID NO: 216      moltype = DNA length = 66
FEATURE            Location/Qualifiers
misc_feature       1..66
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..66
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 216
ggccgggaat caagacgctg gctccatcgt aagcaaacct tagaggttct ggcaaggaga 60
gagatg      66

SEQ ID NO: 217      moltype = DNA length = 47
FEATURE            Location/Qualifiers
misc_feature       1..47
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..47
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 217
ggctccatcg taagcaaacc ttagaggttc tggcaaggag agagatg      47

SEQ ID NO: 218      moltype = DNA length = 78
FEATURE            Location/Qualifiers
misc_feature       1..78
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..78
                   mol_type = other DNA
                   organism = synthetic construct

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SEQUENCE: 218
ggccgggaat caagagtcac ccagactctc tggctccatc gtaagcaaac cttagaggtt 60
ctggcaagga gagagatg 78

SEQ ID NO: 219 moltype = DNA length = 80
FEATURE Location/Qualifiers
misc_feature 1..80
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..80
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 219
ggccgggaat caagagtcac ccagagacag tgaccaacca tcgtaagcaa accttagagg 60
ttctggcaag gagagatg 80

SEQ ID NO: 220 moltype = DNA length = 46
FEATURE Location/Qualifiers
misc_feature 1..46
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..46
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 220
ggtccatcgt aagcaaacct tagaggttct ggcaaggaga gagatg 46

SEQ ID NO: 221 moltype = DNA length = 69
FEATURE Location/Qualifiers
misc_feature 1..69
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..69
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 221
ggccgggaat caagagtcac ccactccat cgtaagcaaa ccttagaggt tctggcaagg 60
agagatg 69

SEQ ID NO: 222 moltype = DNA length = 76
FEATURE Location/Qualifiers
misc_feature 1..76
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..76
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 222
ggccgggaat caagagtcac ccagactctg gtcctcgt aagcaaacct tagaggttct 60
ggcaaggaga gagatg 76

SEQ ID NO: 223 moltype = DNA length = 50
FEATURE Location/Qualifiers
misc_feature 1..50
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..50
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 223
ggccgggaat caagagtcac ccagagacag tgaccaacca tcccatatca 50

SEQ ID NO: 224 moltype = DNA length = 59
FEATURE Location/Qualifiers
misc_feature 1..59
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..59
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 224
ggccgggaat caagagtcac cgtaagcaaa ccttagaggt tctggcaagg agagatg 59

SEQ ID NO: 225 moltype = DNA length = 24
FEATURE Location/Qualifiers
misc_feature 1..24
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source 1..24

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mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 225
 aatgaccact acatccttca aggg 24

SEQ ID NO: 226 moltype = DNA length = 25
 FEATURE Location/Qualifiers
 misc_feature 1..25
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..25
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 226
 aatgaccact acatcctttc aaggg 25

SEQ ID NO: 227 moltype = DNA length = 26
 FEATURE Location/Qualifiers
 misc_feature 1..26
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..26
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 227
 aatgaccact acatcctttt caaggg 26

SEQ ID NO: 228 moltype = DNA length = 22
 FEATURE Location/Qualifiers
 misc_feature 1..22
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..22
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 228
 aatgaccact acatcctaag gg 22

SEQ ID NO: 229 moltype = DNA length = 21
 FEATURE Location/Qualifiers
 misc_feature 1..21
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..21
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 229
 aatgaccact acatcctagg g 21

SEQ ID NO: 230 moltype = DNA length = 20
 FEATURE Location/Qualifiers
 misc_feature 1..20
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 230
 aatgaccact acatcctggg 20

SEQ ID NO: 231 moltype = DNA length = 42
 FEATURE Location/Qualifiers
 source 1..42
 mol_type = unassigned DNA
 organism = Homo sapiens

SEQUENCE: 231
 tatgtgcaat gaccactaca tcctcaaggg cagcaatcgg ag 42

SEQ ID NO: 232 moltype = DNA length = 45
 FEATURE Location/Qualifiers
 misc_feature 1..45
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..45
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 232
 tatgtgcaat gaccactaca tcctcctcaa gggcagcaat cggag 45

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misc_feature 1..34
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..34
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 240
 tatgtgcaat gaccactaca ttggcagcaa tcgg 34
 SEQ ID NO: 241 moltype = DNA length = 21
 FEATURE Location/Qualifiers
 misc_feature 1..21
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..21
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 241
 tatgtgcaat gaccactaca t 21
 SEQ ID NO: 242 moltype = DNA length = 53
 FEATURE Location/Qualifiers
 source 1..53
 mol_type = unassigned DNA
 organism = Homo sapiens
 SEQUENCE: 242
 tcatacagat gatgtctcat catcagagga gcgagaaggt aaagtcaaaa tca 53
 SEQ ID NO: 243 moltype = DNA length = 32
 FEATURE Location/Qualifiers
 misc_feature 1..32
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..32
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 243
 tcatacagat gatacaggta aagtcaaaat ca 32
 SEQ ID NO: 244 moltype = DNA length = 32
 FEATURE Location/Qualifiers
 misc_feature 1..32
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..32
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 244
 tcatacaggt gatgaaggta aagtcaaaat ca 32
 SEQ ID NO: 245 moltype = DNA length = 50
 FEATURE Location/Qualifiers
 misc_feature 1..50
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..50
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 245
 tcatacagat gatgtctcat catcagagcg agaaggtaaa gtcaaaatca 50
 SEQ ID NO: 246 moltype = DNA length = 48
 FEATURE Location/Qualifiers
 misc_feature 1..48
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..48
 mol_type = other DNA
 organism = synthetic construct
 SEQUENCE: 246
 tcatacagat gatgtctcat catcagcgag aaggtaaagt caaaatca 48
 SEQ ID NO: 247 moltype = DNA length = 52
 FEATURE Location/Qualifiers
 misc_feature 1..52
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
 source 1..52
 mol_type = other DNA

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                organism = synthetic construct
SEQUENCE: 247
tcatacagat gatgtctcat catcaggag cgagaaggta aagtcaaaat ca          52

SEQ ID NO: 248      moltype = DNA length = 41
FEATURE            Location/Qualifiers
misc_feature       1..41
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..41
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 248
tcatacagat gatgtctcgc gagaaggtaa agtcaaaatc a                      41

SEQ ID NO: 249      moltype = DNA length = 32
FEATURE            Location/Qualifiers
misc_feature       1..32
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..32
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 249
tcatacagat gatgaaggta aagtcaaaat ca                                32

SEQ ID NO: 250      moltype = DNA length = 29
FEATURE            Location/Qualifiers
misc_feature       1..29
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..29
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 250
tcatacagat gaaggtaaag tcaaaatca                                    29

SEQ ID NO: 251      moltype = DNA length = 42
FEATURE            Location/Qualifiers
misc_feature       1..42
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..42
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 251
tcatacagat gatgtctaca gatgaaggta aagtcaaaat ca                    42

SEQ ID NO: 252      moltype = DNA length = 51
FEATURE            Location/Qualifiers
misc_feature       1..51
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..51
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 252
tcatacagat gatgtctcat catcaggagc gagaaggtaa agtcaaaatc a        51

SEQ ID NO: 253      moltype = DNA length = 34
FEATURE            Location/Qualifiers
misc_feature       1..34
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source             1..34
                   mol_type = other DNA
                   organism = synthetic construct

SEQUENCE: 253
gtcatcctca tcctgataaa ctgcaaaagg ctga                              34

SEQ ID NO: 254      moltype = DNA length = 606
FEATURE            Location/Qualifiers
source             1..606
                   mol_type = unassigned DNA
                   organism = Homo sapiens

SEQUENCE: 254
gctgggtgtct gggttctgtg ccccttcccc accccagccc accccagggtg tctgtccat 60
tctcagggtg gtcacatggg tggctctagg gtgtcccatg agagatgcaa agcgctgaa 120
ttttctgact cttcccatca gaccccccaa agacacatgt gaccaccacc cccatctctg 180

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accatgaggc caccctgagg tgtctggccc tgggcttcta ccctgcggag atcacactga 240
cctggcagcg ggatggcgag gaccaaactc aggacaccga gcttgaggag accagaccag 300
caggagatag aaccttccag aagtgggcag ctgtggtggt gccttctgga gaagagcaga 360
gatacacatg ccatgtacag catgaggggc tgccgaagcc cctcaccctg agatggggta 420
aggaggggga tgaggggtca tatctgttca tatctgttct cagggaaagc aggagccctt 480
ctggagccct tcagcagggg cagggcccct catcttcccc tcctttccca gagccatctt 540
cccagtccac catccccatc gtgggcattg ttgctggcct ggctgtccta gcagttgtgg 600
tcatcg 606

SEQ ID NO: 255      moltype = DNA length = 26
FEATURE           Location/Qualifiers
source            1..26
                  mol_type = unassigned DNA
                  organism = Homo sapiens

SEQUENCE: 255
actaccacag ctccttctct gagtgg 26

SEQ ID NO: 256      moltype = DNA length = 23
FEATURE           Location/Qualifiers
misc_feature      1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source            1..23
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 256
actaccacag ctcctctgag tgg 23

SEQ ID NO: 257      moltype = DNA length = 23
FEATURE           Location/Qualifiers
source            1..23
                  mol_type = unassigned DNA
                  organism = Homo sapiens

SEQUENCE: 257
gtagttggag ctggcggcgt agg 23

SEQ ID NO: 258      moltype = DNA length = 23
FEATURE           Location/Qualifiers
misc_feature      1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source            1..23
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 258
gtagttggag ctagcggcgt agg 23

SEQ ID NO: 259      moltype = DNA length = 23
FEATURE           Location/Qualifiers
source            1..23
                  mol_type = unassigned DNA
                  organism = Homo sapiens

SEQUENCE: 259
gtagttggag ctggtggcgt agg 23

SEQ ID NO: 260      moltype = DNA length = 23
FEATURE           Location/Qualifiers
misc_feature      1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source            1..23
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 260
gtagttggag ctagtggcgt agg 23

SEQ ID NO: 261      moltype = DNA length = 28
FEATURE           Location/Qualifiers
misc_feature      1..28
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source            1..28
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 261
ccatacatca aagatagtca tcttgggg 28

SEQ ID NO: 262      moltype = DNA length = 60
FEATURE           Location/Qualifiers

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source                1..60
                      mol_type = unassigned DNA
                      organism = Homo sapiens

SEQUENCE: 262
ccatacagtc agtatcaatt ctggaagaat ttccagacat taaagatagt catcttgggg 60

SEQ ID NO: 263        moltype = DNA length = 55
FEATURE              Location/Qualifiers
source               1..55
                      mol_type = unassigned DNA
                      organism = Homo sapiens

SEQUENCE: 263
ccatacagtc agtatcaatt ctggaagaat ttccagacat taaagatagt catct      55

SEQ ID NO: 264        moltype = DNA length = 23
FEATURE              Location/Qualifiers
misc_feature         1..23
                      note = Description of Artificial Sequence: Synthetic
                      oligonucleotide
source               1..23
                      mol_type = other DNA
                      organism = synthetic construct

SEQUENCE: 264
ccatacatTA aagatagtca tct                                          23

SEQ ID NO: 265        moltype = DNA length = 23
FEATURE              Location/Qualifiers
source               1..23
                      mol_type = unassigned DNA
                      organism = Homo sapiens

SEQUENCE: 265
agatgactat ctttaatgTC tgg                                          23

SEQ ID NO: 266        moltype = DNA length = 23
FEATURE              Location/Qualifiers
source               1..23
                      mol_type = unassigned DNA
                      organism = Homo sapiens

SEQUENCE: 266
agatgactat ctttaatgTA tgg                                          23

SEQ ID NO: 267        moltype = DNA length = 23
FEATURE              Location/Qualifiers
misc_feature         1..23
                      note = Description of Artificial Sequence: Synthetic
                      oligonucleotide
source               1..23
                      mol_type = other DNA
                      organism = synthetic construct

SEQUENCE: 267
gtagttggag ctgatggcgt agg                                          23

SEQ ID NO: 268        moltype = DNA length = 23
FEATURE              Location/Qualifiers
misc_feature         1..23
                      note = Description of Artificial Sequence: Synthetic
                      oligonucleotide
source               1..23
                      mol_type = other DNA
                      organism = synthetic construct

SEQUENCE: 268
gtagttggag ctggtagcgt agg                                          23

SEQ ID NO: 269        moltype = DNA length = 23
FEATURE              Location/Qualifiers
misc_feature         1..23
                      note = Description of Artificial Sequence: Synthetic
                      oligonucleotide
source               1..23
                      mol_type = other DNA
                      organism = synthetic construct

SEQUENCE: 269
gtagttggag ctggtgacgt agg                                          23

SEQ ID NO: 270        moltype = DNA length = 23
FEATURE              Location/Qualifiers
misc_feature         1..23
                      note = Description of Artificial Sequence: Synthetic
                      oligonucleotide

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source	1..23 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 270		
gtagttggag ctaatggcgt agg		23
SEQ ID NO: 271	moltype = DNA length = 23	
FEATURE	Location/Qualifiers	
misc_feature	1..23 note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..23 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 271		
gtagttggag ctagtagcgt agg		23
SEQ ID NO: 272	moltype = DNA length = 23	
FEATURE	Location/Qualifiers	
misc_feature	1..23 note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..23 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 272		
gtagttggag ctagtgacgt agg		23
SEQ ID NO: 273	moltype = DNA length = 23	
FEATURE	Location/Qualifiers	
source	1..23 mol_type = unassigned DNA organism = Homo sapiens	
SEQUENCE: 273		
caaatgaatg atgcacatca tgg		23
SEQ ID NO: 274	moltype = DNA length = 23	
FEATURE	Location/Qualifiers	
misc_feature	1..23 note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..23 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 274		
caaatgaatg atgcacgtca tgg		23
SEQ ID NO: 275	moltype = DNA length = 23	
FEATURE	Location/Qualifiers	
misc_feature	1..23 note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..23 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 275		
caaatgaatg atgcataatca tgg		23
SEQ ID NO: 276	moltype = DNA length = 23	
FEATURE	Location/Qualifiers	
misc_feature	1..23 note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..23 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 276		
caaatgaatg atgcgcatca tgg		23
SEQ ID NO: 277	moltype = DNA length = 23	
FEATURE	Location/Qualifiers	
misc_feature	1..23 note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..23 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 277		
caaatgaatg atgtacatca tgg		23

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FEATURE	Location/Qualifiers	
misc_feature	1..23	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..23	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 285		
caaatgaatg gtgcacgtca tgg		23
SEQ ID NO: 286	moltype = DNA length = 23	
FEATURE	Location/Qualifiers	
misc_feature	1..23	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..23	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 286		
caaatgagtg atgcacgtca tgg		23
SEQ ID NO: 287	moltype = DNA length = 23	
FEATURE	Location/Qualifiers	
misc_feature	1..23	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..23	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 287		
caaaagaatg atgcacgtca tgg		23
SEQ ID NO: 288	moltype = DNA length = 23	
FEATURE	Location/Qualifiers	
misc_feature	1..23	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..23	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 288		
cgaatgaatg atgcacgtca tgg		23
SEQ ID NO: 289	moltype = DNA length = 23	
FEATURE	Location/Qualifiers	
source	1..23	
	mol_type = unassigned DNA	
	organism = Homo sapiens	
SEQUENCE: 289		
atcataggtc gtcacgtta tgg		23
SEQ ID NO: 290	moltype = DNA length = 23	
FEATURE	Location/Qualifiers	
misc_feature	1..23	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..23	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 290		
atcataggtt gtcacgtta tgg		23
SEQ ID NO: 291	moltype = DNA length = 23	
FEATURE	Location/Qualifiers	
source	1..23	
	mol_type = unassigned DNA	
	organism = Homo sapiens	
SEQUENCE: 291		
atcataggtc gtcacgtta tgg		23
SEQ ID NO: 292	moltype = DNA length = 23	
FEATURE	Location/Qualifiers	
misc_feature	1..23	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..23	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 292		

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atcataggtt gtctgctta tgg	23
SEQ ID NO: 293	moltype = DNA length = 23
FEATURE	Location/Qualifiers
source	1..23
	mol_type = unassigned DNA
	organism = Homo sapiens
SEQUENCE: 293	
ctggacaaga agagtacagt gcc	23
SEQ ID NO: 294	moltype = DNA length = 23
FEATURE	Location/Qualifiers
misc_feature	1..23
	note = Description of Artificial Sequence: Synthetic oligonucleotide
source	1..23
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 294	
ctggaaaaga agagtacagt gcc	23
SEQ ID NO: 295	moltype = DNA length = 23
FEATURE	Location/Qualifiers
source	1..23
	mol_type = unassigned DNA
	organism = Homo sapiens
SEQUENCE: 295	
actccatcga gatttcactg tag	23
SEQ ID NO: 296	moltype = DNA length = 23
FEATURE	Location/Qualifiers
misc_feature	1..23
	note = Description of Artificial Sequence: Synthetic oligonucleotide
source	1..23
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 296	
actccatcga gatttctctg tag	23
SEQ ID NO: 297	moltype = DNA length = 34
FEATURE	Location/Qualifiers
misc_feature	1..34
	note = Description of Artificial Sequence: Synthetic oligonucleotide
source	1..34
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 297	
acttccaggc aacccaagg ggacctcatg cagg	34
SEQ ID NO: 298	moltype = DNA length = 35
FEATURE	Location/Qualifiers
misc_feature	1..35
	note = Description of Artificial Sequence: Synthetic oligonucleotide
source	1..35
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 298	
acttccaggc gaacccaag gggacctcat gcagg	35
SEQ ID NO: 299	moltype = DNA length = 13
FEATURE	Location/Qualifiers
misc_feature	1..13
	note = Description of Artificial Sequence: Synthetic oligonucleotide
source	1..13
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 299	
acttccaggc tcc	13
SEQ ID NO: 300	moltype = DNA length = 13
FEATURE	Location/Qualifiers
misc_feature	1..13
	note = Description of Artificial Sequence: Synthetic oligonucleotide
source	1..13

-continued

	mol_type = other DNA organism = synthetic construct	
SEQUENCE: 300		
acttccaggc tcc		13
SEQ ID NO: 301	moltype = DNA length = 32 Location/Qualifiers	
FEATURE	1..32	
misc_feature	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..32 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 301		
acttccaggc tcccaagggg acctcatgca gg		32
SEQ ID NO: 302	moltype = DNA length = 41 Location/Qualifiers	
FEATURE	1..41	
misc_feature	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..41 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 302		
acttccaggc tccttagggag gccaacccca aggggacctc a		41
SEQ ID NO: 303	moltype = DNA length = 25 Location/Qualifiers	
FEATURE	1..25	
misc_feature	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..25 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 303		
acttccaggc tccacctcat gcagg		25
SEQ ID NO: 304	moltype = DNA length = 32 Location/Qualifiers	
FEATURE	1..32	
misc_feature	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..32 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 304		
acttccaggc tccaccccca agggacctca tg		32
SEQ ID NO: 305	moltype = DNA length = 35 Location/Qualifiers	
FEATURE	1..35	
misc_feature	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..35 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 305		
acttccaggc tccacccaag gggacctcat gcagg		35
SEQ ID NO: 306	moltype = DNA length = 36 Location/Qualifiers	
FEATURE	1..36	
misc_feature	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..36 mol_type = other DNA organism = synthetic construct	
SEQUENCE: 306		
acttccaggc tccaccccaa ggggacctca tgcagg		36
SEQ ID NO: 307	moltype = DNA length = 38 Location/Qualifiers	
FEATURE	1..38	
misc_feature	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..38 mol_type = other DNA organism = synthetic construct	

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SEQUENCE: 307
acttccaggc tccacccacc caaggggacc tcatgcag 38

SEQ ID NO: 308 moltype = DNA length = 38
FEATURE Location/Qualifiers
misc_feature 1..38
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..38
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 308
acttccaggc tccacccacc ccaaggggac ctcatgca 38

SEQ ID NO: 309 moltype = DNA length = 40
FEATURE Location/Qualifiers
misc_feature 1..40
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..40
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 309
acttccaggc tccacccacc ccaaggggac ctcatgcagg 40

SEQ ID NO: 310 moltype = DNA length = 45
FEATURE Location/Qualifiers
misc_feature 1..45
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..45
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 310
acttccaggc tccacccggc gaaccccaag gggacctcat gcagg 45

SEQ ID NO: 311 moltype = DNA length = 34
FEATURE Location/Qualifiers
misc_feature 1..34
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..34
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 311
acttccaggc tccaccctgg ggacctcatg cagg 34

SEQ ID NO: 312 moltype = DNA length = 42
FEATURE Location/Qualifiers
misc_feature 1..42
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..42
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 312
acttccaggc tccacccgaa cccaagggg acctcatgca gg 42

SEQ ID NO: 313 moltype = DNA length = 31
FEATURE Location/Qualifiers
misc_feature 1..31
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..31
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 313
acttccaggc tccacccgaa cctcatgcag g 31

SEQ ID NO: 314 moltype = DNA length = 35
FEATURE Location/Qualifiers
misc_feature 1..35
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..35
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 314
acttccaggc tccacccgag gggacctcat gcagg 35

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SEQ ID NO: 315 moltype = DNA length = 36
FEATURE Location/Qualifiers
misc_feature 1..36
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..36
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 315
acttccaggc tccaccccaa ggggacctca tgcagg 36

SEQ ID NO: 316 moltype = DNA length = 42
FEATURE Location/Qualifiers
misc_feature 1..42
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..42
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 316
acttccaggc tccacccgaa cccaagggg acctcatgca gg 42

SEQ ID NO: 317 moltype = DNA length = 43
FEATURE Location/Qualifiers
misc_feature 1..43
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..43
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 317
acttccaggc tccacccgag accccaaggg gacctcatgc agg 43

SEQ ID NO: 318 moltype = DNA length = 48
FEATURE Location/Qualifiers
misc_feature 1..48
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..48
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 318
acttccaggc tccacccgag ggcgaacccc aaggggacct catgcagg 48

SEQ ID NO: 319 moltype = DNA length = 30
FEATURE Location/Qualifiers
misc_feature 1..30
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..30
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 319
acttccaggc tccacccgac ctcatgcagg 30

SEQ ID NO: 320 moltype = DNA length = 42
FEATURE Location/Qualifiers
misc_feature 1..42
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..42
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 320
acttccaggc tccacccgac cccaagggg acctcatgca gg 42

SEQ ID NO: 321 moltype = DNA length = 42
FEATURE Location/Qualifiers
misc_feature 1..42
 note = Description of Artificial Sequence: Synthetic
 oligonucleotide
source 1..42
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 321
acttccaggc tccacccgac gaagggcccc aaggggacct ca 42

SEQ ID NO: 322 moltype = DNA length = 45

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FEATURE	Location/Qualifiers	
misc_feature	1..45	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..45	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 322		
acttccaggc tccacccgac gaacccaag gggacctcat gcagg		45
SEQ ID NO: 323	moltype = DNA length = 48	
FEATURE	Location/Qualifiers	
misc_feature	1..48	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..48	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 323		
acttccaggc tccacccgac ggcgaacccc aaggggacct catgcagg		48
SEQ ID NO: 324	moltype = DNA length = 51	
FEATURE	Location/Qualifiers	
misc_feature	1..51	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..51	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 324		
acttccaggc tccacccgac gtgcttgagg gcgaaccca aggggacctc a		51
SEQ ID NO: 325	moltype = DNA length = 47	
FEATURE	Location/Qualifiers	
misc_feature	1..47	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..47	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 325		
acttccaggc tccacccgac tcactatctt ctgggctcct ccatgtc		47
SEQ ID NO: 326	moltype = DNA length = 49	
FEATURE	Location/Qualifiers	
misc_feature	1..49	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..49	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 326		
acttccaggc tccacccgac ttggcgaacc ccaaggggac ctcatgcag		49
SEQ ID NO: 327	moltype = DNA length = 51	
FEATURE	Location/Qualifiers	
misc_feature	1..51	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..51	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 327		
acttccaggc tccacccgac ttgcagggcg aacccaagg ggacctcatg c		51
SEQ ID NO: 328	moltype = DNA length = 53	
FEATURE	Location/Qualifiers	
misc_feature	1..53	
	note = Description of Artificial Sequence: Synthetic oligonucleotide	
source	1..53	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 328		
acttccaggc tccacccgac ttggagggcg aacccaagg ggacctcatg cag		53
SEQ ID NO: 329	moltype = DNA length = 53	
FEATURE	Location/Qualifiers	
misc_feature	1..53	

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note = Description of Artificial Sequence: Synthetic
  oligonucleotide
source      1..53
            mol_type = other DNA
            organism = synthetic construct
SEQUENCE: 329
acttccaggc tccaccgcac tttgagggc gaacccaag gggacctcat gca      53

SEQ ID NO: 330      moltype = DNA length = 53
FEATURE            Location/Qualifiers
misc_feature       1..53
note = Description of Artificial Sequence: Synthetic
  oligonucleotide
source      1..53
            mol_type = other DNA
            organism = synthetic construct
SEQUENCE: 330
acttccaggc tccaccgcac tgttgagggg cgaaccccaa ggggacctca tgc      53

SEQ ID NO: 331      moltype = DNA length = 502
FEATURE            Location/Qualifiers
misc_feature       1..502
note = Description of Artificial Sequence: Synthetic
  oligonucleotide
misc_difference    26..480
note = misc_feature - a, c, t, g, unknown or other
source      1..502
            mol_type = other DNA
            organism = synthetic construct
SEQUENCE: 331
acttccaggc tccaccgcac tggagnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 60
nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 120
nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 180
nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 240
nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 300
nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 360
nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 420
nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 480
ggcgaacccc aaggggacct cc      502

SEQ ID NO: 332      moltype = DNA length = 72
FEATURE            Location/Qualifiers
source      1..72
            mol_type = unassigned DNA
            organism = Homo sapiens
SEQUENCE: 332
agctctccct cccaggatcc tctctggctc catcgtaagc aaaccttaga gggtctggca 60
aggagagaga tg      72

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The invention claimed is:

1. A method of inducing a modification of a target endogenous nucleic acid sequence in a nucleus of a eukaryotic cell, comprising:
 - preparing a single-chain guide RNA (sgRNA), wherein the sgRNA comprises a crRNA and a tracrRNA;
 - preparing a Cas9 protein, wherein the Cas9 protein comprises a nuclear localization signal (NLS);
 - combining the sgRNA and the Cas9 protein in vitro to form a Cas9/sgRNA complex, wherein the sgRNA and the Cas9 protein are present in a molar ratio ranging from 29:14 to 29:1.4; and
 - introducing the Cas9/sgRNA complex into the eukaryotic cell, whereby the Cas9/sgRNA complex induces the modification of the target endogenous nucleic acid sequence in the nucleus of the eukaryotic cell.
2. The method of claim 1, wherein the Cas9 protein is a *Streptococcus pyogenes* Cas9 protein.
3. The method of claim 2, wherein the *Streptococcus pyogenes* Cas9 protein is a recombinant protein.
4. The method of claim 1, wherein the Cas9 protein was expressed in *E. coli*.
5. The method of claim 1, wherein the eukaryotic cell is a mammalian cell.
6. The method of claim 5, wherein the mammalian cell is a human cell.
7. The method of claim 1, wherein the Cas9/sgRNA complex is introduced into the eukaryotic cell by a method selected from the group consisting of electroporation, DEAE-dextran treatment, lipofection, nanoparticle-mediated transfection, and protein transduction domain mediated transduction.
8. The method of claim 1, wherein the Cas9/sgRNA complex is introduced into the eukaryotic cell by transfection.
9. The method of claim 1, wherein the Cas9/sgRNA complex is introduced into the eukaryotic cell by electroporation.
10. The method of claim 1, wherein the target endogenous nucleic acid comprises a trinucleotide protospacer adjacent motif (PAM) recognized by the Cas9 protein, wherein the PAM consists of trinucleotide 5'-NGG-3'.
11. The method of claim 1, wherein the NLS is disposed at the C-terminus of the Cas9 protein.
12. The method of claim 1, wherein the modification includes any one of a deletion, insertion, or substitution of at least one nucleotide.

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13. The method of claim 1, wherein the method further comprises allowing the eukaryotic cell to divide into a plurality of cells, each of which comprises the modification to the target endogenous nucleic acid sequence.

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